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(71) Applicants (for all designated States except US): **STATE OF VICTORIA AS REPRESENTED BY DEPARTMENT OF NATURAL RESOURCES AND ENVIRONMENT [AU/AU]**; 15th Floor, 8 Nicholson Street, East Melbourne, Victoria 3002 (AU). **THE UNIVERSITY OF ADELAIDE [AU/AU]**; North Terrace, Adelaide, South Australia 5005 (AU). **INTERNATIONAL MAIZE**

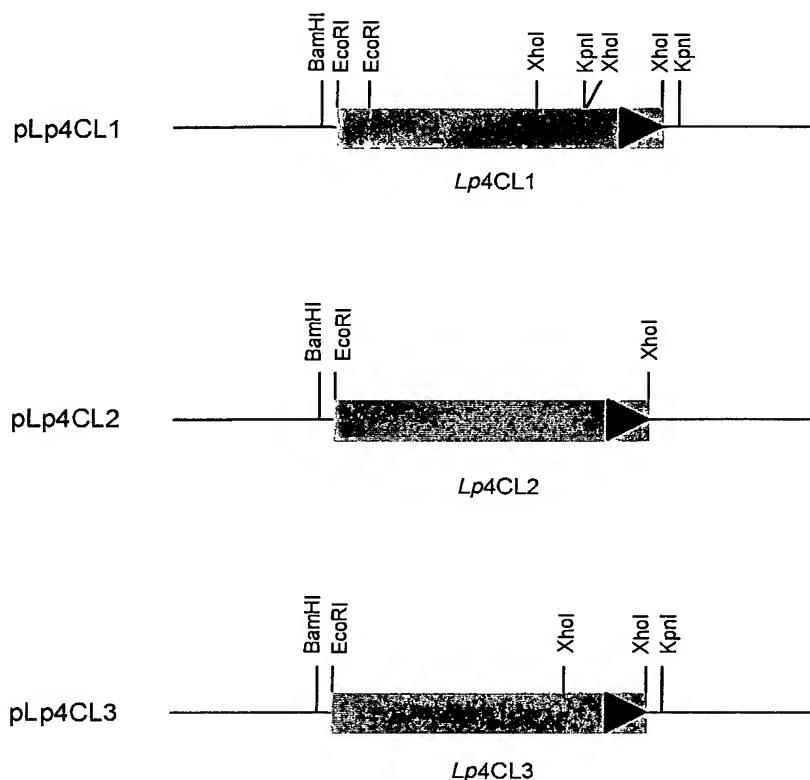
AND WHEAT IMPROVEMENT CENTER [MX/MX]; Lisboa 27, Apartado Postal 6-641, Mexico, D.F. 06600 (MX). **STATE OF SOUTH AUSTRALIA** as represented by **SOUTH AUSTRALIAN RESEARCH AND DEVELOPMENT INSTITUTE [AU/AU]**; Waite Road, Glen Osmond, South Australia 5064 (AU). **SOUTHERN CROSS UNIVERSITY [AU/AU]**; Military Road, Lismore, New South Wales 2580 (AU). **DAIRY RESEARCH AND DEVELOPMENT CORPORATION [AU/AU]**; Level 3, 84 William Street, Melbourne, Victoria 3000 (AU).

(72) Inventors; and

(75) Inventors/Applicants (for US only): **SPANGENBERG, German, Carlos** [UY/AU]; 56 Arthur Street, Bundoora, Victoria 3083 (AU). **LIDGETT, Angela, Jane** [AU/AU]; 13 Moore Street, Richmond, Victoria 3121 (AU). **HEATH, Robyn, Louise** [AU/AU]; 3 Berry Street, Clifton Hill, Victoria 3068 (AU). **MCINNES, Russell, Leigh** [AU/AU]; Glenn College, La Trobe University, Bundoora, Victoria 3083 (AU). **LYNCH, Damian, Paul** [NZ/AU]; Unit 10, 141 Elm Street, Northcote, Victoria 3070 (AU).

[Continued on next page]

(54) Title: MODIFICATION OF LIGNIN BIOSYNTHESIS



(57) Abstract: The present invention relates to the modification of lignin biosynthesis in plants, using the nucleotide sequences encoding the enzymes 4-coumarate CoA-ligase (4CL), cinnamoyl-CoA reductase (CCR), cinnamyl alcohol dehydrogenase (CAD) of the lignin biosynthetic pathway, from ryegrass (*Lolium*) and fescue (*Festuca*). The present invention also relates to regulatory elements, promoters capable of causing expression of exogenous genes in plants, wherein the regulatory elements are from the genes for caffeic acid Omethyl transferase (OMT), 4CL, CCR or CAD. The invention also relates to vectors including the nucleic acids and regulatory elements of the invention, plant cells, plants, plant seeds and other plant parts transformed with the regulatory elements, nucleic acids and vectors and methods using the nucleic acids, regulatory elements and vectors.

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(74) **Agent:** FREEHILLS CARTER SMITH BEADLE;
Level 43, 101 Collins Street, Melbourne, Victoria 3000
(AU).

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MODIFICATION OF LIGNIN BIOSYNTHESIS

The present invention relates to the modification of lignin biosynthesis in plants and, more particularly, to enzymes involved in the lignin biosynthetic pathway and nucleic acids encoding such enzymes.

5 The present invention also relates to a regulatory element and, more particularly, to a promoter capable of causing expression of an exogenous gene in plant cells, such as a gene encoding an enzyme involved in the lignin biosynthetic pathway in plants.

10 The invention also relates to vectors including the nucleic acids and regulatory elements of the invention, plant cells, plants, seeds and other plant parts transformed with the regulatory elements, nucleic acids and vectors, and methods of using the nucleic acids, regulatory elements and vectors.

15 Lignins are complex phenolic polymers that strengthen plant cell walls against mechanical and chemical degradation. The process of lignification typically occurs during secondary thickening of the walls of cells with structural, conductive or defensive roles. Three monolignol precursors, sinapyl, coniferyl and *p*-coumaryl alcohol combine by dehydrogenative polymerisation to produce respectively the syringyl(S), guaiacyl(G) and hydroxyl(H) subunits of the lignin polymer, which can also become linked to cell-wall polysaccharides 20 through the action of peroxidases and other oxidative isozymes. In grasses, biosynthesis of the monolignol precursors is a multistep process beginning with the aromatic amino-acids phenylalanine and tyrosine. It is the final two reduction/ dehydrogenation steps of the pathway, catalysed by Cinnamoyl CoA Reductase (CCR) and Cinnamyl Alcohol Dehydrogenase (CAD) that are 25 considered to be specific to lignin biosynthesis. The proportions of monolignols incorporated into the lignin polymer vary depending on plant species, tissue, developmental stage and sub-cellular location.

Caffeic acid O-methyl transferase (OMT), 4 coumarate CoA-ligase (4CL), cinnamoyl-CoA reductase (CCR) and cinnamyl alcohol dehydrogenase

- 2 -

(CAD) are key enzymes involved in lignin biosynthesis.

Worldwide permanent pasture is estimated to cover 70% of agriculturally cultivated area. Ryegrasses (*Lolium* spp.) together with the closely related fescues (*Festuca* spp.) are of significant value in temperate grasslands. The commercially most important ryegrasses are Italian or annual ryegrass (*L. multiflorum* Lam.) and perennial ryegrass (*L. perenne* L.). They are the key forage species in countries where livestock production is an intensive enterprise, such as the Netherlands, United Kingdom and New Zealand. The commercially most important fescues are tall fescue (*F. anundinacea* Schreb.), meadow fescue (*F. pratensis*) and red fescue (*F. rubra*).

Perennial ryegrass (*Lolium perenne* L.) is the major grass species sown in temperate dairy pastures in Australia, and the key pasture grass in temperate climates throughout the world. A marked decline of the feeding value of grasses is observed in temperate pastures of Australia during late spring and early summer, where the nutritive value of perennial ryegrass based pasture is often insufficient to meet the metabolic demands of lactating dairy cattle. Perennial ryegrass is also an important turf grass.

Grass and legume *in vitro* dry matter digestibility has been negatively correlated with lignin content. In addition, natural mutants of lignin biosynthetic enzymes in maize, sorghum and pearl millet that have higher rumen digestibility have been characterised as having lower lignin content and altered S/G subunit ratio. Thus, lignification of plant cell walls is the major factor identified as responsible for lowering digestibility of forage tissues as they mature.

It would be desirable to have methods of altering lignin biosynthesis in plants, including grass species such as ryegrasses and fescues, by reducing the activity of key biosynthetic enzymes in order to reduce lignin content and/or alter lignin composition for enhancing dry matter digestibility and improving herbage quality. However, for some applications it may be desirable to

- 3 -

enhance lignin biosynthesis to increase lignin content and/or alter lignin composition, for example to increase mechanical strength of wood, to increase mechanical strength of turf grasses, to reduce plant height and reduce lodging or improve disease resistance.

5 While nucleic acid sequences encoding some of the enzymes involved in the lignin biosynthetic pathway have been isolated for certain species of plants, there remains a need for materials useful in the modification of lignin biosynthesis in plants, particularly grass species such as ryegrasses and fescues.

10 Other phenotypic traits which may be improved by transgenic manipulation of plants include disease resistance, mineral content, nutrient quality and drought tolerance.

15 However, transgenic manipulation of phenotypic traits in plants requires the availability of regulatory elements capable of causing the expression of exogenous genes in plant cells.

It is an object of the present invention to overcome, or at least alleviate, one or more of the difficulties or deficiencies associated with the prior art.

20 In one aspect, the present invention provides substantially purified or isolated nucleic acids or nucleic acid fragments encoding the following enzymes from a ryegrass (*Lolium*) or fescue (*Festuca*) species: 4 coumarate CoA-ligase (4CL), cinnamoyl-CoA reductase (CCR) and cinnamyl alcohol dehydrogenase (CAD).

25 The ryegrass (*Lolium*) or fescue (*Festuca*) species may be of any suitable type, including Italian or annual ryegrass, perennial ryegrass, tall fescue, meadow fescue and red fescue. Preferably the ryegrass or fescue species is a ryegrass, more preferably perennial ryegrass (*Lolium perenne*).

The nucleic acid or nucleic acid fragment may be of any suitable type

- 4 -

and includes DNA (such as cDNA or genomic DNA) and RNA (such as mRNA) that is single- or double- stranded, optionally containing synthetic, non-natural or altered nucleotide bases, and combinations thereof.

The term "isolated" means that the material is removed from its original environment (eg. the natural environment if it is naturally occurring). For example, a naturally occurring nucleic acid present in a living plant is not isolated, but the same nucleic acid separated from some or all of the coexisting materials in the natural system, is isolated. Such nucleic acids could be part of a vector and/or such nucleic acids could be part of a composition, and still be isolated in that such a vector or composition is not part of its natural environment.

In a preferred embodiment of this aspect of the invention, the substantially purified or isolated nucleic acid or nucleic acid fragment encoding 4CL includes a nucleotide sequence selected from the group consisting of (a) sequences shown in Figures 2, 3 and 4 hereto (Sequence ID Nos: 1, 3 and 5; respectively) (b) complements of the sequences shown in Figures 2, 3 and 4 hereto (Sequence ID Nos: 1, 3 and 5, respectively); (c) sequences antisense to the sequences recited in (a) and (b); and (d) functionally active fragments and variants of the sequences recited in (a), (b) and (c).

In a further preferred embodiment of this aspect of the invention, the substantially purified or isolated nucleic acid or nucleic acid fragment encoding CCR includes a nucleotide sequence selected from the group consisting of (a) the sequence shown in Figure 10 hereto (Sequence ID No: 7); (b) the complement of the sequence shown in Figure 10 hereto (Sequence ID No: 7); (c) sequences antisense to the sequences recited in (a) and (b); and (d) functionally active fragments and variants of the sequences recited in (a), (b) and (c).

In a still further preferred embodiment of this aspect of the invention, the substantially purified or isolated nucleic acid or nucleic acid fragment encoding CAD includes a nucleotide sequence selected from the group consisting of (a)

- 5 -

the sequences shown in Figures 13, 14, 26 and 27 hereto (Sequence ID Nos: 9, 11, 14 and 16, respectively); (b) complements of the sequences shown in Figures 13, 14, 26 and 27 hereto (Sequence ID Nos: 9, 11, 14 and 16, respectively); (c) sequences antisense to the sequences recited in (a) and (b);
5 and (d) functionally active fragments and variants of the sequences recited in (a), (b) and (c).

By "functionally active" is meant that the fragment or variant (such as an analogue, derivative or mutant) is capable of modifying lignin biosynthesis in a plant. Such variants include naturally occurring allelic variants and non-
10 naturally occurring variants. Additions, deletions, substitutions and derivatizations of one or more of the nucleotides are contemplated so long as the modifications do not result in loss of functional activity of the fragment or variant. Preferably the functionally active fragment or variant has at least approximately 80% identity to the relevant part of the above mentioned
15 sequence, more preferably at least approximately 90% identity, most preferably at least approximately 95% identity. Such functionally active variants and fragments include, for example, those having nucleic acid changes which result in conservative amino acid substitutions of one or more residues in the corresponding amino acid sequence. Preferably the fragment
20 has a size of at least 10 nucleotides, more preferably at least 15 nucleotides, most preferably at least 20 nucleotides.

In a second aspect of the present invention there is provided a vector including a nucleic acid or nucleic acid fragment according to the present invention.

25 In a preferred embodiment of this aspect of the invention, the vector may include a regulatory element such as a promoter, a nucleic acid or nucleic acid fragment according to the present invention and a terminator; said regulatory element, nucleic acid or nucleic acid fragment and terminator being operatively linked.

30 By "operatively linked" is meant that said regulatory element is capable

- 6 -

of causing expression of said nucleic acid or nucleic acid fragment in a plant cell and said terminator is capable of terminating expression of said nucleic acid or nucleic acid fragment in a plant cell. Preferably, said regulatory element is upstream of said nucleic acid or nucleic acid fragment and said terminator is
5 downstream of said nucleic acid or nucleic acid fragment.

The vector may be of any suitable type and may be viral or non-viral. The vector may be an expression vector. Such vectors include chromosomal, non-chromosomal and synthetic nucleic acid sequences, eg. derivatives of plant viruses; bacterial plasmids; derivatives of the Ti plasmid from
10 *Agrobacterium tumefaciens*; derivatives of the Ri plasmid from *Agrobacterium rhizogenes*; phage DNA; yeast artificial chromosomes; bacterial artificial chromosomes; binary bacterial artificial chromosomes; vectors derived from combinations of plasmids and phage DNA. However, any other vector may be used as long as it is replicable or integrative or viable in the plant cell.

15 The regulatory element and terminator may be of any suitable type and may be endogenous to the target plant cell or may be exogenous, provided that they are functional in the target plant cell.

Preferably the regulatory element is a promoter. A variety of promoters which may be employed in the vectors of the present invention are well known
20 to those skilled in the art. Factors influencing the choice of promoter include the desired tissue specificity of the vector, and whether constitutive or inducible expression is desired and the nature of the plant cell to be transformed (eg. monocotyledon or dicotyledon). Particularly suitable promoters include the Cauliflower Mosaic Virus 35S (CaMV 35S) promoter, the
25 maize Ubiquitin promoter, the rice Actin promoter, and ryegrass endogenous OMT, 4CL, CCR or CAD promoters.

A variety of terminators which may be employed in the vectors of the present invention are also well known to those skilled in the art. The terminator may be from the same gene as the promoter sequence or a different gene.
30 Particularly suitable terminators are polyadenylation signals, such as the

- 7 -

CaMV 35S polyA and other terminators from the nopaline synthase (*nos*) and the octopine synthase (*ocs*) genes.

The vector, in addition to the regulatory element, the nucleic acid or nucleic acid fragment of the present invention and the terminator, may include
5 further elements necessary for expression of the nucleic acid or nucleic acid fragment, in different combinations, for example vector backbone, origin of replication (ori), multiple cloning sites, spacer sequences, enhancers, introns (such as the maize Ubiquitin Ubi intron), antibiotic resistance genes and other selectable marker genes [such as the neomycin phosphotransferase (*npt2*) gene, the hygromycin phosphotransferase (*hph*) gene, the phosphinothricin acetyltransferase (*bar* or *paf*) gene], and reporter genes (such as beta-glucuronidase (GUS) gene (*gusA*]). The vector may also contain a ribosome binding site for translation initiation. The vector may also include appropriate sequences for amplifying expression.

15 As an alternative to use of a selectable marker gene to provide a phenotypic trait for selection of transformed host cells, the presence of the vector in transformed cells may be determined by other techniques well known in the art, such as PCR (polymerase chain reaction), Southern blot hybridisation analysis, histochemical GUS assays, northern and Western blot 20 hybridisation analyses.

Those skilled in the art will appreciate that the various components of the vector are operatively linked, so as to result in expression of said nucleic acid or nucleic acid fragment. Techniques for operatively linking the components of the vector of the present invention are well known to those skilled in the art. Such techniques include the use of linkers, such as synthetic linkers, for example including one or more restriction enzyme sites.

30 The vectors of the present invention may be incorporated into a variety of plants, including monocotyledons (such as grasses from the genera *Lolium*, *Festuca*, *Paspalum*, *Pennisetum*, *Panicum* and other forage and turf grasses, corn, oat, sugarcane, wheat and barley), dicotyledons (such as *arabidopsis*,

- 8 -

tobacco, legumes, alfalfa, oak, eucalyptus, maple, canola, soybean and chickpea) and gymnosperms. In a preferred embodiment, the vectors are used to transform monocotyledons, preferably grass species such as ryegrasses (*Lolium* species) and fescues (*Festuca* species), more preferably 5 perennial ryegrass (*Lolium perenne*) including forage and turf type cultivars.

Techniques for incorporating the vectors of the present invention into plant cells (for example by transduction, transfection or transformation) are well known to those skilled in the art. Such techniques include *Agrobacterium* mediated introduction, electroporation to tissues, cells and protoplasts, 10 protoplast fusion, injection into reproductive organs, injection into immature embryos and high velocity projectile introduction to cells, tissues, calli, immature and mature embryos. The choice of technique will depend largely on the type of plant to be transformed.

Cells incorporating the vector of the present invention may be selected, 15 as described above, and then cultured in an appropriate medium to regenerate transformed plants, using techniques well known in the art. The culture conditions, such as temperature, pH and the like, will be apparent to the person skilled in the art. The resulting plants may be reproduced, either sexually or asexually, using methods well known in the art, to produce 20 successive generations of transformed plants.

In a further aspect of the present invention there is provided a plant cell, plant, plant seed or other plant part, including, eg transformed with, a vector of the present invention.

The plant cell, plant, plant seed or other plant part may be from any 25 suitable species, including monocotyledons, dicotyledons and gymnosperms. In a preferred embodiment the plant cell, plant, plant seed or other plant part may be from a monocotyledon, preferably a grass species, more preferably a ryegrass (*Lolium* species) or fescue (*Festuca* species), even more preferably a ryegrass, most preferably perennial ryegrass, including forage- and turf-type 30 cultivars.

- 9 -

The present invention also provides a plant, plant seed or other plant part derived from a plant cell of the present invention.

The present invention also provides a plant, plant seed or other plant part derived from a plant of the present invention.

5 In a further aspect of the present invention there is provided a method of modifying lignin biosynthesis in a plant, said method including introducing into said plant an effective amount of a nucleic acid or nucleic acid fragment and/or a vector according to the present invention.

By "an effective amount" is meant an amount sufficient to result in an
10 identifiable phenotypic trait in said plant, or a plant, plant seed or other plant part derived therefrom. Such amounts can be readily determined by an appropriately skilled person, taking into account the type of plant, the route of administration and other relevant factors. Such a person will readily be able to determine a suitable amount and method of administration. See, for example,
15 Maniatis et al, Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory, Cold Spring Harbor, the entire disclosure of which is incorporated herein by reference.

Using the methods and materials of the present invention, plant lignin biosynthesis may be increased, decreased or otherwise modified relative to an
20 untransformed control plant. It may be increased or otherwise modified, for example, by incorporating additional copies of a sense nucleic acid or nucleic acid fragment of the present invention. It may be decreased, for example, by incorporating an antisense nucleic acid or nucleic acid fragment of the present invention. In addition, the number of copies of genes encoding for different
25 enzymes in the lignin biosynthetic pathway may be manipulated to modify the relative amount of each monolignol synthesized, thereby leading to the formation of lignin having altered composition.

In a still further aspect of the present invention there is provided use of a nucleic acid or nucleic acid fragment according to the present invention, and/or

- 10 -

nucleotide sequence information thereof, and/or single nucleotide polymorphisms thereof, as a molecular genetic marker.

More particularly, nucleic acids or nucleic acid fragments according to the present invention, and/or nucleotide sequence information thereof, and/or 5 single nucleotide polymorphisms thereof, may be used as a molecular genetic marker for qualitative trait loci (QTL) tagging, mapping, DNA fingerprinting and in marker assisted selection, and may be used as candidate genes or perfect markers, particularly in ryegrasses and fescues. Even more particularly, 10 nucleic acids or nucleic acid fragments according to the present invention, and/or nucleotide sequence information thereof, may be used as molecular genetic markers in forage and turf grass improvement, eg. tagging QTLs for dry matter digestibility, herbage quality, mechanical stress tolerance, disease resistance, insect pest resistance, plant stature and leaf and stem colour.

In a still further aspect of the present invention there is provided a 15 substantially purified or isolated polypeptide from a ryegrass (*Lolium*) or fescue (*Festuca*) species, selected from the group consisting of the enzymes 4CL, CCR and CAD.

The ryegrass (*Lolium*) or fescue (*Festuca*) species may be of any suitable type, including Italian or annual ryegrass, perennial ryegrass, tall 20 fescue, meadow fescue and red fescue. Preferably the species is a ryegrass, more preferably perennial ryegrass (*L. perenne*).

In a preferred embodiment of this aspect of the invention, the substantially purified or isolated enzyme 4CL includes an amino acid sequence selected from the group consisting of sequences shown in Figures 2, 3 and 4 25 hereto (Sequence ID Nos: 2, 4 and 6, respectively); and functionally active fragments and variants thereof.

In a further preferred embodiment of this aspect of the invention, the substantially purified or isolated enzyme CCR includes an amino acid sequence selected from the group consisting of the sequence shown in Figure

- 11 -

10 hereto (Sequence ID No: 8); and functionally active fragments and variants thereof.

In a still further preferred embodiment of this aspect of the invention, the substantially purified or isolated enzyme CAD includes an amino acid sequence selected from the group consisting of the sequence shown in Figures 13, 14, 26 and 27 hereto (Sequence ID Nos: 10, 12, 15 and 17, respectively); and functionally active fragments and variants thereof.

By "functionally active" in this context is meant that the fragment or variant has one or more of the biological properties of the enzymes 4CL, CCR and CAD, respectively. Additions, deletions, substitutions and derivatizations of one or more of the amino acids are contemplated so long as the modifications do not result in loss of functional activity of the fragment or variant. Preferably the fragment or variant has at least approximately 60% identity to the relevant part of the above mentioned sequence, more preferably at least approximately 80% identity, most preferably at least approximately 90% identity. Such functionally active variants and fragments include, for example, those having conservative amino acid substitutions of one or more residues in the corresponding amino acid sequence. Preferably the fragment has a size of at least 10 amino acids, more preferably at least 15 amino acids, most preferably at least 20 amino acids.

In a further embodiment of this aspect of the invention, there is provided a polypeptide recombinantly produced from a nucleic acid or nucleic acid fragment according to the present invention. Techniques for recombinantly producing polypeptides are well known to those skilled in the art.

25 In a still further aspect of the present invention there is provided a lignin or modified lignin substantially or partially purified or isolated from a plant, plant seed or other plant part of the present invention.

Such lignins may be modified from naturally occurring lignins in terms of the length, the degree of polymerisation (number of units), degree of branching

- 12 -

and/or nature of linkages between units.

In a still further aspect, the present invention provides an isolated regulatory element capable of causing expression of an exogenous gene in plant cells. Preferably the regulatory element is isolated from a nucleic acid or
5 nucleic acid fragment encoding OMT, 4CL, CCR or CAD.

The regulatory element may be a nucleic acid molecule, including DNA (such as cDNA or genomic DNA) and RNA (such as mRNA) that is single- or double- stranded, optionally containing synthetic, non-natural or altered nucleotide bases, and combinations thereof.

10 Preferably the regulatory element includes a promoter, more preferably an *O*-methyltransferase promoter, even more preferably an *O*-methyltransferase promoter from a ryegrass (*Lolium*) or fescue (*Festuca*) species, more preferably a ryegrass, most preferably perennial ryegrass (*Lolium perenne*).

15 In a particularly preferred embodiment of this aspect of the invention, the regulatory element includes a promoter from the caffeic acid *O*-methyltransferase gene corresponding to the cDNA homologue *LpOMT1* from perennial ryegrass.

20 Preferably the regulatory element includes a nucleotide sequence including the first approximately 4630 nucleotides of the sequence shown in Figure 18 hereto (Sequence ID No: 13); or a functionally active fragment or variant thereof.

25 By "functionally active" in this context is meant that the fragment or variant (such as an analogue, derivative or mutant) is capable of causing expression of a transgene in plant cells. Such variants include naturally occurring allelic variants and non-naturally occurring variants. Additions, deletions, substitutions and derivatizations of one or more of the nucleotides are contemplated so long as the modifications do not result in loss of functional

- 13 -

activity of the regulatory element. Preferably the functionally active fragment or variant has at least approximately 80% identity to the relevant part of the above sequence, more preferably at least approximately 90% identity, most preferably at least approximately 95% identity. Preferably the fragment has a
5 size of at least 100 nucleotides, more preferably at least 150 nucleotides, most preferably at least 200 nucleotides.

In a particularly preferred embodiment of this aspect of the invention, the regulatory element includes a nucleotide sequence selected from the group consisting of:

- 10 Nucleotides – 4581 to –1
- Nucleotides -4285 to –1
- Nucleotides –4020 to –1
- Nucleotides –2754 to –1
- Nucleotides – 1810 to –1
- 15 Nucleotides –831 to –1
- Nucleotides –560 to –1
- Nucleotides –525 to –1
- Nucleotides –274 to –1
- Nucleotides –21 to -1
- 20 of Figure 18 hereto (Sequence ID No: 13);

or a functionally active fragment or variant thereof.

In another preferred embodiment the regulatory element includes a 4 coumarate-CoA ligase promoter, even more preferably a 4 coumarate-CoA ligase promoter from a ryegrass (*Lolium*) or fescue (*Festuca*) species, more
25 preferably a ryegrass, most preferably perennial ryegrass (*Lolium perenne*).

- 14 -

In a particularly preferred embodiment of this aspect of the invention, the regulatory element includes a promoter from the 4 coumarate-CoA ligase gene corresponding to the cDNA homologue *Lp4CL2* from perennial ryegrass.

Preferably the regulatory element includes a nucleotide sequence 5 including the first approximately 2206 nucleotides of the sequence shown in Figure 38 hereto (Sequence ID No: 17); or a functionally active fragment or variant thereof.

By "functionally active" in this context is meant that the fragment or variant (such as an analogue, derivative or mutant) is capable of causing 10 expression of a transgene in plant cells. Such variants include naturally occurring allelic variants and non-naturally occurring variants. Additions, deletions, substitutions and derivatizations of one or more of the nucleotides are contemplated so long as the modifications do not result in loss of functional activity of the regulatory element. Preferably the functionally active fragment or 15 variant has at least approximately 80% identity to the relevant part of the above sequence, more preferably at least approximately 90% identity, most preferably at least approximately 95% identity. Preferably the fragment has a size of at least 100 nucleotides, more preferably at least 150 nucleotides, most preferably at least 200 nucleotides.

20 In a particularly preferred embodiment of this aspect of the invention, the regulatory element includes a nucleotide sequence selected from the group consisting of:

Nucleotides – 2206 to –1

Nucleotides -1546 to –1

25 Nucleotides –1186 to –1

Nucleotides –406 to –1

Nucleotides – 166 to –1

- 15 -

of Figure 38 hereto (Sequence ID No: 17);

or a functionally active fragment or variant thereof.

In another preferred embodiment the regulatory element includes a cinnamoyl-CoA reductase promoter, even more preferably a cinnamoyl-CoA reductase promoter from a ryegrass (*Lolium*) or fescue (*Festuca*) species, 5 more preferably a ryegrass, most preferably perennial ryegrass (*Lolium perenne*).

In a particularly preferred embodiment of this aspect of the invention, the regulatory element includes a promoter from the cinnamoyl-CoA reductase 10 gene corresponding to the *LpCCR1* cDNA from perennial ryegrass.

Preferably the regulatory element includes a nucleotide sequence including the first approximately 6735 nucleotides of the sequence shown in Figure 39 hereto (Sequence ID No: 18); or a functionally active fragment or variant thereof.

15 By "functionally active" in this context is meant that the fragment or variant (such as an analogue, derivative or mutant) is capable of causing expression of a transgene in plant cells. Such variants include naturally occurring allelic variants and non-naturally occurring variants. Additions, deletions, substitutions and derivatizations of one or more of the nucleotides 20 are contemplated so long as the modifications do not result in loss of functional activity of the regulatory element. Preferably the functionally active fragment or variant has at least approximately 80% identity to the relevant part of the above sequence, more preferably at least approximately 90% identity, most preferably at least approximately 95% identity. Preferably the fragment has a 25 size of at least 100 nucleotides, more preferably at least 150 nucleotides, most preferably at least 200 nucleotides.

- 16 -

In a particularly preferred embodiment of this aspect of the invention, the regulatory element includes a nucleotide sequence selected from the group consisting of:

- Nucleotides - 6735 to -1
- 5 Nucleotides -5955 to -1
- Nucleotides -5415 to -1
- Nucleotides -4455 to -1
- Nucleotides - 4035 to -1
- Nucleotides -3195 to -1
- 10 Nucleotides -2595 to -1
- Nucleotides -1755 to -1
- Nucleotides -1275 to -1
- Nucleotides -495 to -1
- Nucleotides -255 to -1
- 15 Nucleotides -75 to -1

of Figure 39 hereto (Sequence ID No: 18);

or a functionally active fragment or variant thereof.

By an "exogenous gene" is meant a gene not natively linked to said regulatory element. In certain embodiments of the present invention the 20 exogenous gene is also not natively found in the relevant plant or plant cell.

The exogenous gene may be of any suitable type. The exogenous gene may be a nucleic acid such as DNA (e.g. cDNA or genomic DNA) or RNA (e.g. mRNA), and combinations thereof. The exogenous gene may correspond to a target gene, for example a gene capable of influencing disease resistance, 25 herbage digestibility, nutrient quality, mineral content or drought tolerance or be a fragment or variant (such as an analogue, derivative or mutant) thereof

- 17 -

which is capable of modifying expression of said target gene. Such variants include nucleic acid sequences which are antisense to said target gene or an analogue, derivative, mutant or fragment thereof. The transgene may code for a protein or RNA sequence depending the target condition and whether down 5 or up-regulation of gene expression is required. Preferably, the target gene is selected from exogenous coding sequences coding for mRNA for a protein, this protein may be of bacterial origin (such as enzymes involved in cell wall modification and cell wall metabolism, cytokinin biosynthesis), or eukaryotic origin (such as pharmaceutically active polypeptides) or of plant origin (such as 10 enzymes involved in the synthesis of phenolic compounds, cell wall metabolism, sugar metabolism, lignin biosynthesis). Preferably, the target gene is selected from the group comprising *O*-methyltransferase, 4 coumarate CoA-ligase, cinnamoyl CoA reductase, cinnamyl alcohol dehydrogenase, cinnamate 4 hydroxylase, phenolase, laccase, peroxidase, coniferol glucosyl 15 transferase, coniferin beta-glucosidase, phenylalanine ammonia lyase, ferulate 5-hydroxylase, chitinase, glucanase, isopentenyltransferase, xylanase.

The plant cells, in which the regulatory element of the present invention is capable of causing expression of an exogenous gene, may be of any suitable type. The plant cells may be from monocotyledons (such as grasses 20 from the genera *Lolium*, *Festuca*, *Paspalum*, *Pennisetum*, *Panicum* and other forage and turf grasses, corn, grains, oat, sugarcane, wheat and barley), dicotyledons (such as arabidopsis, tobacco, legumes, alfalfa, oak, eucalyptus and maple) and gymnosperms. Preferably the plant cells are from a monocotyledon, more preferably a grass species such as a ryegrass (*Lolium*) 25 or fescue (*Festuca*) species, even more preferably a ryegrass, most preferably perennial ryegrass (*Lolium perenne*).

The regulatory element according to the present invention may be used to express exogenous genes to which it is operatively linked in the production of transgenic plants.

30 Accordingly, in a further aspect of the present invention there is provided a vector including a regulatory element according to the present

invention.

In a preferred embodiment of this aspect of the invention, the vector may include a regulatory element according to the present invention, an exogenous gene as hereinbefore described, and a terminator; said regulatory element, exogenous gene and terminator being operatively linked, such that said regulatory element is capable of causing expression of said exogenous gene in plant cells. Preferably, said regulatory element is upstream of said exogenous gene and said terminator is downstream of said exogenous gene.

The vector may be of any suitable type and may be viral or non-viral.

10 The vector may be an expression vector. Such vectors include chromosomal, non-chromosomal and synthetic nucleic acid sequences, eg. derivatives of plant viruses; bacterial plasmids; derivatives of the Ti plasmid from *Agrobacterium tumefaciens*; derivatives of the Ri plasmid from *Agrobacterium rhizogenes*; phage DNA; yeast artificial chromosomes; bacterial artificial chromosomes; binary bacterial artificial chromosomes; vectors derived from combinations of plasmids and phage DNA. However, any other vector may be used as long as it is replicable on integrative or viable in the plant cell.

The terminator may be of any suitable type and includes for example polyadenylation signals, such as the Cauliflower Mosaic Virus 35S polyA (CaMV 35S polyA) and other terminators from the nopaline synthase (*nos*) and the octopine synthase (*ocs*) genes.

The vector, in addition to the regulatory element, the exogenous nucleic acid and the terminator, may include further elements necessary for expression of the nucleic acid, in different combinations, for example vector backbone, origin of replication (ori), multiple cloning sites, spacer sequences, enhancers, introns (such as the maize Ubiquitin Ubi intron), antibiotic resistance genes and other selectable marker genes [such as the neomycin phosphotransferase (*npt*2) gene, the hygromycin phosphotransferase (*hph*) gene, the phosphinothricin acetyltransferase (*bar* or *pat*) gene], and reporter genes (such as beta-glucuronidase (GUS) gene (*gusA*)]. The vector may also

- 19 -

contain a ribosome binding site for translation initiation. The vector may also include appropriate sequences for amplifying expression.

The regulatory element of the present invention may also be used with other full promoters or partial promoter elements.

5 As an alternative to use of a selectable marker gene to provide a phenotypic trait for selection of transformed host cells, the presence of the vector in transformed cells may be determined by other techniques well known in the art, such as PCR (polymerase chain reaction), Southern blot hybridisation analysis, histochemical GUS assays, northern and Western blot
10 hybridisation analyses.

Those skilled in the art will appreciate that the various components of the vector are operatively linked, so as to result in expression of said transgene. Techniques for operatively linking the components of the vector of the present invention are well known to those skilled in the art. Such
15 techniques include the use of linkers, such as synthetic linkers, for example including one or more restriction sites.

The vectors of the present invention may be incorporated into a variety of plants, including monocotyledons, dicotyledons and gymnosperms. In a preferred embodiment the vectors are used to transform monocotyledons,
20 preferably grass species such as ryegrasses (*Lolium* species) and fescues (*Festuca* species), more preferably perennial ryegrass (*Lolium perenne*) including forage- and turf- type cultivars.

Techniques for incorporating the vectors of the present invention into plant cells (for example by transduction, transfection or transformation) are
25 well known to those skilled in the art. Such techniques include *Agrobacterium* mediated introduction, electroporation to tissues, cells and protoplasts, protoplast fusion, injection into reproductive organs, injection into immature embryos and high velocity projectile introduction to cells, tissues, calli, immature and mature embryos. The choice of technique will depend largely

- 20 -

on the type of plant to be transformed.

Cells incorporating the vector of the present invention may be selected, as described above, and then cultured in an appropriate medium to regenerate transformed plants, using techniques well known in the art. The culture 5 conditions, such as temperature, pH and the like, will be apparent to the person skilled in the art. The resulting plants may be reproduced, either sexually or asexually, using methods well known in the art, to produce successive generations of transformed plants.

In a further aspect of the present invention there is provided a plant cell, 10 plant, plant seed or other plant part, including, eg. transformed with, a vector of the present invention.

The plant cell, plant, plant seed or other plant part may be from any suitable species, including monocotyledons, dicotyledons and gymnosperms. In a preferred embodiment the plant cell, plant, plant seed or other plant part is 15 from a monocotyledon, preferably a grass species, more preferably a ryegrass (*Lolium* species) or fescue (*Festuca* species), even more preferably perennial ryegrass (*Lolium perenne*), including forage- and turf-type cultivars.

The present invention also provides a plant, plant seed, or other plant part derived from a plant cell of the present invention.

20 The present invention also provides a plant, plant seed or other plant part derived from a plant of the present invention.

In a still further aspect of the present invention there is provided a recombinant plant genome including a regulatory element according to the present invention.

25 In a preferred embodiment of this aspect of the invention the recombinant plant genome further includes an exogenous gene operatively linked to said regulatory element.

- 21 -

In a further aspect of the present invention there is provided a method for expressing an exogenous gene in plant cells, said method including introducing into said plant cells an effective amount of a regulatory element and/or a vector according to the present invention.

5 By "an effective amount" is meant an amount sufficient to result in an identifiable phenotypic change in said plant cells or a plant, plant seed or other plant part derived therefrom. Such amounts can be readily determined by an appropriately skilled person, taking into account the type of plant cell, the route of administration and other relevant factors. Such a person will readily be able
10 to determine a suitable amount and method of administration. See, for example, Maniatis et al, Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory, Cold Spring Harbor, the entire disclosure of which is incorporated herein by reference.

15 The present invention will now be more fully described with reference to the accompanying Examples and drawings. It should be understood, however, that the description following is illustrative only and should not be taken in any way as a restriction on the generality of the invention described above.

In the Figures

20 Figure 1 shows plasmid maps of the three cDNAs encoding perennial ryegrass 4CL homologues.

Figure 2 shows the nucleotide (Sequence ID No: 1) and amino acid (Sequence ID No: 2) sequences of *Lp4CL1*.

Figure 3 shows the nucleotide (Sequence ID No: 3) and amino acid (Sequence ID No: 4) sequences of *Lp4CL2*.

25 Figure 4 shows the nucleotide (Sequence ID No: 5) and amino acid (Sequence ID No: 6) sequences of *Lp4CL3*.

- 22 -

Figure 5 shows amino acid sequence alignment of deduced proteins encoded by *Lp4CL1* (Sequence ID No: 2), *Lp4CL2* (Sequence ID No: 4) and *Lp4CL3* (Sequence ID No: 6).

Figure 6 shows northern hybridisation analysis of developing perennial ryegrass using *Lp4CL1*, *Lp4CL2* and *Lp4CL3* as hybridisation probes. SR: roots from seedlings (3-5 d post-germination), SS: shoots from seedlings (3-5 d post-germination), ML: leaves from 12-week-old plants, MS: stems from 12-week-old plants. Blots were washed in 0.2 X SSPE, 0.1 % SDS at 65 °C. *Lp4CL1*, *Lp4CL2* and *Lp4CL3* do not cross hybridise at this stringency. Sizes are given in kb.

Figure 7 shows northern hybridisation analysis showing the time course of expression of 4CL mRNA in wounded perennial ryegrass leaves. Sizes are given in kb.

Figure 8 shows genomic Southern hybridisation analysis using *Lp4CL1*, *Lp4CL2* and *Lp4CL3* as hybridisation probes. 10 µg of digested perennial ryegrass genomic DNA or 20 µg of digested tall fescue genomic DNA were separated on a 1.0 % agarose gel, transferred to Hybond N⁺ membranes and then hybridised with ³²P labelled *Lp4CL1*, *Lp4CL2* or *Lp4CL3* probes. The ryegrass *Lp4CL1*, *Lp4CL2* and *Lp4CL3* genes reveal homologous sequences in tall fescue and indicate that the ryegrass 4CL genes can be used to isolate and to manipulate the expression of the tall fescue (*Festuca arundinacea*) 4CL genes.

Figure 9 shows restriction map of *LpCCR1*. An *L. perenne* seedling cDNA library constructed in Uni-ZAPTM (Stratagene) was screened in a solution containing 10xPIPES, 50% deionised formamide and 10% SDS at 42°C. Filters were washed at room temperature, three times in 0.1% SDS, 2× SSPE and then twice in 0.1% SDS, 0.2× SSPE. The location of the probe used for northern and Southern hybridisation analyses is indicated by the black line labelled *LpCCR531*.

- 23 -

Figure 10 shows the nucleotide (Sequence ID No: 7) and amino acid (Sequence ID No: 8) sequences of *LpCCR1*.

Figure 11 shows Southern hybridisation analysis of DNA from double haploid (DH) perennial ryegrass using *LpCCR1* as hybridisation probe. 10 μ g of DH genomic DNA was digested with Dral, BamHI, EcoRI, EcoRV, HindIII or XbaI, separated on a 1% agarose gel and then capillary blotted onto nylon membrane (Amersham Hybond-N). The membrane was probed with the digoxigenin (DIG) labelled LpCCR531 fragment at 25ng/ml in the hybridisation solution. Hybridisation was in 4x SSC, 50% formamide, 0.1% N-Lauroyl-sarcosine, 0.02% SDS, 2% Blocking solution at 42°C. The membrane was washed twice for five minutes in 2x SSC, 0.1% SDS at room temperature, then twice for fifteen minutes in 0.5x SSC, 0.1%SDS at 68°C. Molecular weight was determined by comparison to a DIG-labelled marker (Roche Molecular Biochemicals).

Figure 12 shows northern hybridisation analysis of RNA samples from different organs and developmental stages of perennial ryegrass using *LpCCR1* probe. Roots from seedlings (3-5 d post-germination), shoots from seedlings (3-5 d post-germination), roots from seedlings (7-10 d post-germination), leaves from seedlings (7-10 d post-germination), roots from 6 and 10 week old plants, leaves from 6 and 10 week old plants, stems from 6 and 10 week old plants, whole seedling from 11 day old *Phalaris* and 7 day old *Festuca*.

Total RNA was isolated using Trizol (GibcoBRL) and 15 μ g was separated on a 1.2% Agarose gel containing 6% formamide and then capillary blotted onto nylon membrane (Amersham Hybond-N). The membrane was stained with 0.2% methylene blue/0.3M sodium acetate to visualise the marker and ensure that RNA was evenly loaded. 50 ng LpCCR531 was random-labelled with 32 P-dCTP (Amersham Megaprime) and hybridisation conditions were 4x SSC, 50% formamide, 0.5% SDS, 5x denhardt solution, 5% dextrane sulphate, 0.1% Herring sperm DNA at 42°C over-night. The ryegrass *LpCCR1*

- 24 -

gene reveal homologous transcripts in tall fescue and *Phalaris*, thus indicating that the ryegrass CCR gene can be used to manipulate the expression of the tall fescue (*Festuca arundinacea*) and *Phalaris* CCR endogenous genes.

Figure 13 shows the nucleotide (Sequence ID No: 9) and amino acid (Sequence ID No: 10) sequences of *LpCAD1*.
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Figure 14 shows the nucleotide (Sequence ID No: 11) and amino acid (Sequence ID No: 12) sequences of *LpCAD2*.

Figure 15 shows a plasmid map of a cDNA clone encoding perennial ryegrass CAD homologue *LpCAD1*.

10 Figure 16 shows northern hybridisation analysis of RNA samples from different organs and developmental stages of perennial ryegrass using A) *LpCAD1* and B) *LpCAD2* as hybridisation probes. Roots from seedlings 3-5 d post-germination, 7-10 d post-germination, 6 weeks and 10 weeks, Shoots from seedlings 3-5 d post-germination and 7-10 d post-germination, Leaves
15 from 6 week old and 10 week old plants, stem tissue from 6 and 10 week old plants. RNA isolated from *Phalaris* and *Festuca* 11 and 7 day old seedlings. The ryegrass CAD genes reveal homologous transcripts in tall fescue and *Phalaris*, thus indicating that the ryegrass CAD gene can be used to manipulate the expression of the tall fescue and *Phalaris* CAD endogenous
20 genes.

Figure 17 shows genomic Southern hybridisation analysis. 10 µg of perennial ryegrass genomic DNA digested with a range of restriction enzymes was separated on a 0.8% agarose gel, transferred to Hybond N and then hybridised with a DIG labelled A) *LpCAD1*, and B) *LpCAD2* hybridisation probe.
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Figure 18 shows the nucleotide sequence of the *LpOmt1* promoter (Sequence ID No: 13).

- 25 -

Figure 19 shows a plasmid map of plant transformation vector carrying the reporter β -glucuronidase (GUS) gene (*gusA*) under control of the perennial ryegrass *LpOmt1* promoter.

Figure 20 (upper image) shows PCR analysis of transgenic tobacco plants containing the *gusA* gene under the control of the perennial ryegrass *LpOMT1* promoter (upper figure). PCR reactions using *gusA*-specific primers were performed. Figure 20 (lower images) show histochemical GUS assays, demonstrating xylem-specific *gusA* expression (A and B) and *gusA* expression in glandular leaf trichomes (C and D) in transgenic tobacco plants containing the *gusA* gene under the control of the perennial ryegrass *LpOMT1* promoter.

Figure 21 shows the isolation of the *LpCCR1* genomic clone 1. A) Southern hybridization analysis of CCR genomic clone λ Lp6.1.1a digested with *Xba*I, *Nco*I, *Sac*I, *Xho*I, *Xho*I/*Sac*I DNA was separated on a 0.8% agarose gel, transferred to Hybond N and hybridized with a DIG labelled CCR1 probe. B) Map showing the genomic gene organisation of *LpCCR1* clone 1 based on sequence results. C) Comparison of plant CCR exon size and number in different plant species (*Lolium perenne*, *Lp.*, *Eucalyptus gunni*, *Eg.*, *Eucalyptus saligna*, *Es.*, *Populus balsamifera*, *Pb.*)

Figure 22 shows the isolation of the *LpCCR1* genomic clone 2. A) Southern hybridization analysis of CCR genomic clone λ Lp6.1.1a digested with *Xba*I, *Nco*I, *Sac*I, *Xho*I, *Xho*I/*Sac*I DNA was separated on a 0.8% agarose gel, transferred to Hybond N and hybridized with 200bp of the CCR1 promoter (Figure 21B). B) Map showing the promoter region of *LpCCR1* clone 2 based on sequence results.

Figure 23 shows the isolation of an *Lp4CL* genomic clone. A) Southern hybridisation analysis of 4CL genomic clone λ Lp4CL2 digested with *Bam*HI, *Kpn*I or *Sac*I. DNA was separated on a 0.8% agarose gel, transferred to Hybond N and hybridized with a DIG labelled 4CL1 hybridisation probe. B) 10 μ l of a standard PCR reaction using forward and reverse oligonucleotides

- 26 -

designed to positions outlined on C). The PCR products were separated on a 0.8% agarose gel and stained with ethidium bromide. C) Map showing the genomic gene organisation of $\lambda Lp4CL2$ based on sequence and PCR results.

Figure 24 shows the isolation of an $Lp4CL$ genomic clone. A) Southern hybridisation analysis of 4CL genomic clone $\lambda Lp4CL2$ digested with BamHI, KpnI, Sall. DNA was separated on a 0.8% agarose gel, transferred to Hybond N and hybridized with a DIG labelled 4CL1 probe. B) Map showing the genomic gene organisation of $Lp4CL2$ clone 1 and the promoter region of clone 2.

Figure 25 shows plasmid map of plant transformation vector carrying the *gusA* gene under control of the perennial ryegrass $Lp4CL2$ promoter ($Lp4CL2::gusA$).

Figure 26 shows nucleotide (Sequence ID No: 14) and amino acid (Sequence ID No: 15) sequences of genomic clone CAD2 cv Barlano (Intron 1 and first 111 bp of the coding region are missing).

Figure 27 shows nucleotide (Sequence ID No: 16) and amino acid (Sequence ID No: 15) sequences of coding sequence deduced from genomic clone CAD2 cv Barlano (region in bold is missing from the genomic clone).

Figure 28 shows the isolation of $LpCAD2$ genomic clone. A) Southern hybridization analysis of CAD genomic clone $\lambda LpCAD2$ digested with BamHI, EcoRI, KpnI, Sall or XbaI. DNA was separated on a 0.8% agarose gel, transferred to Hybond N and hybridized with a DIG labelled CAD2 hybridisation probe. B) Map showing the genomic gene organisation of $\lambda LpCAD2$ based on sequence results.

Figure 29 shows A) Sense and antisense $Lp4CL1$, $Lp4CL2$ and $Lp4CL3$ transformation vectors under control of the CaMV 35S promoter; B) Sense and antisense $Lp4CL1$, $Lp4CL2$ and $Lp4CL3$ transformation vectors under control

- 27 -

of the maize ubiquitin promoter.

Figure 30 shows A) Sense and antisense *LpCCR1* transformation vectors under control of the CaMV 35S promoter; B) Sense and antisense *LpCCR1* transformation vectors under control of the maize ubiquitin promoter.

5 Figure 31 shows A) Sense and antisense *LpCAD1* transformation vectors under control of the CaMV 35S promoter; B) Sense and antisense *LpCAD1* transformation vectors under control of the maize ubiquitin promoter.

Figure 32 shows molecular analysis of *Lp4CL1*-transgenic tobacco. A) Plasmid map of transformation vector carrying a chimeric sense *Lp4CL1* gene.
10 B) PCR analysis of independent transgenic tobacco clones using *Lp4CL1* specific primers. C) Southern hybridization analysis of independent transgenic tobacco plants using an *Lp4CL1* specific probe. D) Northern hybridization analysis of independent transgenic tobacco plants using an *Lp4CL1* specific probe.

15 Figure 33 shows molecular analysis of *LpCCR1*-transgenic tobacco. A) Plasmid map of transformation vectors carrying a chimeric sense and antisense *LpCCR1* gene. B) PCR analysis of independent sense transgenic tobacco clones using *LpCCR1* specific primers.

Figure 34 shows protocol for suspension culture-independent production of transgenic perennial ryegrass plants. A) Isolated zygotic embryos, plated on MSM5 medium, day 0; B) Embryogenic callus formation and proliferation, 6 - 8 weeks after embryo isolation; C) Embryogenic calli arranged on high osmotic MSM3Plus medium prior to biolistic transformation; D) Histochemical GUS assay showing GUS expressing foci 3 – 4 days post-20 bombardment of chimeric *gusA* gene; E) Selection of embryogenic calli on MSM3 medium containing 100 mg/l paromomycin (Pm), 2 weeks after microprojectile bombardment; F) Regeneration of Pm resistant shoots on MSK medium containing 100 mg/l Pm, 4 weeks after microprojectile bombardment;
25 G) *In vitro* plant regeneration from PM resistant embryogenic calli, 6 weeks

- 28 -

after microprojectile bombardment; H) Transgenic perennial ryegrass plants 28 weeks after embryo isolation.

Figure 35 shows molecular analysis of transgenic perennial ryegrass plants carrying sense and antisense *LpOmt1* transgenes. Plasmid maps of vectors used for the co-transformation of perennial ryegrass embryogenic calli; pHp23 carrying a chimeric neomycin phosphotransferase (*npt2*) selectable marker gene; pUbiomt1 carrying a maize ubiquitin promoter driven sense *LpOmt1* gene; pUbitmo1 carrying a maize ubiquitin promoter driven antisense *LpOmt1* gene (top). PCR analysis using *npt2*-specific primers of 5 independent transgenic perennial ryegrass plants from biolistic transformation with sense and antisense *LpOmt1* vectors (upper centre). Southern hybridization analysis with an *omt1* hybridization probe of 7 independent perennial ryegrass plants co-transformed with sense (lanes 1-3) and antisense (lanes 4-7) *LpOmt1* vectors (lower centre left). Southern hybridisation analysis with an *npt2* hybridisation probe of independent perennial ryegrass plants (lower centre right). Northern hybridisation analysis of perennial ryegrass plants co-transformed with antisense *LpOmt1* vector (bottom). C = negative control untransformed perennial ryegrass; P = positive plasmid control.

Figure 36 shows biochemical analysis of *LpOmt1*-transgenic perennial ryegrass. OMT activity of leaf samples from selected independent *LpOmt1*-transgenic perennial ryegrass plants (Ell8, Ell11, Ell14 and Ell15) was determined and compared to untransformed perennial ryegrass negative control plant *L. perenne* cv. Ellett (wild type). Mean values and standard deviations of replicate assays are shown.

Figure 37 shows PCR screening of transgenic ryegrass plants. PCR analysis using *npt2*-specific primers of 8 independent transgenic perennial ryegrass plants from biolistic transformation with antisense *LpUbi4CL2* vector.

Figure 38 shows the nucleotide sequence of genomic clone 4CL2 from perennial ryegrass (Sequence ID No: 17).

- 29 -

Figure 39 shows the nucleotide sequence of genomic clone CCR1 from perennial ryegrass (Sequence ID No: 18).

Figure 40 shows the map location of *Lp4CL1*, *Lp4CL3*, *LpCAD1*, *LpCAD2*, *LpCCR1*, *LpOMT1* and *LpOMT2* (in bold) within the genetic linkage 5 map of perennial ryegrass.

EXAMPLE 1

Isolation and characterisation of three 4-Coumarate CoA-Ligase (4CL) cDNAs from *Lolium perenne*

Materials and Methods

10 Plant material

Plants and embryogenic cell suspensions of perennial ryegrass (*Lolium perenne* L.) cv Ellet and tall fescue (*Festuca arundinacea* Schreb.) cv Triumph were established and maintained as previously described (Heath et al., 1998). Wounding experiments were performed with 10-day-old seedlings of perennial 15 ryegrass (cv Ellet) as previously described (Heath et al., 1998).

Screening of a cDNA library

A cDNA library prepared with RNA isolated from perennial ryegrass seedlings (Heath et al., 1998) was screened with a [³²P]dCTP-labelled rice partial 4CL probe. The rice 4CL probe and consisted of a 844 bp 4CL specific 20 sequence inserted into PUC119. This insert has 93 % sequence identity with a rice 4CL cDNA sequence (Genbank, L43362, bases 453-1300). cDNA inserts were excised and recircularized using the ExAssist helper phage with SOLR strain (Stratagene) as described by the manufacturer.

DNA sequencing

25 cDNA clones were digested with 8 restriction enzymes (*Bam*H_I, *Eco*R_I, *Kpn*I, *Not*I, *Pst*I, *Sall*, *Xba*I, *Xho*I) and selected clones were sequenced on both

- 30 -

strands by the dideoxy chain termination method using M13 forward and reverse primers. For sequencing the internal regions of *Lp4CL1*, *Lp4CL2* and *Lp4CL3* synthetic oligonucleotide primers were designed from the DNA sequences previously determined. Sequencing was performed using the ABI
5 dye terminator kit and automatic sequencer. Nucleotide sequences were aligned using the SeqEd program (ABI) and further analysis was performed using the HIBIO DNASIS vs2 program (Hitachi Software Engineering).

Genomic DNA blot analysis

Genomic DNA was isolated from single genotype-derived cell
10 suspensions of perennial ryegrass and tall fescue according to Lichtenstein and Draper (1985). Ten µg of perennial ryegrass DNA and 20 µg of tall fescue DNA was digested with each of the restriction enzymes *Hind*III and *Xba*I, separated on 1 % agarose gels, and transferred to Hybond N⁺ membranes according to the manufacturer's instructions (Amersham). Probes consisted of
15 *Bam*HI/*Kpn*I fragments of *Lp4CL1* (1771 bp), *Lp4CL2* (2034 bp) or *Lp4CL3* (2080 bp) labelled using the Megaprime labelling kit (Amersham) and [³²P]dCTP. Hybridization was performed at 65 °C in 5 X SSPE, 5 X Denhardt's solution, 0.5 % (w/v) SDS, and 200 µg/mL denatured herring sperm DNA. Membranes were washed three times in 2 X SSPE, 0.1 % SDS for 10 min at
20 25 °C and then twice in 0.1 X SSPE, 0.1 % SDS for 20 min at 65 °C.

RNA blot analysis

Total RNA (10 µg) was separated on 1.2 % formaldehyde gels and transferred to Hybond N (Amersham) membranes according to the manufacturers instructions. Membranes were stained with 0.2 % methylene
25 blue to confirm correct loading and transfer of RNA. Hybridisation was performed at 42 °C in 5 X SSPE, 5 X Denhart's solution, 0.5 % SDS, 50 % deionized formamide, 200 µg/mL denatured herring sperm DNA. Preparation of probes and washing of membranes was as for DNA blot analysis except for the tall fescue Northern blot when the final two washes were performed with
30 0.1 X SSPE, 0.1 % SDS for 10 min at 42°C.

- 31 -

Results

Isolation and sequence analysis of perennial ryegrass 4CL cDNAs

A cDNA library prepared from RNA extracted from perennial ryegrass seedlings was screened with a rice 4CL hybridization probe and ten cDNAs were isolated from 2×10^5 pfu. The cDNAs were characterised by restriction analysis with 8 restriction enzymes. All clones were full length (approximately 2.0-2.2 kb) with poly(A) tails and could be separated into three groups: *Lp4CL1* (four clones) *Lp4CL2* (five clones) and *Lp4CL3* (one clone). Plasmid maps for *Lp4CL1*, *Lp4CL2* and *Lp4CL3* are shown (Figure 1). *Lp4CL1*, *Lp4CL2* and *Lp4CL3* were fully sequenced (Figures 2, 3 and 4, respectively).

Lp4CL1 is 2284 bp long with an open reading frame (ORF) of 1710 bp, a 5' noncoding region of 322 bp and a 3' noncoding region of 252 bp including a poly(A) tail. *Lp4CL2* is 1992 bp long with an ORF of 1668 bp, a 5' noncoding region of 61 bp and a 3' noncoding region of 263 bp including a poly(A) tail. *Lp4CL3* is 2038 bp long with an ORF of 1671 bp, a 5' noncoding region of 112 bp and a 3' noncoding region of 255 bp including a poly(A) tail.

Within the coding region, *Lp4CL1* has 70 % nucleic acid sequence identity with both *Lp4CL2* and *Lp4CL3*, while *Lp4CL2* has 79 % sequence identity with *Lp4CL3*. There is little sequence homology in the 3' noncoding regions between clones (52-55 %).

Amino acid sequence comparisons

The putative proteins encoded by the three cDNAs consist of 570 amino acids [60290 u (Da)] for *Lp4CL1*, 556 amino acids (59238 u) for *Lp4CL2* and 557 amino acids (59735 u) for *Lp4CL3*. The deduced amino acid sequences of *Lp4CL1*, *Lp4CL2* and *Lp4CL3* are shown (Figure 5). *Lp4CL2* and *Lp4CL3* share 79 % amino acid sequence identity, *Lp4CL1* and *Lp4CL2* have 61 % amino acid sequence identity, while *Lp4CL1* and *Lp4CL3* have only 58 % amino acid sequence identity. Regions of high sequence homology are more prevalent in the central and c-terminal regions of the enzyme. For example the

- 32 -

sequence identity between amino acids 208 to 568 of each enzyme is 85 % for *Lp4CL2* and *Lp4CL3*, 72 % for *Lp4CL1* and *Lp4CL2* and 67 % for *Lp4CL1* and *Lp4CL3*.

5 *Lp4CL1*, *Lp4CL2* and *Lp4CL3* share several common regions with other plant 4CLs. In particular, they contain the putative AMP-binding domain and the conserved GEICIRG motif, except for *Lp4CL3* where the second isoleucine has been replaced with valine (Figure 5). It has been proposed that domain II is associated with the catalytic activity of 4CL. Also, four Cys residues conserved in plant 4CLs are conserved in *Lp4CL1*, *Lp4CL2* and *Lp4CL3*
10 (Figure 5). These results suggest that the *L. perenne* cDNAs encode three divergent 4CL enzymes that are likely to have originated from three different 4CL genes.

Expression of perennial ryegrass 4CL genes

15 *Lp4CL1*, *Lp4CL2* and *Lp4CL3* were used as hybridization probes in Northern blots with RNA prepared from different organs of perennial ryegrass at two developmental stages. All three probes hybridized to a single mRNA species of approximately 2.2 - 2.3 kb. *Lp4CL1*, *Lp4CL2* and *Lp4CL3* were expressed at both seedling and mature stages of development and in all organs tested. For *Lp4CL2* and *Lp4CL3* the strongest signal was found in RNA
20 samples from seedling roots and mature stems (Figure 6).

25 *Lp4CL1*, *Lp4CL2* and *Lp4CL3* were also used as hybridization probes in Northern blots with RNA prepared from tall fescue. All three probes hybridized to a similar mRNA species (2.3 kb) as that in perennial ryegrass (Figure 6). The strongest signal was found in RNA samples from mature stems with weaker signals in RNA from roots and seedling shoots. No expression of *Lp4CL1*, *Lp4CL2* or *Lp4CL3* was observed in leaves. The three probes varied in their ability to hybridize to the corresponding homologues in tall fescue, with *Lp4CL3* resulting in the highest signal and *Lp4CL1* hybridizing only weakly.

To determine whether 4CL could be induced under stress conditions,

- 33 -

leaves of perennial ryegrass seedlings were wounded. No increase in the transcript level upon wounding was observed with *Lp4CL1*, *Lp4CL2* or *Lp4CL3* (Figure 7).

Genomic organization of perennial ryegrass 4CL genes

5 Perennial ryegrass DNA was digested with two restriction enzymes, *Hind*III or *Xba*I. Restriction sites for these enzymes are not present in the cDNA sequence of *Lp4CL1*, *Lp4CL2* or *Lp4CL3*. When *Lp4CL1*, *Lp4CL2* or *Lp4CL3* was used as a probe, several DNA hybridizing fragments of varying intensity were revealed (Figure 8). Each probe hybridized to a unique set of
10 fragments, suggesting that *Lp4CL1*, *Lp4CL2* and *Lp4CL3* represent three different genes. Furthermore, *Lp4CL1* and *Lp4CL2* hybridized to 2 to 3 major fragments per digest which may represent either alleles of the same gene or indicate the presence of more than one gene in each class. The *Lp4CL1*,
15 *Lp4CL2* and *Lp4CL3* probes also revealed several different size hybridizing DNA fragments in genomic Southern blots from tall fescue under high stringency conditions (Figure 8), suggesting that three similar 4CL genes are present in *F. arundinacea*.

EXAMPLE 2

Isolation and characterisation of a Cinnamoyl CoA Reductase (CCR)

20 **cDNA from *Lolium perenne***

A total of 500,000 phage were screened from a cDNA library constructed from ten-day-old etiolated *L. perenne* seedlings using a maize CCR probe. Ninety-three positive plaques were observed in the primary screen and five were subsequently analysed by restriction enzyme digestion.
25 Four out of the five were identical. One of the four identical cDNAs, *LpCCR1*, was selected for further analysis (Figure 9).

Nucleic acid sequence analysis of perennial ryegrass CCR cDNA

The full nucleotide sequence of *LpCCR1* was obtained and the amino

- 34 -

acid sequence predicted (Figure 10). *LpCCR1* is a 1395 bp cDNA with 149 bp of 5' non-coding region and 160 bp of 3' non-coding region. An open reading frame of 1086 bp encodes a protein of 362 amino acids. The composition of the coding region was found to be 68% G+C rich. Codon usage was also
5 examined and found to be biased towards XXC/G codons (94%), with XCG and XUA codons accounting for only 9% and 0.55% respectively. G+C richness and bias towards G and C in the third position of a codon triplet are previously reported characteristics of monocot genes.

Genomic organization of perennial ryegrass CCR gene

10 The number of CCR genes present in the ryegrass genome was determined by Southern blot analysis of genomic DNA from double haploid plants, using as probe a fragment of the *LpCCR1* cDNA (LpCCR531, Figure 9). Double haploid DNA reduces the complexity associated with allelic variation. Genomic DNA was cut with enzymes that do not cut the cDNA
15 internally; *Dra*I, *Bam*HI, *Eco*RI, *Eco*RV, *Hind*III and *Xba*I, and the membrane was hybridised and washed under medium-stringency conditions. A single strongly hybridising band was evident in each lane (Figure 11) indicating that there is a single copy of the *LpCCR1* gene in the perennial ryegrass genome.

Expression of perennial ryegrass CCR gene

20 To investigate the expression profile of the CCR gene in ryegrass, northern hybridisation analysis was carried out with total RNA extracted from roots and shoots at seedling growth stages (0.5-1cm and 4-6cm shoots) and roots, stem and leaves at mature growth stages (6 and 10 weeks). Seedlings were grown on filter paper in the dark at 25°C and then transferred to soil and
25 glasshouse conditions (25°C) until the 6 and 10-week stages. Whole seedling total RNA from *Festuca* and *Phalaris* was included in the northern analysis. Hybridisation with *LpCCR531* (Figure 9) was performed at medium-stringency and the membrane was then washed at high-stringency. A transcript of approximately 1.5 kb was detected in all tissues, the level of expression
30 varying with maturity and from one tissue type to another (Figure 12). The *LpCCR1* transcript appears to be more abundant in roots and stem than

- 35 -

shoots and leaves. In the stem, transcript abundance increases from 6-weeks to 10-weeks; indicating that transcription in stem tissue is up-regulated as the plant matures. Expression was found predominantly in tissues such as stems and roots that are forming secondary cell walls indicating that *LpCCR1* is
5 constitutively involved in lignification.

EXAMPLE 3

Isolation and characterisation of Cinnamyl Alcohol Dehydrogenase (CAD) cDNAs from *Lolium perenne*

A 558 bp cinnamyl alcohol dehydrogenase (CAD) fragment was
10 amplified from cDNA synthesised from total RNA prepared from perennial ryegrass seedlings. The conserved amino acid domains between *Pinus radiata*, *Medicago sativa*, *Aralia cordata*, *Eucalyptus botryoides* and *Arabidopsis thaliana* CADs were used to design oligonucleotides for the amplification of the perennial ryegrass CAD. The forward oligonucleotide was
15 designed to the conserved amino acid domain CAGVTVYS and the reverse oligonucleotide to the conserved domain DVRYRFV. The 551 bp PCR fragment was cloned and sequenced to confirm that it corresponded to a perennial ryegrass CAD PCR fragment. A cDNA library prepared from RNA extracted from perennial ryegrass seedlings was screened with the 551bp
20 PCR fragment specific for perennial ryegrass CAD. Eight cDNAs were isolated and separated into six groups by restriction digest analysis. One representative clone each from two groups (*LpCAD1*, *LpCAD2*) were selected for further characterisation.

Nucleic acid sequence analysis of perennial ryegrass CAD cDNAs

The complete sequence of the perennial ryegrass CAD homologue
25 *LpCAD1* was determined (Figure 13). The 1325 bp clone had a poly (A) tail, typical start and stop codons and the open reading frame (ORF) of this clone coded for a putative protein of 408 amino acids.

- 36 -

The complete nucleotide sequence of the perennial ryegrass CAD homologue *LpCAD2* was also determined (Figure 14).

Expression of perennial ryegrass CAD genes

A northern hybridisation analysis with RNA samples isolated from 5 perennial ryegrass at different developmental stages hybridised with the full length *LpCAD1* 1325 bp cDNA (Figure 15) was performed to determine patterns of organ and developmental expression. The probe hybridised to a single mRNA species of approximately 1.6 kb. The *LpCAD1* transcript was expressed in all tissue tested: roots, shoots, stem and leaves (Figure 16A).
10 The *LpCAD1* transcript was most abundant in root tissue and the mature stem, this expression pattern is typical of a gene involved in the lignification of plant cell walls. Intergeneric homologies were revealed in *Festuca* and *Phalaris*.

A similar northern hybridisation analysis was performed with *LpCAD2* (Figure 16B), however the transcript was found to be most abundant in mature 15 stem tissue and the shoots.

Genomic organization of perennial ryegrass CAD genes

A Southern hybridisation analysis using DNA samples isolated from a perennial ryegrass double haploid plant digested with *Dra*I, *Bam*HI, *Eco*RI, *Eco*RV, *Hind*III and *Xba*I and hybridised with a 500 bp *LpCAD1* probe was 20 performed. The hybridisation pattern at high stringency revealed the presence of two prominent bands for most digests indicating that *LpCAD1* belongs to a small gene family and exists a multicity gene in perennial ryegrass (Figure 17A).

A similar Southern hybridization analysis was performed with *LpCAD2* 25 (Figure 17B) the hybridisation pattern at high stringency revealed the presence of one or two prominent bands for most digests indicating that *LpCAD2* exists as a single copy gene or a member of a small gene family in perennial ryegrass (Figure 17B).

EXAMPLE 4**Isolation and characterisation of genomic clones and promoters for O-methyltransferase (OMT), cinnamoyl-CoA reductase (CCR), 4 coumarate CoA-ligase (4CL) and cinnamyl alcohol dehydrogenase (CAD) from**5 ***Lolium perenne***

Genomic clones and promoters of O-methyltransferase (OMT), cinnamoyl-CoA reductase (CCR), 4 coumarate CoA-ligase (4CL) and cinnamyl alcohol dehydrogenase (CAD) were isolated from a perennial ryegrass genomic library using the corresponding cDNAs as hybridisation probes.

10 ***Isolation and characterisation of genomic clones and promoters for perennial ryegrass O-methyltransferase (OMT)***

A perennial ryegrass genomic library was screened with the cDNA clone, *LpOmt1*, (Heath *et al.* 1998) encoding O-methyltransferase (OMT). The sequence of the 5' untranslated region and the coding region was found to be identical to that of the *LpOmt1* cDNA previously isolated. The entire 4.8 kb genomic clone was fully sequenced (Figure 18).

To further characterise the promoters, transcriptional fusions of the promoter sequence to the β-glucuronidase (GUS) coding sequence (*gusA*) have been generated (Figure 19). Direct gene transfer experiments to tobacco protoplasts were performed with the corresponding chimeric genes to transgenically express them in a heterologous system for *in planta* expression pattern analysis by histochemical GUS assays. A set of transgenic tobacco plants carrying a chimeric *gusA* gene under the control of the 5' regulatory region of the *LpOmt1* promoter was generated to assess the potential use of the *LpOmt1* promoter for xylem-specificity and targeted downregulation of genes encoding key lignin biosynthetic enzymes.

The transgenic tobacco plants generated using the *LpOmt1* promoter driven chimeric *gusA* transformation vector were screened by PCR and histochemical GUS assays.

- 38 -

A PCR screening was undertaken using *gusA* specific primers for the initial identification of transgenic tobacco plants (Figure 20). PCR positive tobacco plants were screened by histochemical GUS assays for *in planta* expression pattern analysis (Figure 20).

5 ***Isolation and characterisation of genomic clones and promoters for perennial ryegrass cinnamoyl-CoA reductase (CCR)***

A CCR genomic clone from perennial ryegrass was isolated containing 6.5 kb of promoter and the entire gene organisation (intron/exon boundaries). The CCR promoter can be used for targeted expression of foreign genes in
10 transgenic plants.

A perennial ryegrass genomic library was screened with the cDNA clone *LpCCR1* which codes for the lignin biosynthetic enzyme, cinnamyl-CoA reductase (CCR). Four different genomic clones were identified based on restriction digest analysis. Clone 6.1.1a was selected for further analysis. A
15 6.42 kb *Xhol* fragment from clone 6.1.1a, which hybridized strongly to the *LpCCR1* cDNA probe, was subcloned into pBluescriptSK (Figure 21A). Sequence analysis revealed that the 6.42 kb *Xhol* fragment contained the entire *LpCCR1* gene and 200 bp of promoter region. The intron/exon boundaries are illustrated in figure 21B, the location and the size of the exons
20 appear to be conserved in other CCRs from different species (Figure 21C).

To isolate the promoter region of *LpCCR1*, the Southern blot containing digested phage genomic DNA isolated from clone λ Lp6.1.1a was reprobed with the 200bp promoter region. The probe hybridized strongly to a 6.5 kb *Sall* fragment. This genomic fragment *LpCCR1* clone 2, was subcloned into
25 pBluescriptSK and sequenced (Figure 22A). Sequence results revealed that the 6.5 kb *Sall* fragment contained 6.5 kb of promoter (Figure 22B). The full sequence of *LpCCR1* genomic clone containing the promoter and entire gene sequence (exons and introns) was obtained and is shown on Figure 39.

Isolation and characterisation of genomic clones and promoters for perennial ryegrass 4 coumarate CoA-ligase (4CL)

A 4CL2 genomic clone from perennial ryegrass was isolated containing 2.5 kb of promoter and partial gene organisation (intron/exon boundaries). The 5 4CL2 promoter can be used for targeted expression of foreign genes in transgenic plants. The 2.5 kb promoter has been fused to the reporter gene *gusA* for expression analysis.

A perennial ryegrass genomic library was screened with an *Lp4CL* cDNA probe. After tertiary screening positive 4CL genomic clones were 10 obtained and characterised by restriction digest and Southern hybridisation analysis (Figure 23A).

Sequence analysis revealed that the isolated 4CL genomic clone (4CL2) from perennial ryegrass had 100% nucleotide identity to the *Lp4CL2* cDNA clone. To further characterise this 5 kb λ *Lp4CL2* genomic clone and to 15 confirm that it corresponds to the cDNA of *Lp4CL2*, a number of PCR reactions using primers designed to the cDNA were used. PCR results confirmed that the 5 kb genomic fragment was a partial genomic clone corresponding to the *Lp4CL2* cDNA (Figure 23B). Using primer combinations F1 and R1 the entire 4.8kb genomic fragment was amplified. To determine the 20 location of introns additional PCR reactions using the primer combinations F1 / R2 and F2 / R1 were performed, a 1 kb and 3.5 kb bands were amplified respectively. The location and size of the introns could be determined from these results, and further confirmed by sequence analysis. This large 5 kb genomic fragment contains 4 small exons representing the coding sequence of 25 *Lp4CL2* between 508 bp and 1490 bp (Figure 23C).

The genomic clone 1, *Lp4CL2* contained no promoter region. To isolate the promoter region of *Lp4CL2*, the Southern blot containing digested phage genomic DNA isolated from clone λ *Lp4CL2* was reprobed with a 300 bp EcoRI/BglII isolated from the 5' end of the cDNA clone *Lp4CL2*. The 300 bp 30 probe hybridised strongly to a 2.5 kb BamHI fragment. This genomic fragment

- 40 -

5 *Lp4CL2* clone 2, was subcloned into pBluescriptSK and sequenced (Figure 24A). Sequence results revealed that the 2.5 kb *Bam*HI fragment contained the 508 bp of the 5' ORF of *Lp4CL2* missing from genomic clone 1 and 2.0 kb of promoter region (Figure 24B). The full sequence of the *Lp4CL2* genomic clone containing the promoter and partial gene sequence (exons and introns) was obtained and is shown on Figure 39.

The promoter from *Lp4CL2* was thus isolated and used for the production of a chimeric *gusA* reporter gene (Figure 25).

10 ***Isolation and characterisation of genomic clones and promoters for perennial ryegrass cinnamyl alcohol dehydrogenase (CAD)***

A CAD genomic clone from perennial ryegrass was isolated containing the gene organisation (intron/exon boundaries) minus intron 1 containing the first 111 bp of the CAD coding region. The genomic clone has allowed the identification of a G at position 851 bp in the coding region of the CAD2 15 genomic clone isolated from perennial ryegrass cv. Barlano which is absent in the CAD2 cDNA clone isolated from perennial ryegrass cv. Ellett. The SNP (single nucleotide polymorphism) found to exist between the 2 cultivars has the potential utility as a molecular marker for herbage quality, dry matter digestibility, mechanical stress tolerance, disease resistance, insect pest 20 resistance, plant stature and leaf and stem colour.

25 Results below show the isolation of the genomic clone and sequence analysis of deduced coding sequence from the genomic clone CAD2 from perennial ryegrass cv. Barlano compared to the truncated cDNA CAD2 from the cv Ellett. The missing G in the perennial ryegrass cv. Ellett has been highlighted (Figures 26 and 27).

A perennial ryegrass genomic library was screened with a probe corresponding to the 5' end of the *LpCAD2* cDNA clone, which codes for the lignin biosynthetic enzyme cinnamyl alcohol dehydrogenase. Ten positive plaques were identified and isolated in the primary library screening. After a 30 secondary and tertiary screening, two positive plaques were obtained and

- 41 -

corresponding positive genomic clones were further characterised by restriction digest and Southern hybridization analyses. Both genomic clones were found to be identical based on restriction digest analyses. One clone, named $\lambda LpCAD2$ was chosen for further Southern hybridization analyses. A 5 4.5 kb *Bam*H I fragment which hybridized strongly to the *LpCAD2* cDNA probe was subcloned into pBluescriptSK and sequenced (Figure 28A). Sequence analysis revealed that the 4.5 kb *Bam*H I fragment was a partial genomic clone of *LpCAD2*. This large 4.5 kb genomic fragment contains 4 small exons representing the coding sequence of *LpCAD2* between 213 bp and the stop 10 codon at 1213 bp, and the location of the intron/exon boundaries are illustrated in Figure 28B.

EXAMPLE 5

Development of transformation vectors containing chimeric genes with 4CL, CCR and CAD cDNA sequences from perennial ryegrass

15 To alter the expression of the key enzymes involved in lignin biosynthesis 4CL, CCR and CAD, through antisense and/or sense suppression technology and for over-expression of these key enzymes in transgenic plants, a set of sense and antisense transformation vectors was produced. Transformation vectors containing chimeric genes using perennial ryegrass 20 4CL, CCR and CAD cDNAs in sense and antisense orientations under the control of either the CaMV 35S or the maize ubiquitin promoter were generated (Figures 29, 30 and 31).

EXAMPLE 6

Production and characterisation of transgenic tobacco plants expressing 25 chimeric 4CL, CCR and CAD genes from perennial ryegrass

A set of transgenic tobacco plants carrying chimeric 4CL, CCR and CAD genes from perennial ryegrass were produced and analysed.

- 42 -

Transformation vectors with *Lp4CL1*, *Lp4CL2* and *Lp4CL3* full length cDNA sequences in sense and antisense orientations under the control of either the CaMV 35S or the maize ubiquitin promoters were generated. Transformation vectors with *LpCCR1* cDNA in both sense and antisense orientation under the control of either the CaMV 35S and maize ubiquitin promoters were generated. Transformation vectors with 1325 bp full length *LpCAD1* cDNA in sense and 1051 bp partial *LpCAD1* cDNA in antisense orientation under the control of either the CaMV 35S and maize ubiquitin promoters were generated.

10 Direct gene transfer experiments to tobacco protoplasts were performed using these transformation vectors.

The production and molecular analysis of transgenic tobacco plants carrying the perennial ryegrass *Lp4CL1* and *LpCCR1* cDNAs under the control of the constitutive CaMV 35S promoter is described here in detail.

15 A set of transgenic tobacco plants generated using the *Lp4CL1* sense transformation vector was screened by PCR and subjected to Southern and northern hybridization analyses.

20 A PCR screening was undertaken using *npt2* and *Lp4CL1* specific primers for the initial identification of transgenic tobacco plants. Independent transgenic tobacco plants were identified to be co-transformed with both the selectable marker *npt2* and the *Lp4CL1* chimeric genes (Figure 32).

25 Southern hybridisation analysis was performed with DNA samples from PCR positive transgenic tobacco plants to demonstrate the integration of the chimeric *Lp4CL1* transgene in the tobacco plant genome. Independent transgenic tobacco plants carried between 1 and 5 copies of the *Lp4CL1* transgene. No cross-hybridization was observed between the endogenous tobacco 4CL gene and the perennial ryegrass hybridization probe used (Figure

- 43 -

32).

Northern hybridization analysis using total RNA samples prepared from the transgenic tobacco plants carrying the chimeric sense *Lp4CL1* transgene and probed with the *Lp4CL1*-specific hybridization probe revealed the 5 presence of a 1.2 kb *Lp4CL1* transcript strongly expressed in one *Lp4CL1*-transgenic tobacco plant analysed (Figure 32).

The sense and antisense transformation vectors of *LpCCR1* under the control of the CaMV 35S promoter were introduced into tobacco protoplasts via direct gene transfer. A set of transgenic tobacco plants was generated and 10 screened by PCR with specific primers to identify transgenic tobacco plants carrying chimeric *LpCCR1* transgene. The molecular analysis of *LpCCR1*-transgenic tobacco plants is shown (Figure 33).

EXAMPLE 7

Production and characterisation of transgenic perennial ryegrass 15 plants expressing chimeric OMT, 4CL, CCR and CAD genes from perennial ryegrass

An improved transformation method was developed for the production of transgenic perennial ryegrass plants by biolistic transformation of embryogenic cells. Transgenic perennial ryegrass plants were generated using 20 chimeric OMT, 4CL, CCR and CAD genes from perennial ryegrass and the improved transformation method.

Improved method for the production of transgenic perennial ryegrass plants

This improved procedure utilises embryogenic calli produced from 25 mature seed-derived embryos as direct targets for biolistic transformation without requiring the establishment of embryogenic cell suspensions. The protocol relies on a continuous supply of isolated zygotic embryos for callus induction. Transgenic ryegrass plants can be regenerated 24 – 28 weeks after

- 44 -

embryo isolation (Fig. 34). Isolated embryos are plated onto MSM5 medium to produce embryogenic calli suitable as targets for biolistic transformation within 8 weeks. The embryogenic calli, treated on high-osmoticum medium MSM3 Plus prior to microprojectile bombardment, are selected on MSM3 medium
5 containing 100 mg/l paromomycin (Pm) for 2 weeks before being transferred onto MSK with 100 mg/l Pm for further 4 weeks until differentiation of Pm resistant shoot appear. Regenerated shoots are transferred on to fresh selective media MSK with 100 mg/l Pm for a further 4 weeks (Figure 34).

10 ***Production of transgenic perennial ryegrass plants expressing chimeric OMT, 4CL, CCR and CAD genes from perennial ryegrass***

Transgenic perennial ryegrass (*Lolium perenne*) plants were generated using chimeric ryegrass OMT, 4CL, CCR and CAD genes by biolistic transformation of embryogenic calli. Examples of the production and detailed molecular analysis of these transgenic ryegrass plants are described.

15 Transgenic perennial ryegrass plants for OMT down-regulation were produced using biolistic transformation of embryogenic calli and plant transformation vectors pUbiomt1 and pUbitmo1 carrying *LpOmt1* cDNA sequence in sense and antisense orientation under control of the constitutive maize ubiquitin promoter. These transgenic perennial ryegrass plants for
20 down-regulated OMT activity were regenerated from paromomycin resistant calli obtained from biolistic transformation using microprojectiles coated with two plasmids; pHp23 (carrying the chimeric *npt2* gene as the selectable marker) and either the sense or antisense *LpOmt1* transformation vector driven by the maize *Ubi* promoter.

25 Transgenic perennial ryegrass plants were subjected to a polymerase chain reaction (PCR) screening using *npt2*-specific primers. Independent *npt2* PCR-positive transgenic perennial ryegrass plants obtained from biolistic transformation of embryogenic calli – generated from approximately 60,000 isolated mature seed-derived embryos - using *LpOmt1* sense (pUbiomt1) and
30 *LpOmt1* antisense (pUbitmo1) transformation vectors were identified [16 pUbiomt1 transgenic plants and 27 pUbitmo1 transgenic plants] (Figure 35).

- 45 -

Southern hybridization analysis was performed with undigested and *Hind*III-digested DNA samples prepared from the PCR positive transgenic perennial ryegrass plants, to demonstrate their transgenic nature and the integration of the chimeric *npt2* and *LpOmt1* transgenes. Independent 5 transgenic perennial ryegrass plants co-transformed with both, the selectable marker *npt2* gene and *LpOmt1* chimeric genes, were identified (Figure 35). In most instances, the transgenic perennial ryegrass plants recovered contained multiple copies of the selectable marker gene including rearranged transgene 10 copies. No *npt2*-hybridizing bands were detected in the untransformed negative control.

Samples of *Hind*III-digested genomic DNA were included in the analysis when the *LpOmt1* gene-specific hybridization probe (*omt1*) was used. The *omt1* probe hybridized to a number of bands in DNA samples corresponding to both, the transgenic plants and the untransformed negative control. The *omt1*- 15 hybridizing bands shared in all samples correspond to endogenous *LpOmt1* gene sequences represented as a small multigene family in the perennial ryegrass genome (Heath et al. 1998). The different *omt1*-hybridizing bands evident in the samples from the transgenic plants and absent in the untransformed negative control sample correspond to antisense (*tmo1*) and 20 sense (*omt1*) *LpOmt1* transgene integration events (Figure 35).

Northern hybridization analysis using strand-specific *LpOmt1* probes allowed the identification of transgenic perennial ryegrass plants expressing the antisense *LpOmt1* transgene (Figure 35).

The OMT activity of selected antisense and sense *LpOmt1* transgenic 25 perennial ryegrass plants was determined. Biochemical assays for OMT activity were initially established in untransformed plants (such as tobacco and perennial ryegrass). The assays utilise radiolabelled S-adenosylmethionine as the methyl donor for the OMT-catalysed conversion of caffeic acid into ferulic acid. The production of radioactive ferulic acid is measured and allows the 30 OMT activity to be determined.

- 46 -

The OMT activity of selected *LpOmt1*-transgenic perennial ryegrass plants (*L. perenne* cv. Ellett) was determined. Significantly altered OMT activity in individual transformation events was observed (Figure 36). The manipulation of OMT activity in transgenic perennial ryegrass plants due to the expression of the chimeric ryegrass *LpOmt1* gene was thus demonstrated.

Transgenic perennial ryegrass plants were recovered, using biotic transformation of embryogenic calli, for the manipulation of the expression of genes encoding the key lignin biosynthetic enzyme, 4CL. The plant transformation vectors pUbi4CL2 and pUbi2LC4 carrying chimeric *Lp4CL2* cDNA sequences in sense and antisense orientation, respectively, driven by the constitutive maize ubiquitin (*Ubi*) promoter were used. Perennial ryegrass plants for 4CL manipulation were regenerated from Pm-resistant calli obtained from biotic transformation of embryogenic calli using microprojectiles coated with the plasmids pHp23, carrying a chimeric *npt2* gene as selectable marker gene and the antisense pUbi2LC4.

Transgenic perennial ryegrass plants were subjected to a polymerase chain reaction (PCR) screening using *npt2*-specific primers. Independent *npt2* PCR-positive transgenic perennial ryegrass plants were obtained from biotic transformation of embryogenic calli (Figure 37).

Transgenic perennial ryegrass plants were also recovered, using biotic transformation of embryogenic calli, for the manipulation of the expression of genes encoding the key lignin biosynthetic enzymes, CCR and CAD.

EXAMPLE 8

Genetic mapping of perennial ryegrass OMT, 4CL, CCR and CAD genes

Lp4CL1, *Lp4CL3*, *LpCAD1*, *LpCAD2*, *LpCCR1*, *LpOMT1* and *LpOMT2* clones were PCR amplified and radio-labelled for use as probes to detect

- 47 -

restriction fragment length polymorphisms (RFLPs). RFLPs were mapped using 110 progeny individuals of the p150/112 perennial ryegrass reference population restricted with the enzymes described in the table below.

Clones	Polymorphic in p150/112	Enzyme mapped with	Locus	Linkage group
<i>Lp4CL1</i>	Y	<i>Dra</i> I	<i>Lp4CL1</i>	2
<i>Lp4CL3</i>	Y	<i>EcoRV</i>	<i>Lp4CL3</i>	6
<i>LpCAD1</i>	Y	<i>EcoRV</i>	<i>LpCAD1</i>	2
<i>LpCAD1.2.1</i>	Y	<i>EcoRI</i>	<i>LpCAD2a</i> <i>LpCAD2b</i> <i>LpCAD2c</i>	7 - 2
<i>LpCCR1</i>	Y	<i>EcoRI</i>	<i>LpCCR1</i>	7
<i>LpOMT1</i>	Y	<i>Dra</i> I	<i>LpOMT1</i>	7
<i>LpOMT2</i>	Y	<i>EcoRV</i>	<i>LpOMT2</i>	6

5

Lp4CL1, *Lp4CL3*, *LpCAD1*, *LpCAD2*, *LpCCR1*, *LpOMT1* and *LpOMT2* loci mapped to the linkage groups as indicated in the table and in Figure 40. These gene locations can now be used as candidate genes for quantitative trait loci for lignin biosynthesis associated traits such as herbage quality, dry matter digestibility, mechanical stress tolerance, disease resistance, insect pest resistance, plant stature and leaf and stem colour.

10

REFERENCES

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Lichtenstein, C, And J. Draper (1985) Genetic engineering of plants. In: D.M. Glover (ed.), DNA Cloning, Vol. 2, pp. 67-119, IRL Press, Washington.

20

Finally, it is to be understood that various alterations, modifications and/or additions may be made without departing from the spirit of the present invention as outlined herein.

- 48 -

It will also be understood that the term "comprises" (or its grammatical variants) as used in this specification is equivalent to the term "includes" and should not be taken as excluding the presence of other elements or features.

Documents cited in this specification are for reference purposes only
5 and their inclusion is not an acknowledgement that they form part of the common general knowledge in the relevant art.

CLAIMS

1. A substantially purified or isolated nucleic acid or nucleic acid fragment encoding an enzyme selected from the group consisting of 4 coumarate CoA-ligase (4CL), cinnamoyl-CoA reductase (CCR) and cinnamyl alcohol dehydrogenase (CAD), from a ryegrass (*Lolium*) or fescue (*Festuca*) species.
2. A nucleic acid or nucleic acid fragment according to claim 1 wherein said ryegrass or fescue species is perennial ryegrass (*Lolium perenne*).
3. A nucleic acid or nucleic acid fragment according to claim 1 encoding 4CL and including a nucleotide sequence selected from the group consisting of (a) sequences shown in Figures 2, 3 and 4 hereto (Sequence ID Nos: 1, 3 and 5; respectively) (b) complements of the sequences shown in Figures 2, 3 and 4 hereto (Sequence ID Nos: 1, 3 and 5, respectively); (c) sequences antisense to the sequences recited in (a) and (b); and (d) functionally active fragments and variants of the sequences recited in (a), (b) and (c).
4. A nucleic acid or nucleic acid fragment according to claim 1 encoding CCR and including a nucleotide sequence selected from the group consisting of (a) the sequence shown in Figure 10 hereto (Sequence ID No: 7); (b) the complement of the sequence shown in Figure 10 hereto (Sequence ID No: 7); (c) sequences antisense to the sequences recited in (a) and (b); and (d) functionally active fragments and variants of the sequences recited in (a), (b) and (c).
5. A nucleic acid or nucleic acid fragment according to claim 1 encoding CAD and including a nucleotide sequence selected from the group consisting of (a) the sequences shown in Figures 13, 14, 26 and 27 hereto (Sequence ID Nos: 9, 11, 14 and 16, respectively); (b) complements of the sequences shown in Figures 13, 14, 26 and 27 hereto (Sequence ID Nos: 9,

- 50 -

11, 14 and 16, respectively); (c) sequences antisense to the sequences recited in (a) and (b); and (d) functionally active fragments and variants of the sequences recited in (a), (b) and (c).

6. A vector including a nucleic acid or nucleic acid fragment
5 according to claim 1.

7. A vector according to claim 6 further including a promoter and a terminator, said promoter, nucleic acid or nucleic acid fragment and terminator being operatively linked.

8. A plant cell, plant, plant seed or other plant part, including a
10 vector according to claim 6.

9. A method of modifying lignin biosynthesis in a plant, said method including introducing into said plant an effective amount of a nucleic acid or nucleic acid fragment according to claim 1 and/or a vector according to claim 6.

15 10. Use of a nucleic acid or nucleic acid fragment according to claim 1, and/or nucleotide sequence information thereof, and/or single nucleotide polymorphisms thereof as a molecular genetic marker.

11. A substantially purified or isolated polypeptide from a ryegrass (*Lolium*) or fescue (*Festuca*) species, selected from the group consisting of the
20 enzymes 4CL, CCR and CAD.

12. A polypeptide according to claim 11 wherein said ryegrass or fescue species is perennial ryegrass (*Lolium perenne*).

13. A polypeptide according to claim 11 wherein said polypeptide is 4CL and includes an amino acid sequence selected from the group consisting of sequences shown in Figures 2, 3 and 4 hereto (Sequence ID Nos: 2, 4 and 25 6, respectively); and functionally active fragments and variants thereof.

- 51 -

14. A polypeptide according to claim 11 wherein said polypeptide is CCR and includes an amino acid sequence selected from the group consisting of the sequence shown in Figure 10 hereto (Sequence ID No: 8); and functionally active fragments and variants thereof.

5 15. A polypeptide according to claim 11 wherein said polypeptide is CAD and includes an amino acid sequence selected from the group consisting of the sequence shown in Figures 13, 14, 26 and 27 hereto (Sequence ID Nos: 10, 12, 15 and 17, respectively); and functionally active fragments and variants thereof.

10 16. A lignin or modified lignin substantially or partially purified or isolated from a plant, plant seed or other plant part according to claim 8.

15 17. An isolated regulatory element capable of causing expression of an exogenous gene in plant cells, wherein said regulatory element is isolated from a nucleic acid or nucleic acid fragment encoding a protein selected from the group consisting of: O-methyl transferase (OMT), 4 coumarate CoA-ligase (4CL), cinnamoyl-CoA reductase (CCR) and cinnamyl alcohol dehydrogenase (CAD).

18. A regulatory element according to claim 17 wherein said regulatory element includes an O-methyltransferase promoter.

20 19. A regulatory element according to claim 17 wherein said regulatory element includes a 4 coumarate CoA-ligase promoter.

20. A regulatory element according to claim 17 wherein said regulatory element includes a cinnamoyl CoA-reductase promoter.

25 19. A regulatory element according to claim 17 from a ryegrass (*Lolium*) or Fescue (*Festuca*) species.

20. A regulatory element according to claim 17 including the first

- 52 -

approximately 4630 nucleotides of the sequence shown in Figure 18 hereto (Sequence ID No: 13); or a functionally active fragment or variant thereof.

21. A regulatory element according to claim 17 including the first approximately 2206 nucleotides of the sequence shown in Figure 38 hereto
5 (Sequence ID No: 17); or a functionally active fragment or variant thereof.

22. A regulatory element according to claim 17 including the first approximately 6735 nucleotides of the sequence shown in Figure 39 hereto (Sequence ID No: 18); or a functionally active fragment or variant thereof.

23. A vector including a regulatory element according to claim 17.

10 24. A vector according to claim 23 further including an exogenous gene and a terminator, said regulatory element, exogenous gene and terminator being operatively linked, such that said regulatory element is capable of causing expression of said exogenous gene in plant cells.

15 25. A plant cell, plant, plant seed or other plant part, including a vector according to claim 23.

26. A method for expressing an exogenous gene in plant cells, said method including introducing into said plant cells an effective amount of a regulatory element according to claim 17 and/or a vector according to claim 23.

1/76

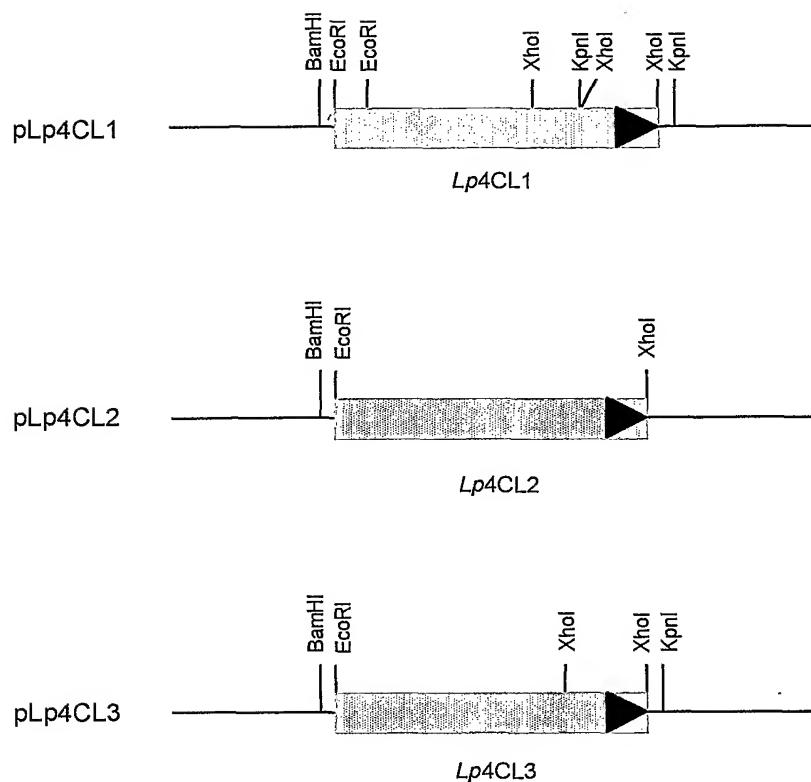


FIGURE 1

1	CGGCACGAGTGGACTTTCCGACGCCGGAGTCGCCGATGATGACCGCCTGAGGAGGTAGT -----+-----+-----+-----+-----+-----+-----+-----+	60
61	CGTAGTCGTCCCTCCGCCCTGTACGCGCCGCTGCCGCCATTTCCTTCCTCGCCTCGCGG -----+-----+-----+-----+-----+-----+-----+-----+	120
121	TCCTCCTCCCCGACCTGCGCTAGGCTCTGGATCTCGCGGGTTGGCGCGCGTCC -----+-----+-----+-----+-----+-----+-----+-----+	180
181	CTGTGAGCTCGTGCGAATTGGCACGCCACCTTCGAGGCGTGCAGTGGTACGAGCTC -----+-----+-----+-----+-----+-----+-----+-----+	240
241	GCGAGCCATTGTCAGTGCAGTGAGGCTCTGCTACTCGTGGCCATTCCAAGAACGCTC -----+-----+-----+-----+-----+-----+-----+-----+	300
301	TGCTCCCTGAAACCAGAGGATCATGATCACGGTGGCGGCCGAGGTGCAGCAGCGCA -----+-----+-----+-----+-----+-----+-----+-----+	360
	M I T V A A P E V Q Q P Q	
361	GATCGCGGGCGGCTGCTGCGGCCGTGGAGGCGGCACCGGAGGCACGACGATCTCCG -----+-----+-----+-----+-----+-----+-----+-----+	420
	I A A A A A V E A A A P E A T T I F R	
421	GTCCAGGCTCCGGACATCGACATCCGACCCACATGCCCTGCACGACTATTGCTTCGC -----+-----+-----+-----+-----+-----+-----+-----+	480
	S R L P D I D I P T H M P L H D Y C F A	
481	GACGGCAGCCTCGGCCCCGGACGCCGCGCTGCCTCATCACCGCGGCCACGGGGAAAGACCTA -----+-----+-----+-----+-----+-----+-----+-----+	540
	T A A S A P D A P C L I T A A T G K T Y	
541	CACGTTGCCGAGACGCACCTGCTGTGCCGCAAGGCCGGCGCTGCACGGGCTCGG -----+-----+-----+-----+-----+-----+-----+-----+	600
	T F A E T H L L C R K A A A A A L H G L G	
601	CGTGGCCACGGGGACCGGATCATGCTGCTGCCAGAACCTCGTGGAGTCGCGCTCGC -----+-----+-----+-----+-----+-----+-----+-----+	660
	V R H G D R I M L L Q N S V E F A L A	
661	CTTCCTCGCGCGTCCATGCTCGCGCCGTCAAGCACGGCGGAACCGTTCTGCACGCC -----+-----+-----+-----+-----+-----+-----+-----+	720
	F F G A S M L G A V S T A A N P F C T P	
721	GCAGGAGATCCACAAGCAGCTCGTGGCTCCGGCGGAAGCTGGTCGTACGCAGTCCGC -----+-----+-----+-----+-----+-----+-----+-----+	780
	Q E I H K Q L V A S G A K L V V T Q S A	

FIGURE 2

781	CTACGTCGACAAGCTCCGGCACGAGGCCTCCCCGAATCGGCAGGCCCTCACCGTGAT Y V D K L R H E A F P R I G E A L T V I	840
841	CACCATCGACGAGGACGACGGCACCCCGAACGGCTGCCAGCCTTCTGGGCCCTCGTGTC T I D E D D G T P D G C Q P F W A L V S	900
901	AGCCGCCGACGAGAACAGCGTCCCGGAGTCTCCCATCTGCCGGACGACGCCGTGGCGCT A A D E N S V P E S P I S P D D A V A L	960
961	GCCCTACTCGTCGGGCACGACGGGCTGCCAACGGCGTGGTGCACGCACGGGGGCT P Y S S G T T G L P K G V V L T H G G L	1020
1021	GGTGTGAGCGTGGCGCAGCAGGTGGACGGCGAGAACCGAACCTGCACATGCCGGCGGG V S S V A Q Q V D G E N P N L H M R A G	1080
1081	GGAGGACGTGGTGCCTGCGTGCCTGCCACATCTCTCGCTCAACTCGGTGCT E D V V L C V L P L F H I F S L N S V L	1140
1141	GCTGTGCGCGCTGCCGGCGCCGCGCCGTGATGCTGATGCCATTGAGATGGGGC L C A L R A G A A V M L M P R F E M G A	1200
1201	CATGCTGGAGGGCATCGAGCGGTGGCGCGTCACGGTGGCGCCGTGGTGCCGCCGCTGGT M L E G I E R W R V T V A A V V P P L V	1260
1261	GCTCGCGCTCGCCAAGAACCCGGGTGGAGAACGACGACCTCAGCTCCATTGGATCGT L A L A K N P G V E K H D L S S I R I V	1320
1321	GCTCTCCGGCGCCGCGCCGCTCGCAAGGAGCTCGAGGACGCCCTACGTGGCCGCTGCC L S G A A P L G K E L E D A L R G R L P	1380
1381	GCAGGCCATCTCGGACAGGGCTACGGGATGACGGAGGCCGGCGGTGCTGTCCATGTG Q A I F G Q G Y G M T E A G P V L S M C	1440
1441	CCCGCGTTCGCGCGGGAGCCGACGCCGGCAAGTCCGGCTCGTGC GGACCGTGGTGC P A F A R E P T P A K S G S C G T V V R	1500

FIGURE 2 CONTINUED

1501	CAACGCCAGCTAAGGTGGTCGACCCCGACACCGGCGTCTCCCTCGGCCGCAACCTCCC -----+-----+-----+-----+-----+-----+ N A Q L K V V D P D T G V S L G R N L P	1560
1561	CGCGAGATCTGCATCCGGCCCGCAGATCATGAAAGGATACTTGAATGATCCCGTGGC -----+-----+-----+-----+-----+ G E I C I R G P Q I M K G Y L N D P V A	1620
1621	CACGCCGCGACCATCGACGTCGAGGGTGGCTCACACCGGCGACATCGGCTACGTCGA -----+-----+-----+-----+-----+ T A A T I D V E G W L H T G D I G Y V D	1680
1681	CGACGACGAGGTCTTCATCGTCGACCGCGTCAAGGAGCTCATCAAGTTCAAGGGCTT -----+-----+-----+-----+-----+ D D D E V F I V D R V K E L I K F K G F	1740
1741	CCAGGTACCGCCGGCCGAGCTCGAGGCTCTGCTCATCGCGATCCGTCCATGCCGACGC -----+-----+-----+-----+-----+ Q V P P A E L E A L L I A H P S I A D A	1800
1801	GGCCGTCGTCCCGCAAAAGGATGATGCCGCCGGCGAGGTCCCGTTGCCTCGTGGTCCG -----+-----+-----+-----+-----+ A V V P Q K D D A A G E V P V A F V V R	1860
1861	CGCCGCCACTCCGACATGCCGAGGAGGCCATCAAGGAGTTCGTATCCAAGCAGGTGGT -----+-----+-----+-----+-----+ A A D S D I A E E A I K E F V S K Q V V	1920
1921	GTTCTACAAGAGGCTGCACAAGGTCTACTTCACCCACGCGATAACCAAGTCGGCGTCGGG -----+-----+-----+-----+-----+ F Y K R L H K V Y F T H A I P K S A S G	1980
1981	GAAGATACTCAGGAAAGAACTCAGAGCTAAACTCGCCGCCGGCCACTGCCTGAAGAGT -----+-----+-----+-----+-----+ K I L R K E L R A K L A A P A T A * R V	2040
2041	GGTCATGGCTTCATGCTAACATTGATCAGAAAGGCACTCTAGCATATATGTTCCA -----+-----+-----+-----+-----+ V H G F M L I I S I R K A L L A Y M F H	2100
2101	CCTTTGTTTCATGGAAAGATTGTATTCCAGCTAGTGGCCAGTGACTGAGTAAGGGATG -----+-----+-----+-----+-----+ L L F H L E D C I P A S G Q *	2160
2161	GGGATAAAAGTTTGTCTACGTTTCTTTACGCTACTCTCCATTGGGAGTACAATG -----+-----+-----+-----+-----+ TATCAGGGATTCTGTATTGAAGTTAACAGATTGGTTCAATTATAAAAAAAAAAAAAA	2220
2221	AAAAA 2281 ---- 2284	2280

FIGURE 2 CONTINUED

1	CGGCACGAGCGCCATTCCCTCACCTTCAGCTCCGCCAAAGATTCCATCCGGCGAGATC	60
61	CATGGGCTCCATCGCGCGACGCGCCTCCGCCGGAGCTGGTGTCCGGTCCAAGCTCCC M G S I A A D A P P A E L V F R S K L P	120
121	GGACATCGAGATCCCGACCCACCTGACGCTGCAGGACTACTGCTTCCAGCGCCTGCCGA D I E I P T H L T L Q D Y C F Q R L P E	180
181	GCTCTCCCGCGCGCCTGCCTCATCGACGGCGCCACGGCGCCGCTCACCTACGGCGA L S A R A C L I D G A T G A A L T Y G E	240
241	GGTGGACGCCCTGTCCCCGCCGCTGCGCCGCCGGGCTGCGCCGCTCGCGTCGGCAAGGG V D A L S R R C A A G L R R L G V G K G	300
301	CGACGTGTCATGGCGCTCCCGCAACTGCCGGAGTTCGCCCTCGTGTTCCTCGGC D V V M A L L R N C P E F A F V F L G A	360
361	GGCCCGGCTCGGCCGCCACCACCGCCAACCCGTTCTACACGCCAACGAGATCCA A R L G A A T T T A N P F Y T P H E I H	420
421	CCGCCAGGCCACCGCCGCCGGGCCAGGGTCATGTCACCGAGGGCTGCGCCGTCGAGAA R Q A T A A G A R V I V T E A C A V E K	480
481	GGTGGCGCCTTCGCCGCCGAGAGAGGGATTCCCGTCGTCTCCGTCGACGAGGGCGTCGA V R A F A A E R G I P V V S V D E G V D	540
541	CGGGGGCTGCCTCCGTTGCCGAGACTCTGCTGGGAAGAAAGCGGGGAGCGGTTCGT G G C L P F A E T L L G E E S G E R F V	600
601	CGACGAGGCCGGTCGACCCGACGACGTGGTGGCGCTGCCGTACTCGTCCGGCACCGG D E A V D P D D V V A L P Y S S S G T T G	660
661	CCTGCCAAGGGCGTCATGTCACCCACCGCAGCCTCGTCACCGCGTCGCCAGCAGGT L P K G V M L T H R S L V T S V A Q Q V	720
721	GGACGGTGAGAACCGAACCTGCACTTCAGCTCGTGGACGTGCTGCTGTGCGTGCTGCC D G E N P N L H F S S S D V L L C V L P	780

FIGURE 3

781	GCTGTTCCACATCTACTCGCTCAACTCGGTGCTGCTGCCGGTCTCCGCCGGGTGC L F H I Y S L N S V L L A G L R A G C A	840
841	GATCGTGATCATGCGCAAGTTGACCGACGGCGCGCTGGTGGACCTGGTGCGCACGC I V I M R K F D H G A L V D L V R T H G	900
901	CGTCACCGTGGCGCCATTGCGGCCATCGTGGTGGAGATGCCAAGAGCGCGCGGGT V T V A P F V P P I V V E I A K S A R V	960
961	GACCGCCGCGGACCTGGCGTCCATCCGGCTGGTCATGTCGGGGCGGCCATGGCAA T A A D L A S I R L V M S G A A P M G K	1020
1021	GGAGCTGCAGGACCGCGTTCATGGCCAAGATCCCCAACGCCGTGCTCGGCCAGGGATATGG E L Q D A F M A K I P N A V L G Q G Y G	1080
1081	GATGACCGAGGCCGGCCCTGTGCTGGCGATGTGCCCTGGCCTCGCCAAGGAGGCCGTTCGC M T E A G P V L A M C L A F A K E P F A	1140
1141	GGTCAAGTCCGGTTCTGGCACCCTCGTCAGGAACGCCAGCTCAAGATCGTCGACCC V K S G S C G T V V R N A E L K I V D P	1200
1201	CGACACCGGCCCTCCCTGGCCGAACTGCCGGGGAGATCTGCATCCGGCAAGCA D T G A S L G R N L P G E I C I R G K Q	1260
1261	GATCATGAAAGGTTACCTAAATGATCCGGTGGCCACAAAGAACACCATTGACAAGGACGG I M K G Y L N D P V A T K N T I D K D G	1320
1321	TTGGCTGCATACTGGTGACATTGGTTATGTCGATGATGACGACGAGATCTTATTGCGA W L H T G D I G Y V D D D D E I F I V D	1380
1381	CAGACTGAAGGAGATAATTAAATATAAGGGATTCCAAGTACCTCCGGCGGAACTTGAAGC R L K E I I K Y K G F Q V P P A E L E A	1440
1441	CCTTCTCATTACACACCCTGAAATCAAGGATGCTGCTGCGTATCGATGCAAGACGAACT L L I T H P E I K D A A V V S M Q D E L	1500
1501	TGCTGGTGAAGTTCCGGTTGCGTTGTTGCGGACTGAGGGTTCAAGAGATCAGCGAAAA A G E V P V A F V V R T E G S E I S E N	1560

FIGURE 3 CONTINUED

1561	CGAGATCAAGCAGTCGTTGAAAAGAGGTTGTTCTACAAGAGGATCTGCAAAGTGT -----+-----+-----+-----+-----+-----+ E I K Q F V A K E V V F Y K R I C K V F	1620
1621	CTTCGGATTCCATTCCAAAGAGTCCATCTGGCAAGATCCTCAGGAAGGACCTGAGAGC -----+-----+-----+-----+-----+-----+ F A D S I P K S P S G K I L R K D L R A	1680
1681	AAAGCTGCCGCAGGCATTCCCAGCAGTAATACCACACAGTCCAAAAGCTAAGTCAGATA -----+-----+-----+-----+-----+-----+ K L A A G I P S S N T T Q S K S *	1740
1741	TATTGTTCCAACCTTACACACCTCTGTCCAACACCATGTAATGTTCTTAATATAAACG -----+-----+-----+-----+-----+-----+ -----+-----+-----+-----+-----+-----+	1800
1801	GAAATTATTACATATAGAAGGGCTGATTCTTTACTAGATGTGTCCAACATATGATATG -----+-----+-----+-----+-----+-----+ -----+-----+-----+-----+-----+-----+	1860
1861	CTTGTAGGCCGATGATGTAAACCTGTCATGTATAGATACCGCCTTTTGACAAGAA -----+-----+-----+-----+-----+-----+ -----+-----+-----+-----+-----+-----+	1920
1921	AGGCTGATTATAATGTATACCGTGAACGTGAATATTTGTTCAGGGAGATCAAAAAAAA -----+-----+-----+-----+-----+-----+ -----+-----+-----+-----+-----+-----+	1980
1981	AAAAAAAAAAA -----+--- 1992	

FIGURE 3 CONTINUED

1 CGGCACGAGATCTCCCACGACTAATTAGAAGAAGATTTACTTAGTCTCTGCTTCTCGCT 60
 1-----+-----+-----+-----+-----+-----+-----+
 61 CGATCGCCGGCCGGTGAGGTAGCTAGCTACTCGTACTAGACCATTACCATGGGTC 120
 61-----+-----+-----+-----+-----+-----+-----+
 M G S
 121 CGTGCCGGAGGAGTCAGTGGTGGCGGTGGCACCGCGGAGACGGTGTCCGGTCGAAGCT 180
 121-----+-----+-----+-----+-----+-----+-----+
 V P E E S V V A V A P A E T V F R S K L
 181 CCCCCACATCGAGATCAACAACGAGCAGACGCTGCAGAGCTACTGCTTCGAGAAGATGGC 240
 181-----+-----+-----+-----+-----+-----+-----+
 P D I E I N N E Q T L Q S Y C F E K M A
 241 CGAGGTCGCGTCCCGCCCTGCATCATCGACGGCCAGACGGGCGCTCCTACACCTACAC 300
 241-----+-----+-----+-----+-----+-----+-----+
 E V A S R P C I I D G Q T G A S Y T Y T
 301 GGAGGTCGACTCCCTGACCCGTCGCGCCGGCGGGCTGCGCCGCATGGCGTGGGAA 360
 301-----+-----+-----+-----+-----+-----+
 E V D S L T R R A A A G L R R M G V G K
 361 GGGCGACGTGGTGTGATGAACCTGCTGCGCAACTGCCGGAGTTCGCCTCTCCTCCTGGG 420
 361-----+-----+-----+-----+-----+-----+
 G D V V M N L L R N C P E F A F S F L G
 421 CGCGCGCGGCTGGCGCCACCACCGCCAACCCGTTCTACACCCCGCACGAGAT 480
 421-----+-----+-----+-----+-----+-----+
 A A R L G A A T T T A N P F Y T P H E I
 481 CCACCGCCAGGCGGAGGCGCGGGCGCCAAGCTGATCGTACCGAGGCCTGCGCCGTGGA 540
 481-----+-----+-----+-----+-----+-----+
 H R Q A E A A G A K L I V T E A C A V E
 541 GAAGGTGCTGGAGTTCGCGCGGGGGCGGGCGTGGTACCGTCGACGGAGGC 600
 541-----+-----+-----+-----+-----+-----+
 K V L E F A A G R G V P V V T V D G R R
 601 CGACGGGTGCGTGGACTTCGCGGAGCTGATCGCCGGCGAGGAGCTGCCCGAGGCGAACGA 660
 601-----+-----+-----+-----+-----+-----+
 D G C V D F A E L I A G E E L P E A D E
 661 GGCGGGGTCTCCCCGACGACGTCGTCGCCCTGCCCTACTCCTCCGGCACCCACCGGGCT 720
 661-----+-----+-----+-----+-----+-----+
 A G V L P D D V V A L P Y S S G T T G L
 721 CCCCAAGGGCGTCATGCTACCCACCGCAGCCTCGTCACCAGCGTCGCCAGCTGGTCGA 780
 721-----+-----+-----+-----+-----+-----+
 P K G V M L T H R S L V T S V A Q L V D

FIGURE 4

781	CGGGTCGAACCCATAACGTGTGCTTCAACAAGGACCGACCGCCTGCTGTGCCTGCTGCCGCT G S N P N V C F N K D D A L L C L L P L	840
841	CTTCCACATCTACTCGCTGCACACGGTGTGCTGGCGGGCTCCGCGTCGGCGCCGCGCAT F H I Y S L H T V L L A G L R V G A A I	900
901	CGTCATCATGCGCAAGTTGACGTGGCGCGCTGGTGACCTCGTCCGCGCGACCGCAT V I M R K F D V G A L V D L V R A H R I	960
961	CACCATCGGCCATTGTGCCGCCATCGTGGAGATGCCAAGAGCGACCGCGTCGG T I A P F V P P I V V E I A K S D R V G	1020
1021	CGCCGACGACCTCGCATCCATCGCATGGTGCTCTCCGGCGCGCCATGGGCAAGGA A D D L A S I R M V L S G A A P M G K D	1080
1081	CCTCCAGGACGCCCTCATGGCCAAGATCCCCAACGCCGTGCTGGACAGGGGTACGGGAT L Q D A F M A K I P N A V L G Q G Y G M	1140
1141	GACCGAGGCTGGCCGGTGCTGGCATGTGCCTGGCGTCGCCAAGGAGCCGTTCAAGGT T E A G P V L A M C L A F A K E P F K V	1200
1201	CAAGTCCGGGTGCGGAACCGTGGTGCGCACGCCGAGCTCAAGGTGTCGACCCGA K S G S C G T V V R N A E L K V V D P D	1260
1261	CACCGGCGCATCCCTGGCCGGAACCGCTGGCGAGATTGCGTCCGGGGAAAGCAGAT T G A S L G R N Q P G E I C V R G K Q I	1320
1321	CATGATAGGTTACCTGAACGACCCAGAGTCGACCAAGAACACCATCGACAAGGACGGCTG M I G Y L N D P E S T K N T I D K D G W	1380
1381	GCTGCACACCGGAGACATCGGCTGGATGACGACGAGATCTTCATCGTCGACAG L H T G D I G L V D D D E I F I V D R	1440
1441	GCTCAAGGAGATCATCAAGTACAAGGGCTTCCAAGTGGCGCCGGAGCTCGAGGCCCT L K E I I K Y K G F Q V A P A E L E A L	1500
1501	CCTCCTCACGAACCGGAGGTCAAGGACGCCGCCGTGTAAGGGTGAAGGATGATCTCTG L L T N P E V K D A A V V G V K D D L C	1560

FIGURE 4 CONTINUED

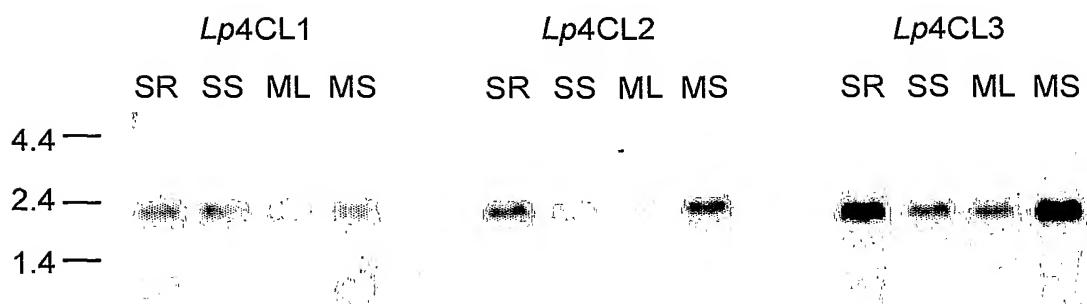
1561	CGGCGAAGTCCGGTCGCCTTCATTAAGAGGATCGAAGGATCTGAGATCAACGAGAACGA -----+-----+-----+-----+-----+-----+-----+ G E V P V A F I K R I E G S E I N E N E	1620
1621	GATCAAGCAATT CGTCTCAAAGGAGGTGTTCTACAAGAGGATCAACAAGGTCTACTT -----+-----+-----+-----+-----+-----+ I K Q F V S K E V V F Y K R I N K V Y F	1680
1681	CACCGACTCCATTCCCAAGAACCCCTCCGGCAAGATCCTAAGGAAGGACTTGAGAGCCAG -----+-----+-----+-----+-----+-----+ T D S I P K N P S G K I L R K D L R A R	1740
1741	GCTCGCCGCTGGCATCCCCACCGAAGTTGCCCGCCGAGAACGCTAAGGGCCGTTCTCAG -----+-----+-----+-----+-----+-----+ L A A G I P T E V A A P R S *	1800
1801	GAACGCAGTCACCCATGGT GCT GTT TAGGT GCT GTT ATAGACCACACCAAATGGGAAAG -----+-----+-----+-----+-----+-----+ 	1860
1861	AAACTACGGGAGGGGATCATATTATTGTTGCAGGAGATATCAGTTGTTGATTGCCCTG -----+-----+-----+-----+-----+-----+ 	1920
1921	CTTG GTAAT GTT GATAAAATGAAATGATATAATAGATGTGTTGTTTATTTTGACCA -----+-----+-----+-----+-----+-----+ 	1980
1981	TGTAAGAACAAAGGCTGTTTATACACTTATTTTGAAAAA -----+-----+-----+-----+-----+-----+ 	2038

FIGURE 4 CONTINUED

FIGURE 5

10	20	30	40	50	60
<i>Lp4CL1</i>	MITVAAPENVQQPQIAAAAAA VEAAAPEATTI	FRSRLPDIDIP	PTHMPILHDYCFATAASAPD		
<i>Lp4CL2</i>	MGSIAADAPPAL..	VFRSKLPDIEIP	THLTLQDYCFQLPELSA		
<i>Lp4CL3</i>	MGSVPEESVVAVAPAETV	FRSKL	PDIEINNEQT	QSYCFEKMAEVAS	
70	80	90	100	110	120
<i>Lp4CL1</i>	APCLITAATGKTYTFAETHL	LCKAAAALHGL	GVRHGDRIMLL	SNSVEFALAFEGASML	
<i>Lp4CL2</i>	RACLIIDGATGAALTYGEVDAL	SRRCAAGLRRRL	GVGKGDVVMALL	RNCPEFAVFLGAARL	
<i>Lp4CL3</i>	RPCIIDGQTGASYTYTEVDSL	TTRRAAGLRRM	VGKGDVVMNLL	RNCPEFAFSFLGAARL	
130	140	150	160	170	180
<i>Lp4CL1</i>	GAVSTAANPFC	TPOEIH	KOLVASGAKL	IVVTQSA	YVOKL
<i>Lp4CL2</i>	GAATT	TANPFC	YTPHEI	HQRATAAGARV	IVTEACAV
<i>Lp4CL3</i>	GAATT	TANPFC	YTPHEI	HROAAAGAKL	IVTEACAV
190	200	210	220	230	240
<i>Lp4CL1</i>	TPDGCQPFWALVSAADE	NSVPESPIS.	PDDAVALPYSSGTT	GLPKGVVL	THGGLVSSVA
<i>Lp4CL2</i>	GVDGGCLPFAET	LLGEESGERFV	DEAVDPDDV	VALPYSSGTT	GLPKGVML
<i>Lp4CL3</i>	RRDGCVDF.	AELIAGEE	LPEADEAGVL.	PDDVVALPYSSGTT	GLPKGVML
250	260	270	280	290	300
<i>Lp4CL1</i>	QKVDG	ENPNLHM	RAGED	DVVL	CALRAGAA
<i>Lp4CL2</i>	QKVDG	ENPNLHF	SS.	SDVLL	AGLRA
<i>Lp4CL3</i>	QKVDG	SNPNVCFNK.	DDALLC	LLPLFHI	YSLHTVLL
310	320	330	340	350	360
<i>Lp4CL1</i>	ERWRVITVA	AVVPPIVL	LALAKNPGV	EKHDLSSIRI	VLSGAAPLGKE
<i>Lp4CL2</i>	RTHGVITV	APFPVPIV	VI	IAKSARVTAADLASIRI	LVMSGAPV
<i>Lp4CL3</i>	RAHRITI	APFPVPIV	VI	IAKSDRVGADDLASIRI	VLSGAAPV
370	380	390	400	410	420
<i>Lp4CL1</i>	QGYGMTEAGPVLSMC	PAFA	REPTPAKSGSCGT	VVRNAOLKVV	VDPTGVS
<i>Lp4CL2</i>	QGYGMTEAGPVLM	AMCL	LAFAKEP	FAVKSGSCGT	VVRNAELKVV
<i>Lp4CL3</i>	QGYGMTEAGPVLM	AMCL	LAFAKEP	EKFVKSGSCGT	VVRNAELKVV
430	440	450	460	470	480
<i>Lp4CL1</i>	RGPQIMKGYLN	DPVATA	ATIDVEGWLHTGDI	GYVDDDDEVF	IIVDRVKE
<i>Lp4CL2</i>	RGPQIMKGYLN	DPVATA	KNTIDKD	GWLHTGDI	GYVDDDDEVF
<i>Lp4CL3</i>	RGPQIMKGYLN	DPESTKNTI	DKD	GWLHTGDI	GYVDDDDEVF
490	500	510	520	530	540
<i>Lp4CL1</i>	ELEALLI	AAHESIADA	AVVPKQDDA	AGEGPVAFVV	RADSDIA
<i>Lp4CL2</i>	ELEALLI	THEIKDA	AVVSMQ	DELAGEGPVAFVV	TEGSEISE
<i>Lp4CL3</i>	ELEALLI	LTNPEVKDAA	AVVGVKDDLC	GEGPVFAFIKR	IEGSEINE
550	560	570			
<i>Lp4CL1</i>	HKVYFTHA	IPKSASGK	KILRKELRA	KLAA	PATA
<i>Lp4CL2</i>	CKVFADS	IPKSPSGK	KILRKDLRA	LAAG	GIPSSNTTQS
<i>Lp4CL3</i>	NKVYFTDS	IPKNPSGK	IILRKDLRA	LAA	GIPTEVAAPRS

FIGURE 6



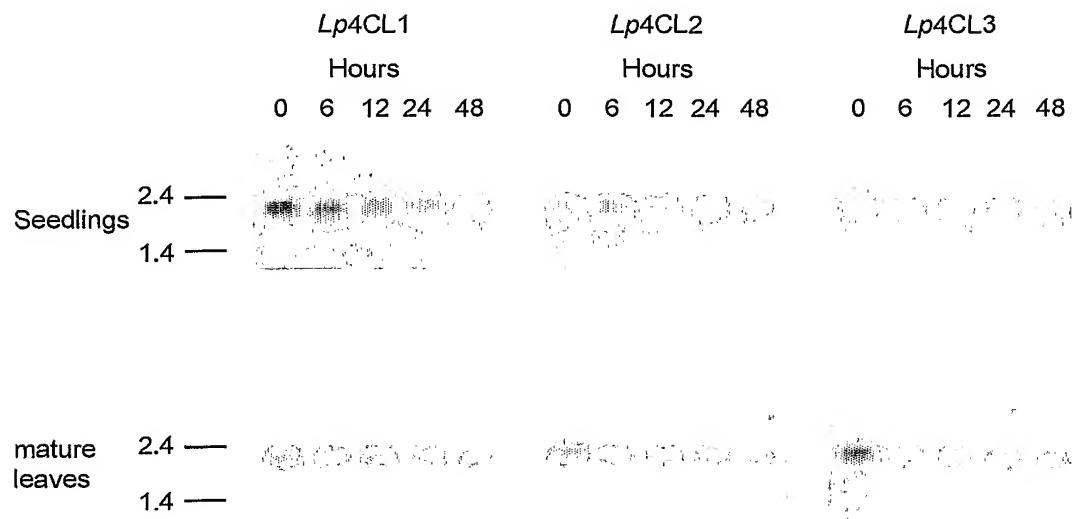


FIGURE 7

FIGURE 8

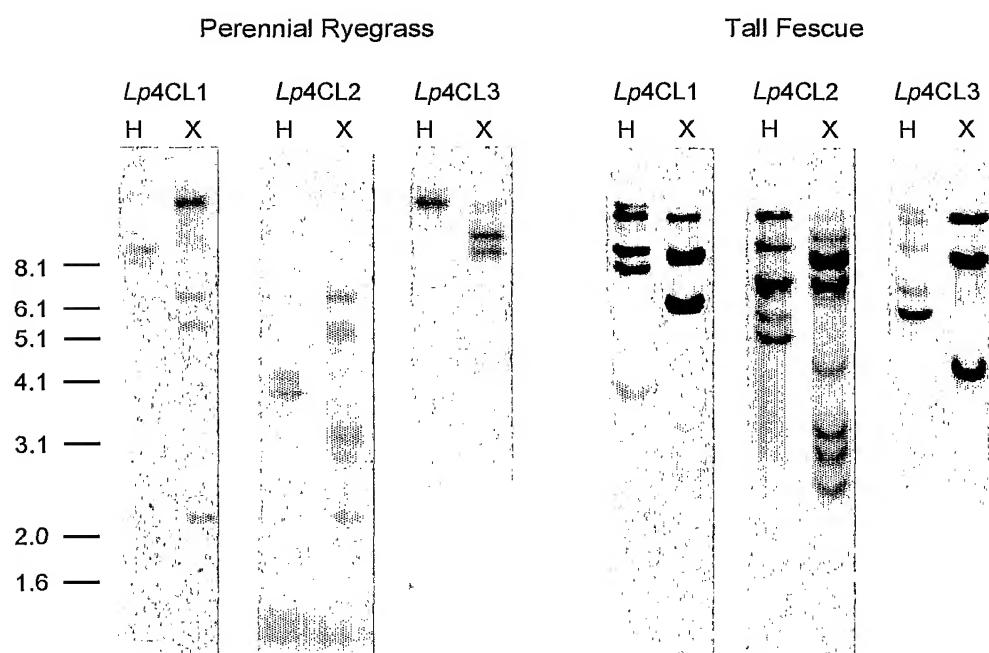
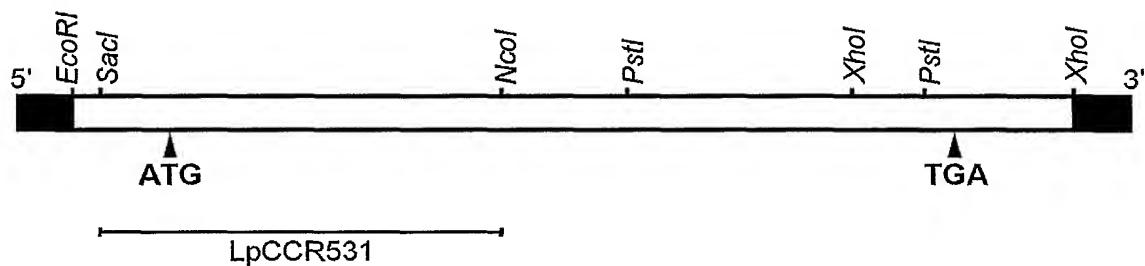


FIGURE 9



16/76

1 GGACGAGGAATCCTACCAACCGAGCTACCAGATCCTCTACTAATCGAGCTCCCTA 60
 61 CGCTGCTCCGCCTGTCTCGTTCCGCCTCACCGCCGGCGGTTCTCCGCTCCAAGCTAC 120
 121 GTCCGTCCGTCCACATATATAGCATCGACATGACCATCGCCGAGGTCTGGCTGCCCGAG 180
 181 M T I A E V V A A G D
 ACACCGCCGCCGCGGGTGGTGCAGCCCGCCGGAAACGGGCAGACCGTGCGTGACCGCGC 240
 181 T A A A V V Q P A G N G Q T V C V T G A
 241 CCGCCGGGTACATCGCGTCGTGGCTCGTCAAGCTGCTGGAGAAGGGGTACACCGTCA 300
 241 A G Y I A S W L V K L L E K G Y T V K
 301 AGGGCACCGTCAGGAACCCAGACGACCCGAAGAACGCGCACCTGAGGGCGCTCGACGGCG 360
 301 G T V R N P D D D P K N A H L R A L D G A
 361 CCGCCGACCGGCTGGTCCTCTGCAAGGCCACCTCCTCGACTACGACGCCATCCGCCCG 420
 361 A D R L V L C K A D L L D Y D A I R R A
 421 CCATCGACGGCTGCCACGGCGTCTCCACACCGCGTCCCCCGTCACCGACGACCCCGAGC 480
 421 I D G C H G V F H T A S P V T D D P E Q
 481 AAATGGTGGAGCCGGCGGTGAGGGCACGCAGTACGTATAGACGCGCGGGAGGCCG 540
 481 M V E P A V R G T Q Y V I D A A A E A G
 541 GCACGGTGGCGGCGATGGTGCACCTCCATCGCCGGCGTCACCATGGACCCCAACC 600
 541 T V R R M V L T S S I G A V T M D P N R
 601 GCGGGCCGGACGTGGTCGTGACGAGTCGTGGAGCGACCTCGACTCTGCAAGAAAA 660
 601 G P D V V V D E S C W S D L D F C K K T
 661 CCAGGAACCTGGTACTGCTACGGGAAGGCGGTTGGAGGCAGGGCATCGGAGTTGGCGC 720
 661 R N W Y C Y G K A V A E Q A A S E L A R
 721 GGCAGCGCGCGTGGACCTTGTGGTGAACCCGGTGCTGGTATCGGCCCCCTGCTGC 780
 721 Q R G V D L V V V N P V L V I G P L L Q

FIGURE 10

781	AGCCGACGGTGAACGCCAGCATCGGCCACATCCTCAAGTACCTGGACGGGTGGCCAGCA -----+-----+-----+-----+-----+-----+-----+ P T V N A S I G H I L K Y L D G S A S K	840
841	AGTTCGCCAACGCCGTGCAGGCGTACGTGGACGTCCCGACGTGGCGACGCCACCTCC -----+-----+-----+-----+-----+-----+ F A N A V Q A Y V D V R D V A D A H L R	900
901	GCGTCTTCGAGTGC CGCCGCCCGTCCGGCCGCCACCTCTGCGCCGAGCGCGTCCCTCCACC -----+-----+-----+-----+-----+-----+ V F E C A A A S G R H L C A E R V L H R	960
961	GCGAGGACGTGCGCATCCTCGCCAAGCTCTCCCCGAGTACCCGTCCCCACCAAGGT -----+-----+-----+-----+-----+-----+ E D V V R I L A K L F P E Y P V P T R C	1020
1021	GCTCTGATGAGACGAACCGAGGAAGCAGCCATACAAGATGTCGAACCAGAAAGCTCCAGG -----+-----+-----+-----+-----+-----+ S D E T N P R K Q P Y K M S N Q K L Q D	1080
1081	ACCTCGGACTCGAGTTCAGGCCGGTGAGCCAGTCCTGTACGAGACGGTGAAGAGCCTCC -----+-----+-----+-----+-----+-----+ L G L E F R P V S Q S L Y E T V K S L Q	1140
1141	AGGAGAAGGGCCACCTTCCGGTGCTCAGCGAGCAGGCAGAGGGGGACAAGGAAACCTTAG -----+-----+-----+-----+-----+-----+ E K G H L P V L S E Q A E A D K E T L A	1200
1201	CTGCCGAGCTGCAGGCAGGGTTACCATCCGAGCATGAGGAACAAGAAATCAACCATGTC -----+-----+-----+-----+-----+-----+ A E L Q A G V T I R A *	1260
1261	CATACTGCTACTGTCATGTAAACCAGCTGTTGAATGCCTAAAATCTAACGTTGTAATA	1320
1321	CTGTGTTGTTCATGTGGACTAGATTGATCGAATAAACATCTCTACACAAGGTTGCTAAA	1380
1381	AAAAAAAAAAAAAA -----+----- 1395	

FIGURE 10 CONTINUED

FIGURE 11

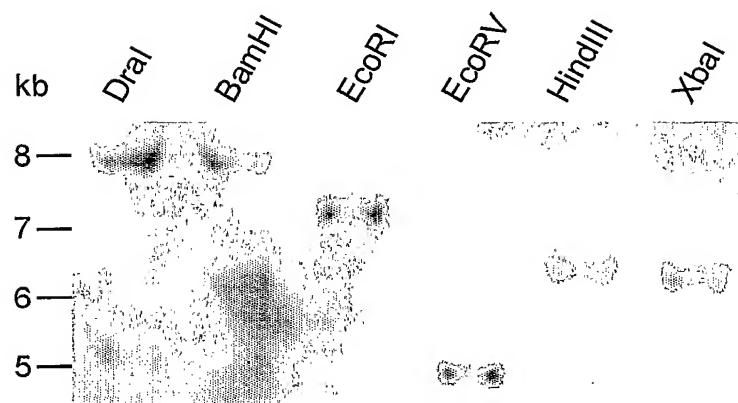
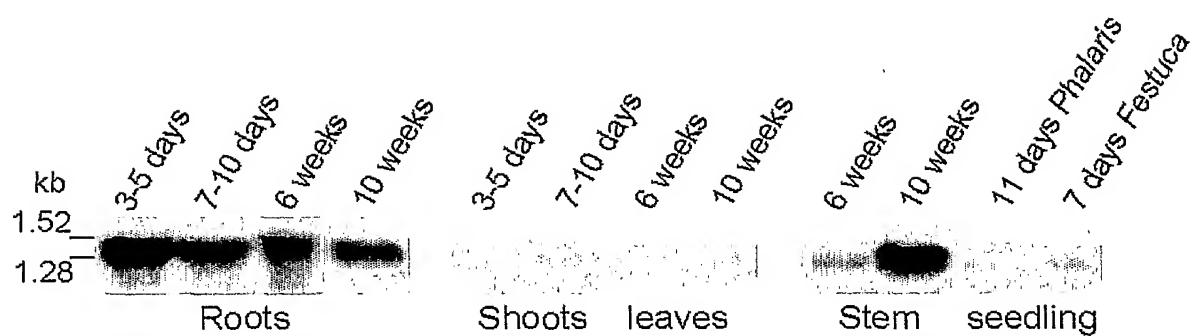


FIGURE 12



1	GGCACGAGCAACAAGTCATCAATGGCGGAAGGCTTGCCTGGCGCTGGTTGGGCTGCGAGG -----+-----+-----+-----+-----+-----+ M A E G L P A L G W A . A R	60
61	GACGCCCTCGGTACACCTCTCCCTTACAGCTCTCGAGAAGCGTTCCGAAGGGACGACgAT -----+-----+-----+-----+-----+-----+ D A S G H L S P Y S F S R S V P K D D D	120
121	GTGACGATCAAGGTGCTCTCTGCCACACTGACCTCCACATCATCAAGAAC -----+-----+-----+-----+-----+-----+ V T I K V L F C G I C H T D L H I I K N	180
181	GACTGGGGCAACGCCCTCTACCCATCGTCCCAGGGCATGAGATCGTGGCGTCGTGCC -----+-----+-----+-----+-----+-----+ D W G N A L Y P I V P G H E I V G V V A	240
241	AGCGTCGGCAGCGCGTCAGCAGCTTCAAGGCCGGCgACACGGTGGCGTGGGCTACTTC -----+-----+-----+-----+-----+-----+ S V G S G V S S F K A G D T V G V G Y F	300
301	CTCGACTCCTGCCGACCTGCTACAGCTGCAGCAAGGGTACGAGAACTTCTGCCACC -----+-----+-----+-----+-----+-----+ L D S C R T C Y S C S K G Y E N F C P T	360
361	CTGACGCTCACCTCCAACGGCGTCAGCGCGGCCACCACCCAGGGCGCTTCTCC -----+-----+-----+-----+-----+-----+ L T L T S N G V D G G G A T T Q G G F S	420
421	GACGTCCCTCGTCGTCAACAAGGACTACGTCATCCGCGTCCGGACAACCTGCCCTGGCC -----+-----+-----+-----+-----+-----+ D V L V V N K D Y V I R V P D N L P L A	480
481	GGCGCGGCACCTCTCCTCTGCGCCGGCGTCACAGTCTACAGCCCTATGGTGGAGTACGGC -----+-----+-----+-----+-----+-----+ G A A P L L C A G V T V Y S P M V E Y G	540
541	CTCAACGCCCGGAcGGAAGCACyTCGGcGTCGTCGGCTGGCGGGCTCGGCCACGTCGcC -----+-----+-----+-----+-----+-----+ L N A P G K H X G V V G L G G L G H V A	600
601	GTCAAGTTGGCAAGGCCTTCGGGATGACCGTCACCGTCATCAGCTCTGGACAGGAAG -----+-----+-----+-----+-----+-----+ V K F G K A F G M T V T V I S S S D R K	660
661	CGCGACGAGGCCGCTCGGCCCTCGGCCGACGCCCTTCCTCGTCAGCAGCGACCCCGAG -----+-----+-----+-----+-----+-----+ R D E A L G R L G A D A F L V S S D P E	720

FIGURE 13

721	CAGATGAAGGCGGC GGCGGGCACCATGGACGGCATCGACACGGTGTCC CGCGGGCCAC -----+-----+-----+-----+-----+-----+ Q M K A A A G T M D G I I D T V S A G H	780
781	CCGATCGTGCCGCTGCTCGACCTGCTCAAGCCC ATGGGGCAGATGGTCGTGGTGGCGCG -----+-----+-----+-----+-----+-----+ P I V P L L D L L K P M G Q M V V V G A	840
841	CCCAGCAAGCCGCTCGAGCTCCGGCTTCGCCATCGGCGGGCAAGCGCCTCGCC -----+-----+-----+-----+-----+ P S K P L E L P A F A I I G G G K R L A	900
901	GGGAGCGGCACCGGCAGCGTCGCACACTGCCagGCCATGCTCGACTTCGCGGGCAAGCAC -----+-----+-----+-----+-----+ G S G T G S V A H C Q A M L D F A G K H	960
961	GGCATCACCGCCGACGTCGAGGTCTCAAGATGGACTACgGTCAACACCGCCATCGAGCG -----+-----+-----+-----+-----+ G I T A D V E V V K M D Y G Q H R H R A	1020
1021	GCTAGAGAAGAACGACGT CAGGTACCGCTTCGTATCGACGTCGCCGGCAGCCACCTGCA -----+-----+-----+-----+-----+ A R E E R R Q V P L R H R R R Q P P A	1080
1081	GGGCACCGCCCTTAAC TTGTGCTACACAATGTGGACGCGCCTCGTTGGTCCAGAAAA -----+-----+-----+-----+-----+ G H R R L T C A T Q C G R A L V W S R K	1140
1141	AGGTTCGCCGGCTCACAGCCACATGAACAAGTCAATGAGTCGTTGGTGTGTTGTTATCT -----+-----+-----+-----+-----+ R F A G S Q P H E Q V N E S L V C C L S	1200
1201	TCATTCCACATATGGGACGCAGTTCCAGATTTCATGTCAAATAATTGCGTCGTGCGG -----+-----+-----+-----+-----+ S F H I W D A V P D F H V K	1260
1261	TTGTCAAGACTCAAATAGGAGAAAAAAAGACTCGTGATTCGTTTGCAAAAAAAAAAAA -----+-----+-----+-----+-----+ AAAAAA	1320
1321	----- 1325	

FIGURE 13 CONTINUED

1	GGCACGAGTCGCCTCCAACGTCTTCCCTTAACC GGCGTCCCTACGCtTGCA CCACCACC	60
61	ACGCACAGACAGAGCAGTTCCCAGCCCCGCCGAACCGGATGGCACCCACGGCGCG M A P T A A E	120
121	AGCAGACGGAGCACCA CAGCAC ACCAGGAAGGCGGTGGGCTGGCGCGCGACGACG Q T E H H Q H T R K A V G L A A R D D A	180
181	CCGGCCACCTCTCCCCGCTGCCATCACACGGAGGAGCACAGGAGACGACGATGTGGTGA G H L S P L A I T R R S T G D D D V V I	240
241	TAAAGATTTGTACTGCGGAATCTGCCACTCTGACCTGCACGCCCTGAAGAACGACTGGA K I L Y C G I C H S D L H A L K N D W K	300
301	AGAACTCAAGGTACCCGATGATCCCCGGCACGAGATGCCGGCGAGGTACGGAGGTGG N S R Y P M I P G H E I A G E V T E V G	360
361	GCAAGAACGTGAGCAAGTTCAAGGCCGGCACCGCGTGGCGTCGGTGCATGGTAACT K N V S K F K A G D R V G V G C M V N S	420
421	CGTGCCGGTCGTGCGAGAGCTGCGACAAGGGCTTCGAGAACCACTGCCGGCATGATCC C R S C E S C D K G F E N H C P G M I L	480
481	TCACCTACAACCTCGGTCGACGTCGACGGCACCGTCACCTACGGCGCTACTCCAGCATGG T Y N S V D V D G T V T Y G G Y S S M V	540
541	TGGTGGTGCACGAGCGGTTCGTGGTCCGGTCCCGACGCCATGCCGCTGGACAAGGGCG V V H E R F V V R F P D A M P L D K G A	600
601	CGCCGCTGCTGCGCCGGCATACCGTGTACAGCCCCATGAAGTACCA CGGGCTCAACG P L L C A G I T V Y S P M K Y H G L N V	660
661	TTCCCGGGCTGCACCTCGGCGTGGCTGGGCTGGCGGGCTGGCCACGTTGCGGTCAAGT P G L H L G V L G L G G L G H V A V K F	720
721	TCGGCAAGGCCTTCGGAATGAAAGTGACGGTATCAGCTCGTCGCCGGGAAGAAGGAGG G K A F G M K V T V I S S S P G K K E E	780

FIGURE 14

23 / 76

781	AGGCCCTGGGGCGGCTGGGCACGCGTTCATCGTCAGCAAGGACGCCGACGAGATGA -----+-----+-----+-----+-----+-----+ A L G R L G A D A F I V S K D A D E M K	840
841	AGGCTGTGATAGCACCATGGATGGCATCANTAAACACGGTATCTGCAAACATCCCCCTGA -----+-----+-----+-----+-----+-----+ A V I A P W M A S X N T V S A N I P L T	900
901	CCCCCTCTCTTCGGGCTGCTCAAGCCAACGGCAAGATGATCATGGTCGGCCTCCCCGAGA -----+-----+-----+-----+-----+-----+ P L F G L L K P N G K M I M V G L P E K	960
961	AGCCCATCGAGATTCCCTCCCTCGCTCTAGTTGCCACGAATAAGACCCCTGGCCGGGAGCA -----+-----+-----+-----+-----+-----+ P I E I P P F A L V A T N K T L A G S I	1020
1021	TCATCGGGCATGAGCGACACGCAGGAGATGCTGGACCTCGCGCGAACGACGGCGTGA -----+-----+-----+-----+-----+-----+ I G G M S D T Q E M L D L A A K H G V T	1080
1081	CGGCCGACATCGAGGTGGTCGGCGCGAGTATGTGAACACGGCCTGGAGCGCCTGCCA -----+-----+-----+-----+-----+-----+ A D I E V V G A E Y V N T A L E R L A K	1140
1141	AGAACGACGTCAGGTATCGCTTCGTATCGACATCGCAACACCCCTCGACAATGTTGG -----+-----+-----+-----+-----+-----+ N D V R Y R F V I D I G N T L D N V A A	1200
1201	CCACCACCGAGTGAACGTACTCAGCACTGCTTACGATCTACGTTGTTCCACTGTTAGTGC -----+-----+-----+-----+-----+-----+ T T E *	1260
1261	TCCGTAGTAAACAATAACGATCAAAACTCTTGTATCTGGTGCATTGGTAGACATGG -----+-----+-----+-----+-----+-----+ TTGTTGCGAGGAAACTGAGTTGAAGGATGGATGGATAAAAAAAAAAAAAAAA	1320
1321		1378

FIGURE 14 CONTINUED

FIGURE 15

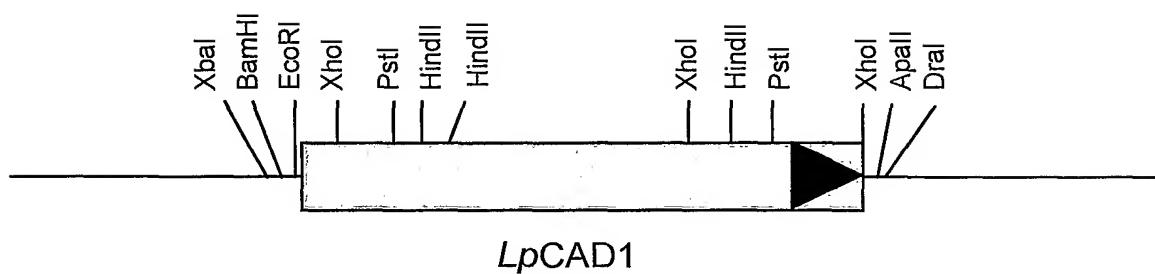
*LpCAD1*

FIGURE 16

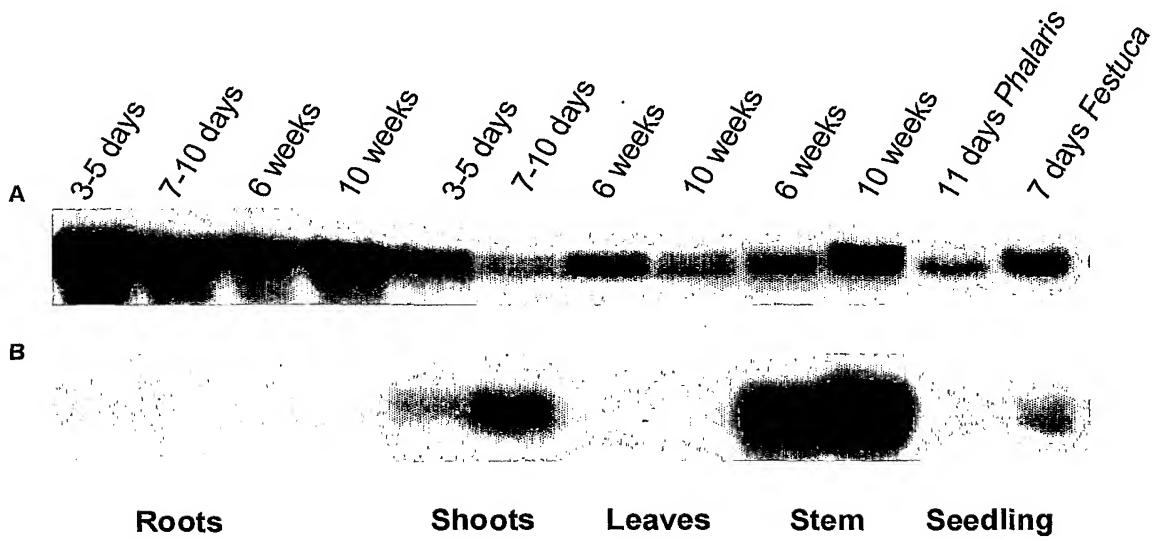
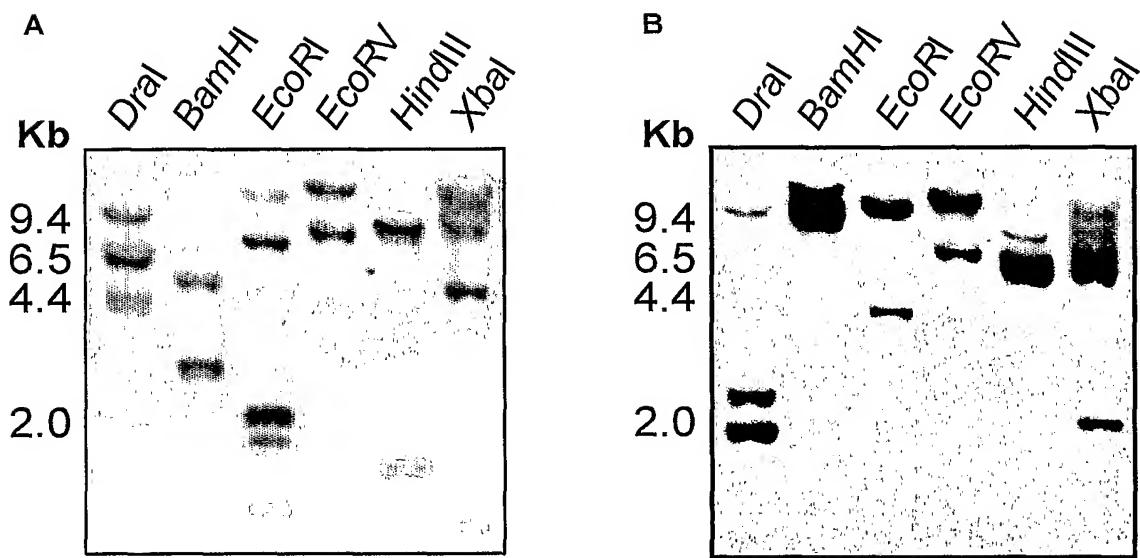


FIGURE 17



26/76

	pBluescript	
-4580	GC GGCCGCTCTAAA ACTAGTGGATCCCCGGCTGCAGGAATTGATATCAAGCTTATnG SalI ATACCGTCGACAGCGGTnCAAATCGCCGGCTCTGGGGTGGAAAGTGnAGCAGTGGGAAGA +	-4581
-4520	TGTGTGCGAGGGGTTGTGTTGGATGnAAGACAGGCCAGTGGAGAACAGAGAGA +	-4521
-4520	ACCGGAGAGGCCAAAGTATCCGCAGCCCCGAAACAAGGCCTAGATTGGGTTAACGTTG +	-4461
-4460	GGTCGTCTCAGACACCGCGGCCATCCTTTAGGTGGTCCGCGCTGGACCCTATTTTTA +	-4401
-4400	TCTGAGTTGACCCATT CAGACGCGCAGACACGAGATGGATGGTGCAGTwAgAGATGACCT +	-4341
-4340	AAGTACAArAACCTCTCCCCGA . GCTGCCGCATCcGTCACTTACCGAGCGAcAAAGTT HindIII +	-4281
-4280	CCCACTTGCATCACACTCAGCCCAGCAAGCATACTGATGGTGAGGCCACTCCGGCTGTGC +	-4221
-4220	CCACCGACCCCCACGCCATCCAAAACCAACTCTACTTTTCACCAmCACCAACAAAAGACAAA +	-4161
-4160	ATATGGTGGATTTGTGATGAGATGGAAGCGGAGCTTGTCAAGATGGAAACGCATAAAAT +	-4101
-4100	CGAGAACACGTATACAGTGCTGGAAATTGGATGACTAAGCCCCAACGGTTAGAAAAAAA +	-4041
-4040	XbaI TnAGACCATGTCTAGATGGAATTAGACATTTTGATATAATAGAAGCGGGACTTGGCGC +	-3981
-3980	GACAATTCAAACCTCGCCCTAACAGGTATCGAActTTCGAtAGTTAGCGTGTGCTACT +	-3921
-3920	GCggAcCCCCAACCAACCACTGTGTTAACGCCACATCgGTAAAGGCCAACGGTTAGATGAAA +	-3861
-3860	GTACCAATCTCACTCATTTGCGACTAGCTACAAAACCTTGCTTTCACATGTACGGTCATA +	-3801
-3800	CTACAATTGGACCTTGGTAACGTAAGTATGGACTGTATGGTGTGCTAACGGTGTGGC +	-3741
-3740	AGCTCAAATAAACCAAAATTCACACACGTCAACCATGAACTGAGATTACACACCAAC +	-3681
-3680	GGCTGAGCCGTCTCCTTAAAGATAGAGGGAGAAAACCATAATCACCATTGGTGGTCAT +	-3621
-3620	GTGTGAGTGTGCAAGCAAAAAAAATGGAGAACGCCAACCCGTTGAGAGAGTGGAGAG +	-3561
-3560	CATACAAGAACACCAACAAAGTGTGAAGGAGAAAAGAATATGAGATAAGATTCGGA +	-3501

FIGURE 18

-3500	AATACTTTGCACACCCATGCATGGGTGTTCCGTACCGTCTATGTATTCCTC +-----+-----+-----+-----+-----+-----+-----+-----+-----+-----	-3441
-3440	GAAATTCACTGCCACCAGGTAGATAAAAATATTTTTCTCTCCTCTTTTATTCAA +-----+-----+-----+-----+-----+-----+-----+-----+-----+-----	-3381
-3380	ATCTCAAAGCATAAkrArTGGTGACAGAACGATAAGATTCTACCTAGCTTCTGAGATC +-----+-----+-----+-----+-----+-----+-----+-----+-----+-----	-3321
-3320	CCACTAGTTATCTCAAGCTGGTGAAGGATTAACCATGCTTGAATTAGATTGGCT +-----+-----+-----+-----+-----+-----+-----+-----+-----+-----	-3261
-3260	TCAAACCTGGTAGTAGCTTGTTCATACTTGATTACTTGGTATGGTAGTTGGTTGA +-----+-----+-----+-----+-----+-----+-----+-----+-----+-----	-3201
-3200	GATTTGGTCAATGTAGAATCAGATTTGAGAGCGATTGTCAGCTTGAATTGCCGCAGTT +-----+-----+-----+-----+-----+-----+-----+-----+-----+-----	-3141
-3140	TAGCACATACTAGTTGGATAGATGAACAGTTGGAGAGACAAATAATGCTATACGAGC +-----+-----+-----+-----+-----+-----+-----+-----+-----+-----	-3081
-3080	TCATCGGATAATATTAGTCTATGGCTTGTGCTCGGTGTCGGCTCTGCAAACCTTACCCC +-----+-----+-----+-----+-----+-----+-----+-----+-----+-----	-3021
-3020	TCTGTAGATGGTAGGATTTCTGATATCCTTCATGGTTAAGGGTGTGCGTGTAAAGGAA +-----+-----+-----+-----+-----+-----+-----+-----+-----+-----	-2961
-2960	CGGGAGATACGGATCACACCTTTCGTCTACACTTACAAGCATGTAACACCTAACGATT +-----+-----+-----+-----+-----+-----+-----+-----+-----+-----	-2901
-2900	GATTGATATCTAGGCCTACACCCATGGAGGTAAACTAATATTATTGAAATGCGACTTT +-----+-----+-----+-----+-----+-----+-----+-----+-----+-----	-2841
-2840	TCAAAAGTCCAATATAACCTTGACGATGATCTTACAACACTCGCGCCAGTCTGTATG +-----+-----+-----+-----+-----+-----+-----+-----+-----+-----	-2781
-2780	<i>KpnI</i> ATATCAGATTGGCCGAGGATCGTGGTACCTTGTAGTGGACTATGATGCTCATGGAGTT +-----+-----+-----+-----+-----+-----+-----+-----+-----+-----	-2721
-2720	GTATGGACATGTTGTAATGCTGGTTCTCTAGGTTTTCTAATCAACTTGGCATTCTT +-----+-----+-----+-----+-----+-----+-----+-----+-----+-----	-2661
-2660	CTCCTAACACATAATAAGAGGAAACACCTCCATACATTATTCTGAAAAAAGCATGGCA +-----+-----+-----+-----+-----+-----+-----+-----+-----+-----	-2601
-2600	ACAATGAAACAGAAAAGTACGACAGTCTATACCCGACCCAAACAATGGCTCAGGTCTT +-----+-----+-----+-----+-----+-----+-----+-----+-----+-----	-2541
-2540	TCACGATGCATAGTTGTTAGCATGTATTTATAGTAGGAACATAAAATTAAAGACAAC +-----+-----+-----+-----+-----+-----+-----+-----+-----+-----	-2481
-2480	TGcnAAAACAATTGTCTTGTAGTGTGTTTAAGGATGCGGCAATTATCGATTATAACA +-----+-----+-----+-----+-----+-----+-----+-----+-----+-----	-2421
-2420	TTACATATGTGATTGGATAGCCAACTTTTGTCTTCCgATGATCATATGAAAGGGTTGT +-----+-----+-----+-----+-----+-----+-----+-----+-----+-----	-2361
-2360	ATCTTAGGGCATCTCCAATGGGnAGACTCAAATGCAAAAAAAAtnGTCCGTTGGGTCTC +-----+-----+-----+-----+-----+-----+-----+-----+-----+-----	-2301
-2300	CnGGACAAAACCTGCTCCCCAACGGGGCAACCCAACTTAAACGGACAGGTGCAAGCGTCC +-----+-----+-----+-----+-----+-----+-----+-----+-----+-----	-2241

FIGURE 18 CONTINUED

28 / 76

-2240	GGCnTGACCCAAAAGTGCACAAATTGGnAnATTTTGGGGCnAGCCAGACGAACGCGG +-----+-----+-----+-----+-----+-----+-----+	-2181
-2180	GCGTCCACTGTATCCGACTATGTCGCCATCCTGGCCCCTGACAGTGACACAAAATACA +-----+-----+-----+-----+-----+-----+-----+	-2121
-2120	ACCACATGCGCCCCCACCCTCTCTCCGTCGCCCTGGAAAnChGTCC +-----+-----+-----+-----+-----+-----+-----+	-2061
-2060	TCGCTCCTCGCCGGAATTGATCTCGCTAACCATGCTCCGCCACCcTCGcCTkAAGG +-----+-----+-----+-----+-----+-----+-----+	-2001
-2000	CCCCAgCCGCCGCTACcTCCTTTGTCAGCCCTATTgGAAGTCGCCgAGTTGAAACGA +-----+-----+-----+-----+-----+-----+-----+	-1941
-1940	GCGCCGCCAGCCTcGACACCGCCGAGCAAGACGAAGACTGGCCGGAGCTGCCGAGACGG +-----+-----+-----+-----+-----+-----+-----+	-1881
-1880	GACGGGGACGGAGCTGCCATGCGTGCCTCGCAGGGGCCGATGGGGCCGGAGCTGCCG +-----+-----+-----+-----+-----+-----+-----+	-1821
<i>PstI</i>		
-1820	TGGCTGG <u>TGCA</u> GACCTCGGGCCGCTGCTAGCCGTGCCACGACGCGAGCATGCGCCTCG +-----+-----+-----+-----+-----+-----+-----+	-1761
-1760	ACGCCGCCCCGTGCTACCTCGTCGCCGCCAGGGCCGCCCGCCCTGCCGACCGgCGg +-----+-----+-----+-----+-----+-----+-----+	-1701
-1700	CGgAgACGCGAcCTTCGCGgACGTGCCGGCGCAGAGACGCCCTCGCACAGCGCC +-----+-----+-----+-----+-----+-----+-----+	-1641
-1640	CTCCTCGATCTCGTCGAGCCGCATAACGCgGcTAGAGGGACGCCGCTCCCGGTGTC +-----+-----+-----+-----+-----+-----+-----+	-1581
-1580	GGCCTCCGTTGTGGCGCATCGCGGCCCTCCGTCGAGGCGCGCGAGCTTCCCTCGCGCG +-----+-----+-----+-----+-----+-----+-----+	-1521
-1520	TCGTGGCGCAGCCTGCCCTGATTGGCTCTGAGGCGCGCGAGCTTCCCTCGCGCG +-----+-----+-----+-----+-----+-----+-----+	-1461
-1460	GCGGGCGGAGCCTCCCTGCGGCCGACCTGCTCTGCCGGTCCGAGACGCCGCG +-----+-----+-----+-----+-----+-----+-----+	-1401
-1400	GGCAGAGCTCCTCGCGCGCTGGGCGCGCTCCCTCGCGCGATGGCGCTTCCAGGC +-----+-----+-----+-----+-----+-----+-----+	-1341
-1340	TCGCACGCCCTCCGGCGTGGCGCAGCAGAGCGCAGCCTCCGGTGAGTTAGGCACAGG +-----+-----+-----+-----+-----+-----+-----+	-1281
-1280	CGCGACACGACATCCCCGGCTCGGCCCTCCGGCGTGGCGCAGCGCAGCGCAGCG +-----+-----+-----+-----+-----+-----+-----+	-1221
-1220	TAGGTTGGCAACTAGTaCTACGAGGAAGAAAGAGGAGAAACAATTATTGGGTACAGCG +-----+-----+-----+-----+-----+-----+-----+	-1161
-1160	TTGGCGTACTGTGCGATCCAAACGGACACCCgGACGCGAaACGATGTCAGCGTGTCCGC +-----+-----+-----+-----+-----+-----+-----+	-1101
-1100	GTGGcGACCCAAACGACCCGAAACGGACGTCcGTGTTGGGTGGCGTTGGAGATGCCCT +-----+-----+-----+-----+-----+-----+-----+	-1041
-1040	TACTCCCCATCCTCAAATGAGTCTAATTATATCTTGTGTAAGTTTAAAAAGTTAA +-----+-----+-----+-----+-----+-----+-----+	-981

FIGURE 18 CONTINUED

29 / 76

-980 ACTTTGATCAACATTAGTAATGATAGTAGCAACGAATACAAAATTAAATTGTAAAAATAT
 -920 ATTATGAAACTTATTAAAGATGGATCTAGTTATACTAATTCTCGGGATGGAGGAAG
 -860 TAGCTAAATATTGTTAATTCTAAATAAAAATTAAAACCTTAACCTAAAACAAAAGTTA
 -800 Putative Myb Binding domains CAAGCATAATTATCTGtGGATGGAGGAAGtAGCTAAGATAACCCAATCCTCTCTCAT
 -740 TACCTAGCATGCCACATCAGGAAACTATTAGGATAAGCTCCAAGGAACCACCCAGAACAA
 -680 ACAATTACATGGCCTGGCTAAACCTAATGACAATTCCGAGCAACTGGTGGTGGTAC
 -620 GCGTTCCCTGTTCAATTGTCCTATTACAAGAGTGGCCCTGTATAGGTAAAAAAAAATAA
 -560 *HindIII* *PstI* CAAGCTTCCAGGACGGCCATGTTCCCTTGCAGGCTGCACGTACTCACGACGAAG
 -500 TGTATCTCGTGTCTGGACATTGTCCTCGCGCATTGTAACCATGAAATTAAAAATGTG
 -440 GTGGCCTGCTATATCTGTATGGGGGTATCATGCACTCCTCGCAGAGGAATCCAGACGAC
 -380 GATTACACGTGTTCCACCTTAGCTTTTAAGTGTGTGTAAGGAACGATCATATA
 -320 *XbaI* ACTGCCCTGAATGCTGCATATATATAACCGACTCCATCATGTACTCGAGACAAGGTCG
 -260 TCAAGAAAACAAACTATGCCATCTCACTAGCAATGATTGAGAGTACAGCTTTCCGG
 -200 TGCCATATTTCCATATATCTTTCTGAAGAACAGAAAAAAACAGTGTGGTGT
 -140 GGTGGTTGGTGAAGCGAGAAAGCCCCATATAAGCCCTGCTCACCCCTCCCGCAAAGCACA
 -80 *PvuI* ACTCATAGCTGGGTCTCGCTCACACCAAAATGCCACCAGCAGCATCTCGA
 -20 TCGGAGACGCATAGATCGATGGGCTCCACCGCCGACATGGCCGCGTCCGGACGA
 M G S T A A D M A A S A D E
 40 GGACGCGTGCATGTTGCCCTCCAGCTCGCTTCTCGTCGGTCCCTCCGATGACGCTGAA
 D A C M F A L Q L A S S S V L P M T L K
 100 GAACGCCATCGAGCTTGGCCCTGGAGATCCTGGTGGCCGCCGGCAAGTCGCTGAC
 N A I E L G L L E I L V A A G G K S L T
 160 CCCGACCGAGGTGGCCGCCAGCTCCCGTCCGGCGAACCCGGAAAGCGCCGGACATGGT
 P T E V A A K L P S A A N P E A P D M V
 219

FIGURE 18 CONTINUED

220 GGACCGCATACTCCGGCTGCTCGCGTCGTACAACGTCGTGACGTGCCTGGTGGAGGAGGG 279
 +-----+-----+-----+-----+-----+-----+-----+
 D R I L R L L A S Y N V V T C L V E E G

 280 CAAGGACGGCCGCCCTCTCCGGAGCTACGGCGCCGCCGTGTGCAAGTTCCCTACCCCC 339
 +-----+-----+-----+-----+-----+-----+-----+
 K D G R L S R S Y G A A P V C K F L T P

 340 CAACGAGGACGGCGTCTCCATGGCGGCCGCTCGCGTCATGAACCAGGACAAGGTCCCTCAT 399
 +-----+-----+-----+-----+-----+-----+-----+
 N E D G V S M A A L A L M N Q D K V L M

 Intron/exon boundary
 400 GGAGAGCTGJGTGAGTCTCTCAGTGGAGCTAGTTACTGTAGATCCGAATTGTTCCCTTA 459
 +-----+-----+-----+-----+-----+-----+-----+
 E S
 460 *SalI* pBluescript
 GTGAGGGTTAATTCCGGCCGCCGTCGACCTCGAGGGGGGCCGGTACCCAATTGCC 744
 +-----+-----+-----+

TATACTGAGTCGTATTACGCGCGCTCACTGGCGCTCGTTTACAACGTCGTGACTGGGAA
 AACCCCTGGCGTTACCCAACCTTAATCGCCTTGCAAGCACATCCCCCTTCGCCAGCTGGCGT
 AATAGCGAAGAGGCCCGCACCGATCGCCCTTCCAACAGTTGCGCAGCTGAATGGCGAA
 TGGGACGCGCCCTGTAGCGCGCATTAAAGCGCGCGGGTGTGTG

FIGURE 18 CONTINUED

FIGURE 19

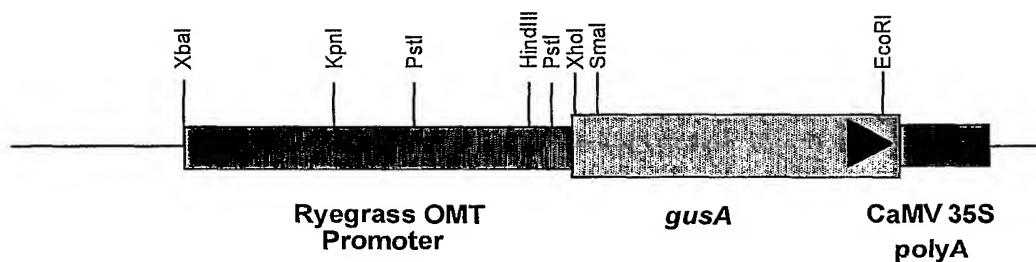


FIGURE 20

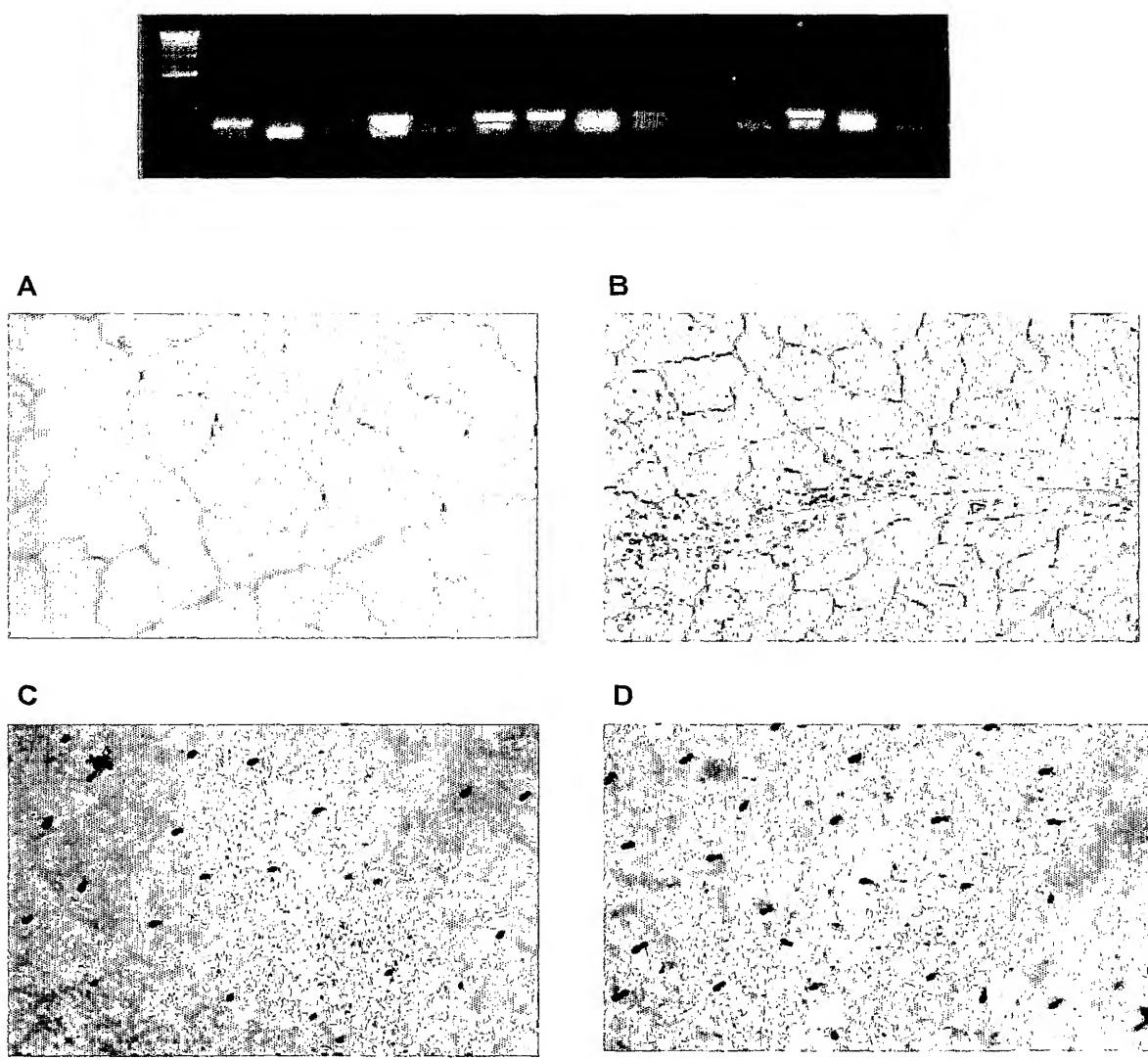
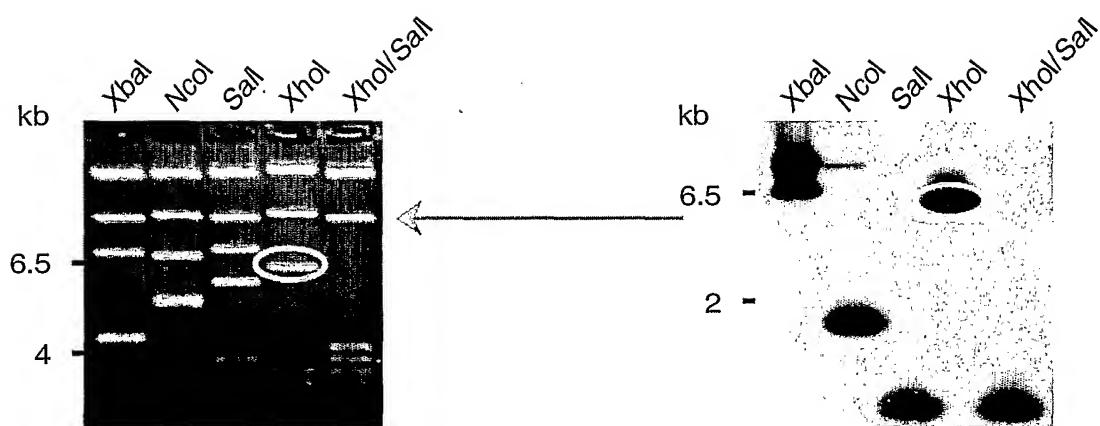
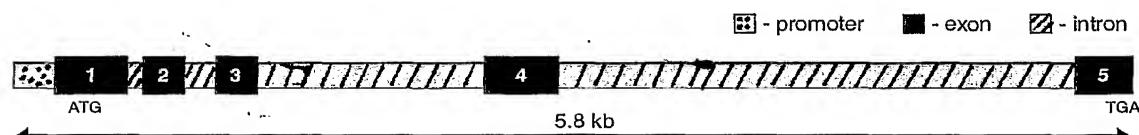


FIGURE 21

A**B****C**

Exon	<i>LpCCR1</i>	<i>EgCCR1</i>	<i>EsCCR1</i>	<i>PbCCR1</i>
1	173 bp	133 bp	133 bp	139 bp
2	155 bp	155 bp	155 bp	155 bp
3	189 bp	186 bp	186 bp	186 bp
4	353 bp	353 bp	353 bp	353 bp
5	220 bp	218 bp	184 bp	184 bp

FIGURE 22

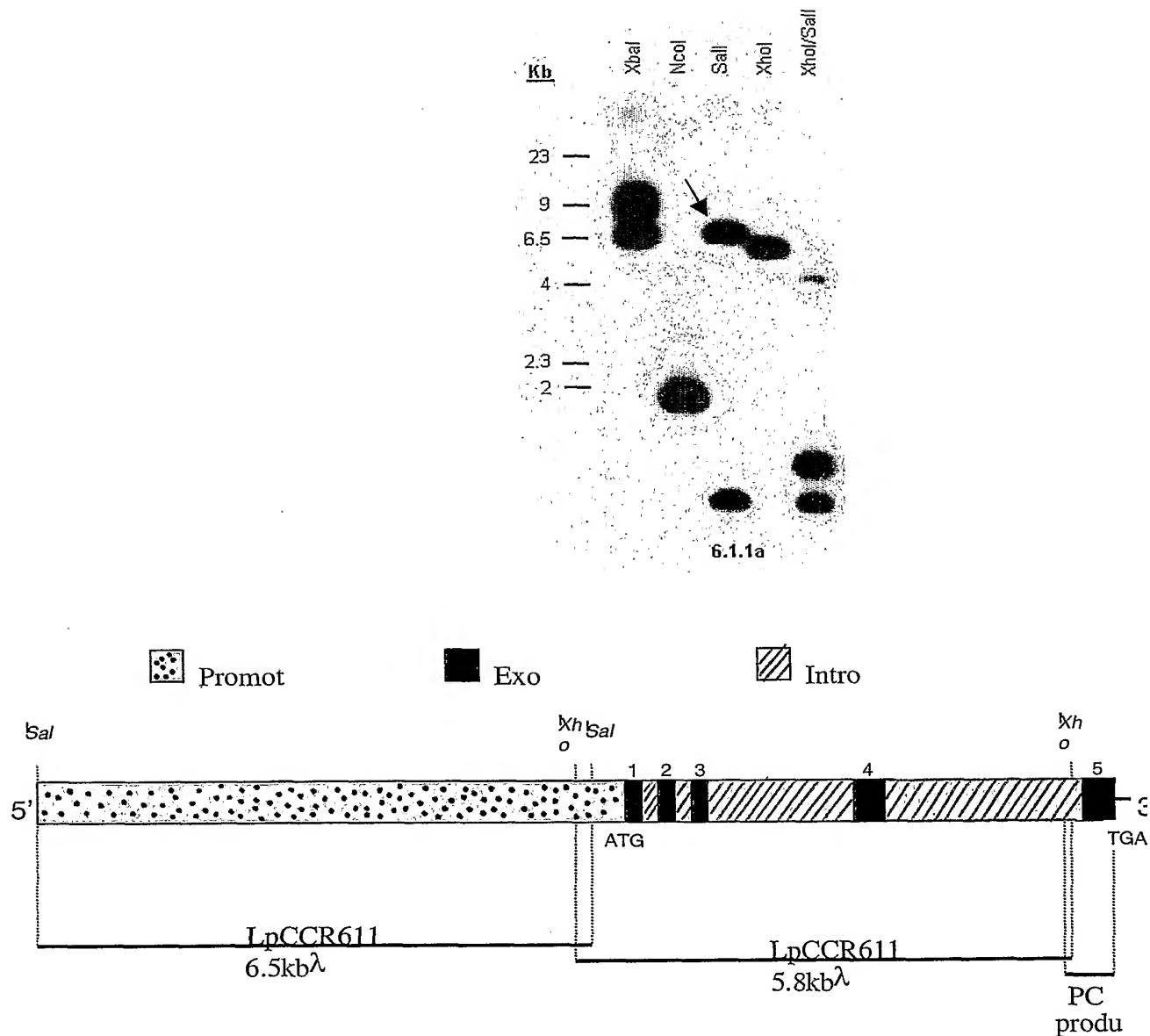
A

FIGURE 23

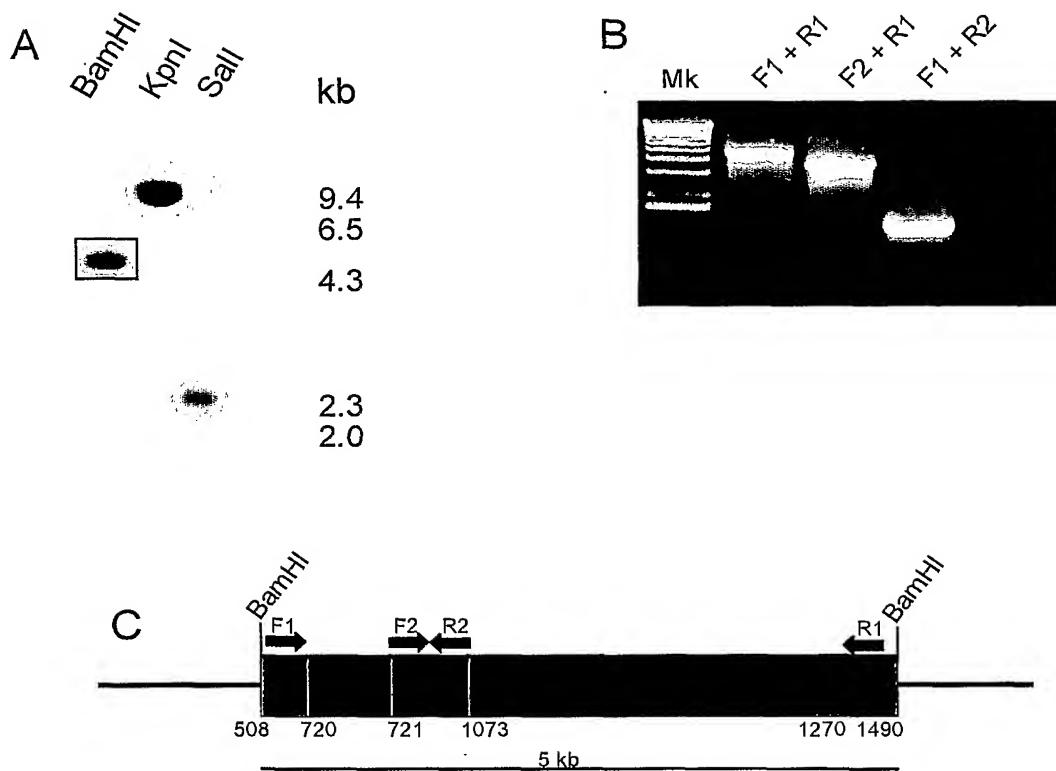


FIGURE 24

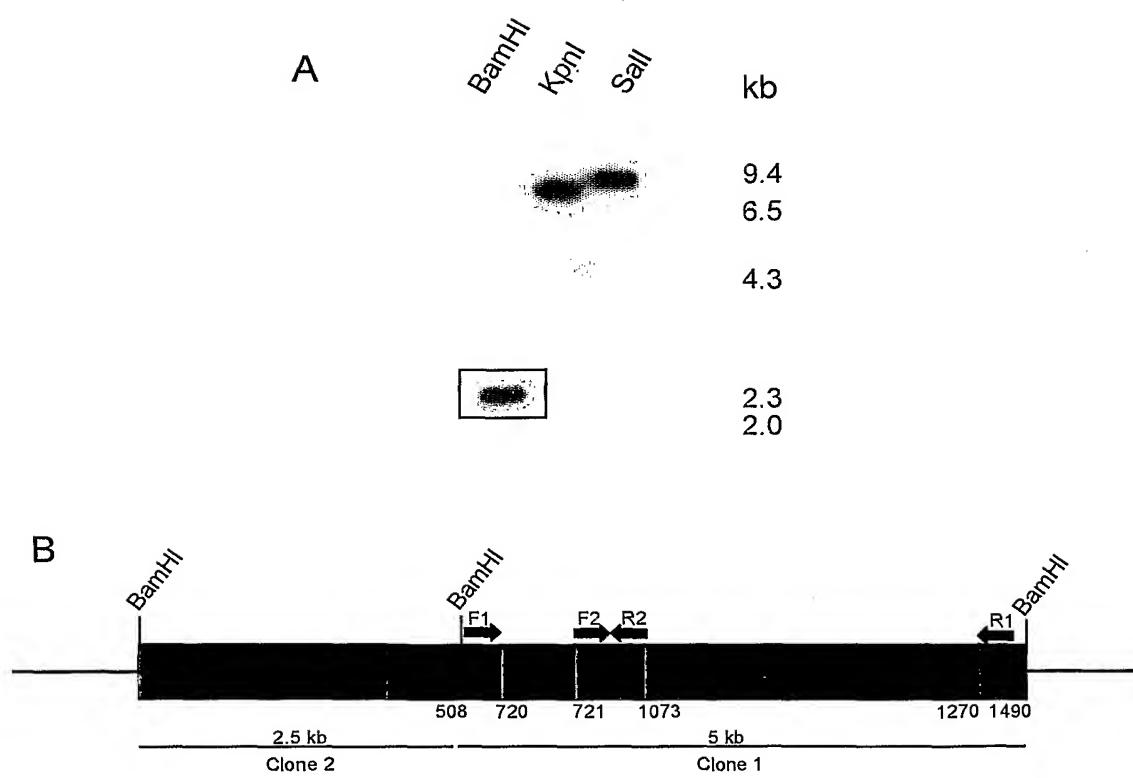
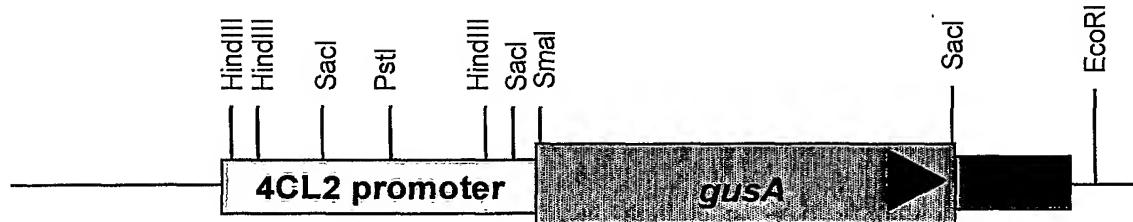


FIGURE 25



38/76

	TCCCGTATCTTCACAGTGACACCCCTACACTTCTGCTTGGAGATTACACAC	60
1	-----+-----+-----+-----+-----+-----+-----+	
	ACGGCAATTACCAAGGAGTATCTCCTAGATTATTTTTCGATAAGGATCTCCAGATAT	120
61	-----+-----+-----+-----+-----+-----+-----+	
	AGCATGTGAATCTCTGTACTACTACTGTTGTCAGCAAGAAAATTAAACATTGACATCAGTGT	180
121	-----+-----+-----+-----+-----+-----+-----+	
	TTTGTTGGGGCAGCGGAATCTTGACGCCCTTCTGCCTCTCAAGACATGTCACCCCT	240
181	-----+-----+-----+-----+-----+-----+-----+	
	CACTAGTTAGTGTGCCAGCTGGTAGTACTACGTACGATGCTCCCTCCCTCCGTAATTATT	300
241	-----+-----+-----+-----+-----+-----+-----+	
	CAACCTTTTGCTCTCTTTTATAAAGTCAAACCTTTAAATCTGACCAGATATCTGC	360
301	-----+-----+-----+-----+-----+-----+-----+	
	TAAAAAATTAGCAGACATGCATACATCAAAGCAGTAGTCCTCCCTCCGTTAAAATTACC	420
361	-----+-----+-----+-----+-----+-----+-----+	
	TGGTTTATTCAAATAAAGTCAAACTCTGTAATTCAATTAAATATTAGAAAATCTA	480
421	-----+-----+-----+-----+-----+-----+-----+	
	ACAGCACCTGTAGTATAAAAGTATGCTCCCTCTGTTGTAAGCTAACACTTTT	540
481	-----+-----+-----+-----+-----+-----+-----+	
	TGAGATACGGATAAATCTTAGCTAAACATGTCTATACCTTTGTATCTAGATAAAGT	600
541	-----+-----+-----+-----+-----+-----+-----+	
	TGGAAAGCTTTTAGAAACAGACAAAGTATGTTGACATTATGAATGTTGAGTATT	660
601	-----+-----+-----+-----+-----+-----+-----+	
	TTCCCTCTAACTTGATCAAATTACAAATTGCTTGAATAGAGGGACCATTATTAGT	720
661	-----+-----+-----+-----+-----+-----+-----+	
	ATGAAAATACATAATTGTAAAACACTCAACATAATTACGTGGTCAGTGATAGCAC	780
721	-----+-----+-----+-----+-----+-----+-----+	
	TAACCTAGCTTTCTAAATGCCACTGCTTTCAATAGAGCATGAAGCAGGACAATTAA	840
781	-----+-----+-----+-----+-----+-----+-----+	
	TTCGTGTGACTTGAATAGAGGGAGCCTGTTGGTCAACTCACCCGCATGTGTCTT	900
841	-----+-----+-----+-----+-----+-----+-----+	
	CATCCCCCTTGCTCTTCTATCTGTGGTGTCAATTGAGTGTCCCACGTGCATGTGGCGA	960
901	-----+-----+-----+-----+-----+-----+-----+	

FIGURE 26

961	AACTTGAACTAGAAATTGACATGCTCCACTGCCGGAGCGGAGTATCTTGTGCTTG -----+-----+-----+-----+-----+-----+-----+	1020
1021	TTACCCATTATTGTTGCTACGTACTACAGTGGTAGATTGAACTTCATAATCAAAGAAC -----+-----+-----+-----+-----+-----+-----+	1080
1081	TTAGTTCTACAATTTTGCTAAGCAATATAATGAGCAATCAAACCTCTATATCTGTG -----+-----+-----+-----+-----+-----+-----+	1140
1141	GCAAATAACTAACCTTATAGTTACAGTTAGATGCAGACGCCAGTGTCTTCCCCCTT -----+-----+-----+-----+-----+-----+-----+	1200
1201	TTCGGAAAAAGCTATTCCATAATAAGTGGAAATTAAATAATGGGTACTACGAATT -----+-----+-----+-----+-----+-----+-----+	1260
1261	TGAAAAAAAAGTGTCAAAAATTCACTAAGAAAGTACGTAGTACAAATTAAACTAAGAT -----+-----+-----+-----+-----+-----+-----+	1320
1321	TCCGACACTTATTAGGATCGGAGAGAGTAAGTAGCAAACACTACTCCATCCACCTAAAA -----+-----+-----+-----+-----+-----+-----+	1380
1381	CACGTGATTAACTTTGTCTAGATACGGATAGAAAGTTGGGATACATCCGTATCTTAAAA -----+-----+-----+-----+-----+-----+-----+	1440
1441	AAAAACGCACTTATTAGACGAAGGAGGGAGTATTCAACCTTGATTAAACGGAATC -----+-----+-----+-----+-----+-----+-----+	1500
1501	TACAAAGGAAATACATGGATTGTACAAGTGGCTGACCGTATCCATTATGTACTCGTACT -----+-----+-----+-----+-----+-----+-----+	1560
1561	TTGCAGTTGAAAGCAAAGGCTAGTGTAAATTGTAGGTGGTCTAGCGTCTAGCTGTT -----+-----+-----+-----+-----+-----+-----+	1620
1621	CATGGCGTTATCACAGCCGTGCCAGTGTGCTCAGGGCCGTACATAAGTTGCTTGGTGTAT -----+-----+-----+-----+-----+-----+-----+	1680
1681	GTGTCGATCTAGGATTGCGTCTTACAATTGCTTCCAATTATTCTGTAAAGAG -----+-----+-----+-----+-----+-----+-----+	1740
1741	ATCGATGTGAACTTCTCTGCGAGTAAACTGAAATTGTCTGAATAATATAACTCGGCAG -----+-----+-----+-----+-----+-----+-----+	1800
1801	ATTATGTTTATCGTTGCGTAACAGGCTACACAAATTGCTCGAGTCAGCAGCGAG -----+-----+-----+-----+-----+-----+-----+	1860
1861	TTGAGCTACAACGAATCCATCAGAAAAACTATACTATAGTAGCACATCGTTCTT -----+-----+-----+-----+-----+-----+-----+	1920

FIGURE 26 CONTINUED

40/76

1921	TTTCATGACGTTCTGTTCTTCTTAACCTTCCAGGAGCACGGAGACGACGATGTGGTG -----+-----+-----+-----+-----+-----+ R S T G D D D V V	1980
1981	ATAAAGATTTGTAUTGCCACTCTGACCTGCACGCCCTGAAGAACGACTGG -----+-----+-----+-----+-----+ I K I L Y C G I C H S D L H A L K N D W	2040
2041	AAGAACTCAAGGTACCGATGATCCCCGGCAGCAGATCGCCGGCAGGTACGGAGGTG -----+-----+-----+-----+-----+ K N S R Y P M I P G H E I A G E V T E V	2100
2101	GGCAAGAACGTGAGCAAGTTCAAGGCCGGCAGCGCTGGCGTCGGTGCATGGTGAAC -----+-----+-----+-----+-----+ G K N V S K F K A G D R V G V G C M V N	2160
2161	TCGTGCCGGTCGTGCGAGAGCTGCGACAAGGGCTTCGAGAACCATGCCGGCATGATC -----+-----+-----+-----+-----+ S C R S C E S C D K G F E N H C P G M I	2220
2221	CTCACCTACAACCTGGTCGACGTCGACGGCACCGTCACCTACGGGGCTACTCCAGCATG -----+-----+-----+-----+-----+ L T Y N S V D V D G T V T Y G G Y S S M	2280
2281	GTGGTGGTGCACGAGCGGTCTGGTCCGGTTCCCGACGCCATGCCGCTGGACAAGGG -----+-----+-----+-----+-----+ V V V H E R F V V R F P D A M P L D K G	2340
2341	GCGCCGCTGCTGTGCGCCGGCATCACCGTGTACAGCCCCATGAAGTACCACGGCTAAC -----+-----+-----+-----+-----+ A P L L C A G I T V Y S P M K Y H G L N	2400
2401	GTTCCCGGGCTGCACCTCGCGTGCTGGGCTGGCGGGCTGGCCACGTTGCGGTCAAG -----+-----+-----+-----+-----+ V P G L H L G V L G L G G L G H V A V K	2460
2461	TTCGGCAAGGCCCTCGGAATGAAAGTGACGGTGATCAGCTCGCCGGGAAGAAGGAG -----+-----+-----+-----+-----+ F G K A F G M K V T V I S S S P G K K E	2520
2521	GAGGCCCTGGGGCGGCTGGCGCCGACCGTTCATCGTCAGCAAGGACGCCGACGAGATG -----+-----+-----+-----+-----+ E A L G R L G A D A F I V S K D A D E M	2580
2581	AAGGTAGGCGGACCCGCTGGTTCAAGTTACTTCCCTGTCCGGTGCAGAAGAAAGAGGAA -----+-----+-----+-----+-----+ K	2640

FIGURE 26 CONTINUED

G at 851 bp (coding sequence) missing from cDNA in cv Ellett

	▼	
2641	CTTGAGGGTTCATGTTGTTGCGTTGGTATGTCTTGCAAGGCTGTGATGAGCACCAT	2700
	A V M S T M	
2701	GGATGGCATCATAAACACGGTATCTGCAAACATCCCCCTGACCCCTCTTCGGGCTGCT	2760
	D G I I N T V S A N I P L T P L F G' L L	
2761	CAAGCCCAACGGCAAGATGATCATGGTCGGCCTCCCCGAGAACGCCATCGAGATTCCCTCC	2820
	K P N G K M I M V G L P E K P I E I P P	
2821	CTTCGCTCTAGTTGCCAGTAAGTCTTAGGATCTCTTGCAATAAGGAGAAATCATGCACTG	2880
	F A L V A	
2881	ATCGATCAGAGAAATGAGATAGCATCCTGATGAACATTGTACGTGTGCAGCGAATAAG	2940
	N K	
2941	ACCCCTGGCCGGGAGCATCATCGGCGGATGAGCGACACGCAGGAGATGCTGGACCTCGCG	3000
	T L A G S I I G G M S D T Q E M L D L A	
3001	GCGAAGCACGGCGTGACGGCGACATCGAGGTGGTCGGCGGGAGTATGTAAACACGGCC	3060
	A K H G V T A D I E V V G A E Y V N T A	
3061	TTGGAGCGCCTTGCCAAGAACGACGTCAGGTATCGCTTCGTATCGACATCGAACACACC	3120
	L E R L A K N D V R Y R F V I D I G N T	
3121	CTCGACAAGGTTGCGGCCACCACCGAGTGAAACGTACTCAGCACTGCTTACGATCTACGTT	3180
	L D K V A A T T E *	
3181	GTTCCACTGTTAGTGCTCCGTAGTAAACAATAACGATCAAACCTTTGTCATCTGGTGC	3240
3241	ATTGGTGTAGACATGGTTGGTGGAGGAAACTGAGTTGAAGGATGGATGGATAAGTTG	3300
3301	CTTCTTGCCTGTTAATGGATTACCTACTTAGCTCACTGCAATTAAACAAATTAAAGAAC	3360
3361	GACACACCCAAAAGACTTTCGTCAGTTTCTGGATTATACAAGTCGTTATGGTTGGGTG	3420

FIGURE 26 CONTINUED

3421	TCAGTGTGTCACAGATAATCATACTATGGTATTTAACCTGGAAGATCGTTTTGGCGG -----+-----+-----+-----+-----+-----+-----+	3480
3481	CAACTCAGTGGGTTTCCCACATGTAATTATAAAATTCAACAAGTCATGAGGTACA -----+-----+-----+-----+-----+-----+-----+	3540
3541	AAGGGTTGTTGCTAGAGGATAGCAACAAAGAAGCTAGCCAAAAGATCATAGGCTAAAAAA -----+-----+-----+-----+-----+-----+-----+	3600
3601	GAGAGAAAAGAAAACAAAAGCTATAGTTATCGAAATCTCTCAGCTCAAATTTAAAAAC -----+-----+-----+-----+-----+-----+-----+	3660
3661	CAGCATAAGACTTTCTAGAACGCCTTATGAACAAGAAGAGCTAGCTCATCTTAAACCTTT -----+-----+-----+-----+-----+-----+-----+	3720
3721	TCCTGCATCTGAAAGATTGAGGGTGCAACCCTGAATATAAAATCATTCTGTCATCCA -----+-----+-----+-----+-----+-----+-----+	3780
3781	GATAGACTATGAGTCAAAATAGTCATTCCATGAAGAAGGGCACTTTAATACATTTT -----+-----+-----+-----+-----+-----+-----+	3840
3841	GAGACTTGGTATGATACTCTGAATGTCAACACCCCTGGAAGATCTTTCACTCCTATGGAA -----+-----+-----+-----+-----+-----+-----+	3900
3901	GGACAAGAAAGCATTCAACTCCTTTACTAAGGAAGAGATTGACAAGGTGATTGAGAGA -----+-----+-----+-----+-----+-----+-----+	3960
3961	ATTCCTTAGACACTATAGAAAGTCACAAGGTGCCAACGGCGAACCTGTGCCGACGGC -----+-----+-----+-----+-----+-----+-----+	4020
4021	TTTTTATCGGGGAAGCCAGCATCGTACCGAGACCGGCAGCCCACCAACTAGGCCGTCGG -----+-----+-----+-----+-----+-----+-----+	4080
4081	CACACATCCTCCAGTGTGGCGGCCAACATCGGCATAAGTTGGCCGTTGGGCATCAACT -----+-----+-----+-----+-----+-----+-----+	4140
4141	CCCCCGTCGGAACAGGTCTAGCGCATGGACCGTCGTATGGGGCGAACGACGTCATC -----+-----+-----+-----+-----+-----+-----+	4200
4201	CTATGCCGACGGCCTAGCCGCGGCCTAGCTTGCCAGCGCTATGCCGACGTCACATTGCC -----+-----+-----+-----+-----+-----+-----+	4260
4261	ATCGGCACATGCTAGTTTTTTCTTTTCTACATGCCAATTGTATATGTATATATA -----+-----+-----+-----+-----+-----+-----+	4320
4321	CTCATTTACTTATTACTCCAATTATTTAATGTGTATATATTGCTCACCAATTGTAC -----+-----+-----+-----+-----+-----+-----+	4380

FIGURE 26 CONTINUED

4381 GAATTTGTAACCTCCGAGAAATTGCTAAAATGATGGAGTGACCTACAACGAGCCTGGAT 4440
4441 ATGTGAGTTCTTCTTGCCCCATTGCACAAAAATTGTAAATATTAGGGTTACTGGATCCA 4500
A) CTAGTTCTAGAGCGGCCACCAGCGGGGAGCTCCAGCTTTGTTCCCTTAGTA 4555

FIGURE 26 CONTINUED

44/76

<pre> GGCACGAGTCGCCTCCAACGTCTTCCCTAACCGGCCGTCCCTACGCTTGACACCACC 1 -----+-----+-----+-----+-----+-----+-----+ +-----+-----+-----+-----+-----+-----+ ACGCACAGACAGAGCAGTTCCCAGCCCCGCCGGAACCGGATGGCACCCACGGCGGG 61 -----+-----+-----+-----+-----+-----+-----+ M A P T A A E +-----+-----+-----+-----+-----+ AGCAGACGGAGCACCACCAAGCACACCAGGAAGGCAGTGGGCTGGCGCGCGACGACG 121 -----+-----+-----+-----+-----+-----+-----+ Q T E H H Q H T R K A V G L A A R D D A +-----+-----+-----+-----+-----+ CCGGCCACCTCTCCCCGCTCGCCATCACACGGAGGAGCACAGGAGACGACGATGTGGTGA 181 -----+-----+-----+-----+-----+-----+-----+ G H L S P L A I T R R S T G D D D D V V I +-----+-----+-----+-----+-----+ TAAAGATTTGTACTCGGAAATCTGCCACTCTGACCTGCACGCCCTGAAGAACGACTGGA 241 -----+-----+-----+-----+-----+-----+-----+ K I L Y C G I C H S D L H A L K N D W K +-----+-----+-----+-----+-----+ AGAACTCAAGGTACCCGATGATCCCCGGCACGAGATCGCCGGCGAGGTACGGAGGTGG 301 -----+-----+-----+-----+-----+-----+-----+ N S R Y P M I P G H E I A G E V T E V G +-----+-----+-----+-----+-----+ GCAAGAACGTGAGCAAGTTCAAGGCCGGCACCGCGTGGCGTCGGTGCATGGTGAAC 361 -----+-----+-----+-----+-----+-----+-----+ K N V S K F K A G D R V G V G C M V N S +-----+-----+-----+-----+ CGTGCCGGTGTGCGAGAGCTGCGACAAGGGCTTCGAGAACCACTGCCGGCATGGTACCC 421 -----+-----+-----+-----+-----+-----+-----+ C R S C E S C D K G F E N H C P G M I L +-----+-----+-----+-----+ TCACCTACAACACTGGTCGACGTCGACGGCACCGTCACCTACGGCGGCTACTCCAGCATGG 481 -----+-----+-----+-----+-----+-----+-----+ T Y N S V D V D G T V T Y G G Y S S M V +-----+-----+-----+-----+ TGGTGGTGCACGAGCGGTTCGTGGTCCGGTCCCGACGCCATGCCGTGGACAAGGGCG 541 -----+-----+-----+-----+-----+-----+-----+ V V H E R F V V R F P D A M P L D K G A +-----+-----+-----+-----+ CGCCGCTGCTGTGCGCCGGCATCACCGTGTACAGCCCCATGAAGTACCAAGGGCTCAACG 601 -----+-----+-----+-----+-----+-----+-----+ P L L C A G I T V Y S P M K Y H G L N V +-----+-----+-----+-----+ TTCCCGGGCTGCACCTCGGCGTGTGGGCTGGGGCTGGCGGGCTGGGCCACGTTGCGGTCAAGT 661 -----+-----+-----+-----+-----+-----+-----+ P G L H L G V L G L G G L G H V A V K F +-----+-----+-----+-----+ TCGGCAAGGCCTTCGGAATGAAAGTGACGGTGATCAGCTCGTCGCCGGGAAGAAGGAGG 721 -----+-----+-----+-----+-----+-----+-----+ G K A F G M K V T V I S S S P G K K E E +-----+-----+-----+-----+ </pre>	60 120 180 240 300 360 420 480 540 600 660 720 780
----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------	----------------------------------------------------------------------------------------

FIGURE 27

781 AGGCCCTGGGGCGGCTGGCGCCGACCGTTCATCGTCAGCAAGGACGCCGACGAGATGA
 840 A L G R L G A D A F I V S K D A D E M K
 G at 851 bp (coding sequence) missing from cDNA in cv Ellett
 ▼
 841 AGGCTGTGATGAGCACCATGGATGGCATCATAAACACGGTATCTGCAAACATCCCCCTGA
 900 A V M S T M D G I I N T V S A N I P L T
 960 CCCCTCTCTCGGGCTGCTCAAGCCAACGGCAAGATGATCATGGTCGGCCTCCCCGAGA
 961 P L F G L L K P N G K M I M V G L P E K
 1020 AGCCCATCGAGATTCCCTCCCTCGCTCTAGTTGCCACGAATAAGACCCCTGGCCGGAGCA
 1021 P I E I P P F A L V A T N K T L A G S I
 1080 TCATCGCGGCATGAGCGACACG CAGGAGATGCTGGACCTCGCGCGAACGACCGCGTGA
 1081 I G G M S D T Q E M L D L A A K H G V T
 1140 CGGCCGACATCGAGGTGGTCGGCGCGGAGTATGTGAACACGGCTTGGAGCGCCTGCCA
 1141 A D I E V V G A E Y V N T A L E R L A K
 1200 AGAACGACGTCAGGTATCGCTTCGTCATCGACATCGGAAACACCCCTGACAATGTTGCGG
 1201 N D V R Y R F V I D I G N T L D N V A A
 1260 CCACCACCGAGTGAACGTACTCAGCACTGCTTACGATCTACGTTGTTCCACTGTTAGTGC
 1261 T T E *
 1320 TCCGTAGTAAACAATAAACGATCAAAACTCTTGTATCTGGTGCATTGGTGTAGACATGG
 A) TTGTTTGCAGGAAACTGAGTTGAAGGATGGATGGATAAAAAAAAAAAAAAAA
 1378 GGCACAGAGTCGCCTCCAACGTCTTCCCTTAACCGGCCGTCCCTACGCCTTGCA
 1 G ACCACC
 60
 61 ACGCACAGACAGAGCAGTTCCAGCCCCGGCAACCGGATGGCACCCACGGCGCG
 120 M A P T A A E
 121 AGCAGACGGAGCACCAACCAGCACACCAGGAAGGGGGTGGGCTGGCGCGCGACGACG
 180 Q T E H H Q H T R K A V G L A A R D D A

FIGURE 27 CONTINUED

181 CCGGCCACCTCTCCCCGCTCGCCATCACACGGAGGAGCACAGGAGACGACGATGTGGTGA 240
 G H L S P L A I T R R S T G D D D V V I

 241 TAAAGATTTGTAUTGCAGGAATCTGCCACTCTGACCTGCACGCCCTGAAGAACGACTGGA 300
 K I L Y C G I C H S D L H A L K N D W K

 301 AGAACTCAAGGTACCCGATGATCCCCGGCACGAGATGCCGGCAGGTCACGGAGGTGG 360
 N S R Y P M I P G H E I A G E V T E V G

 361 GCAAGAACGTGAGCAAGTTCAAGGCCGGCACCGCGTGGCGTCGGGTGCATGGTGAAC 420
 K N V S K F K A G D R V G V G C M V N S

 421 CGTCCGGTCGTGCGAGAGCTGCGACAAGGGCTTCGAGAACCACCTGCCGGCATGATCC 480
 C R S C E S C D K G F E N H C P G M I L

 481 TCACCTACAACTCGGTCGACGTCGACGGCACCGTACCTACGGCGCTACTCCAGCATGG 540
 T Y N S V D V D G T V T Y G G Y S S M V

 541 TGGTGGTGCACGAGCGGTTCTGGTCCGGTCCCGACGCCATGCCGTGGACAAGGGCG 600
 V V H E R F V V R F P D A M P L D K G A

 601 CGCCGCTGCTGTGCGCCGGCATCACCGTGACAGCCCCATGAAGTACCAACGGGCTCAACG 660
 P L L C A G I T V Y S P M K Y H G L N V

 661 TTCCCGGGCTGCACCTCGGCGTGCTGGGGCTGGGGCTGGGCCACGTTGCGGTCAAGT 720
 P G L H L G V L G L G G L G H V A V K F

 721 TCGGCAAGGCCTCGGAATGAAAGTGACGGTATCAGCTCGCCGGGAAGAACGGAGG 780
 G K A F G M K V T V I S S S P G K K E E

 781 AGGCCCTGGGGCGGCTGGCGCCGACCGTTCATCGTCAGCAAGGACGCCACGAGATGA 840
 A L G R L G A D A F I V S K D A D E M K

 G missing at 851 bp in the cDNA isolated from cv Ellett
 resulted in a premature stop codon (truncated CAD2)

 841 AGGCTGTGATAGCACCATTGGATGGCATATAAACACGGTATCTGCAAACATCCCCCTGAC 900
 A V I A P W M A S *

FIGURE 27 CONTINUED

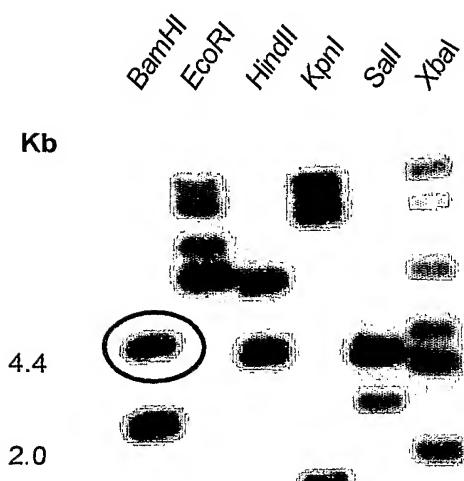
47/76

901	CCCTCTTCTGGGCTGCTCAAGCCAAACGGCAAGATGATCATGGTCGGCTCCCCGAGAA -----+-----+-----+-----+-----+-----+-----+	960
961	GCCCATCGAGATTCCCTCCCTCGCTCTAGTTGCCACGAATAAGACCCTGGCCGGAGCAT -----+-----+-----+-----+-----+-----+-----+	1020
1021	CATCGGCGGCATGAGCGACACGCAGGAGATGCTGGACCTCGCGCGAAGCACGGCGTGAC -----+-----+-----+-----+-----+-----+-----+	1080
1081	GGCCGACATCGAGGTGGTCCGCGGGAGTATGTGAACACGGCCTTGGAGGCCCTGCCAA -----+-----+-----+-----+-----+-----+-----+	1140
1141	GAACCGACGTCAAGGTATCGCTTCGTCATCGACATCGCAACACCCCTCGACAATGTTGCC -----+-----+-----+-----+-----+-----+-----+	1200
1201	CACCAACCGAGTGAACGTACTCAGCACTGCTTACGATCTACGTTGTTCCACTGTTAGTGCT -----+-----+-----+-----+-----+-----+-----+	1260
1261	CCGTAGTAAACAATAAACGATCAAACCTTGTCACTGGTGCATTGGTAGACATGGT -----+-----+-----+-----+-----+-----+-----+	1320
A)	TGTTTGCAGGAAACTGAGTTGAAGGATGGATGGATAAAAAAAAAAAAAAAA -----+-----+-----+-----+-----+-----+-----+	1377

FIGURE 27 CONTINUED

FIGURE 28

A



B

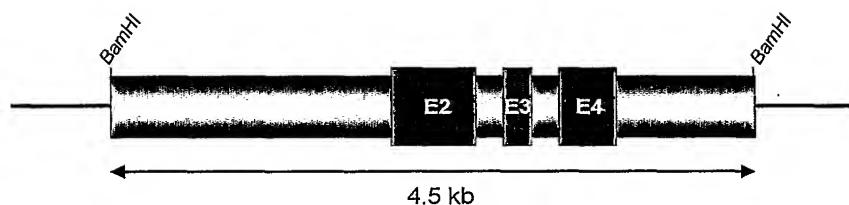
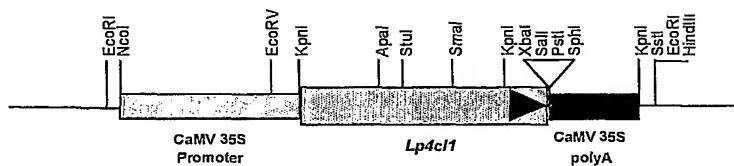


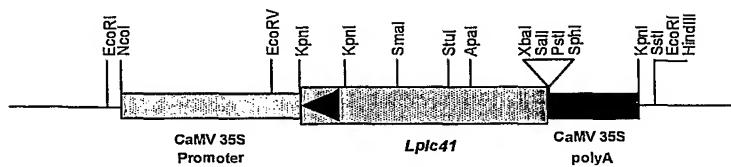
FIGURE 29A

A)

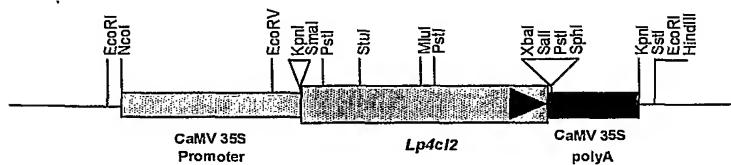
p35S4cl1



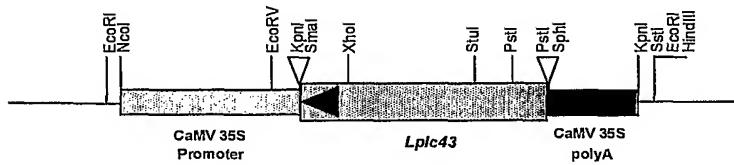
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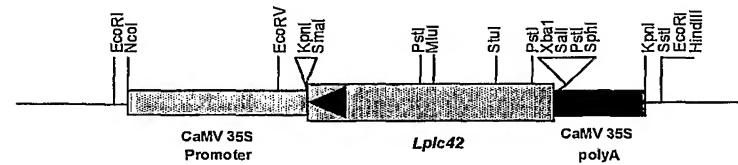
p35S4cl2



p35S4cl3



p35S4cl42



p35S4cl3

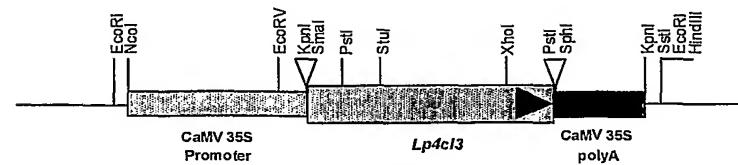


FIGURE 29B

B)

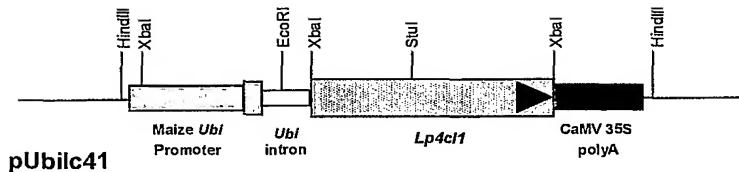
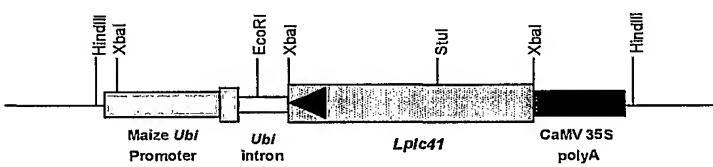
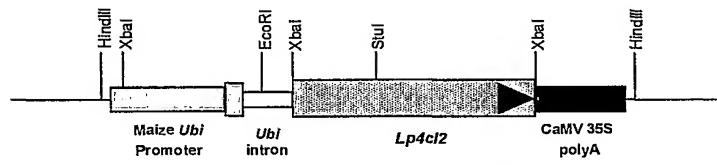
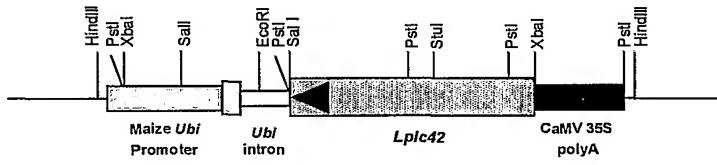
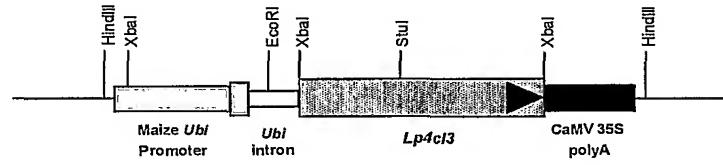
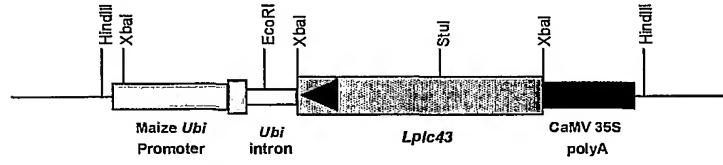
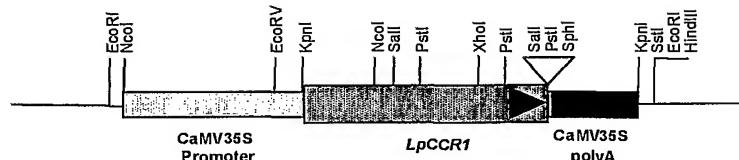
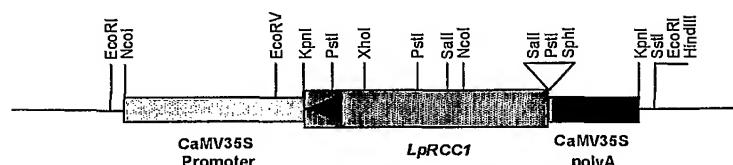
pUbi4cl1**pUbilc41****pUbi4cl2****pUbilc42****pUbi4cl3****pUbilc43**

FIGURE 30

A)

p35SCCR1**p35SRCC1**

B)

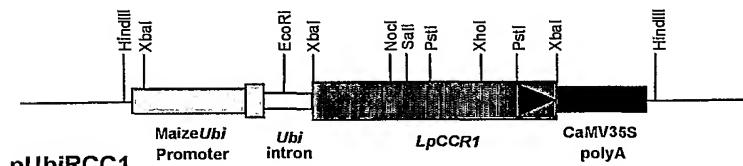
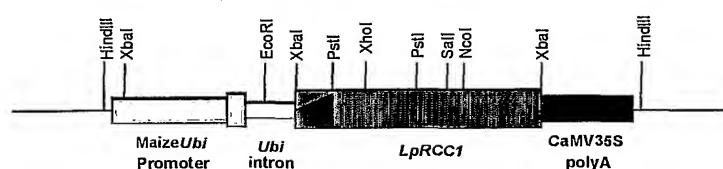
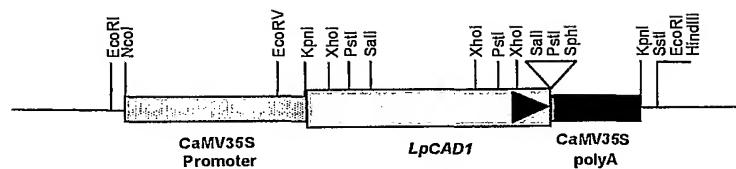
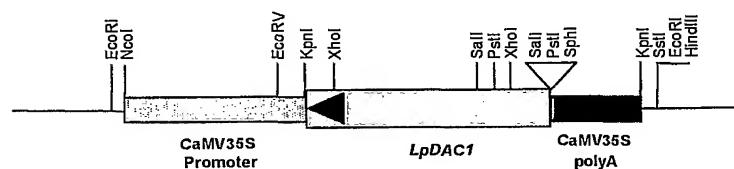
pUbiCCR1**pUbiRCC1**

FIGURE 31

A)

p35SCAD1**p35SDAC1**

B)

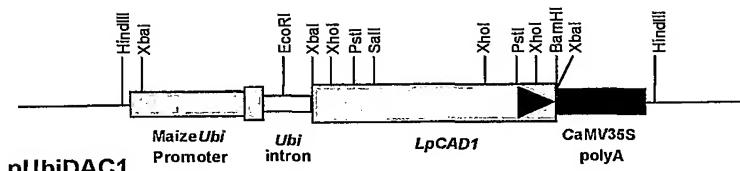
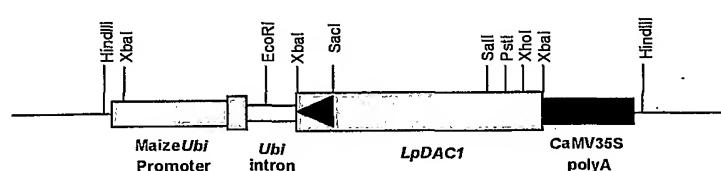
pUbiCAD1**pUbiDAC1**

FIGURE 32

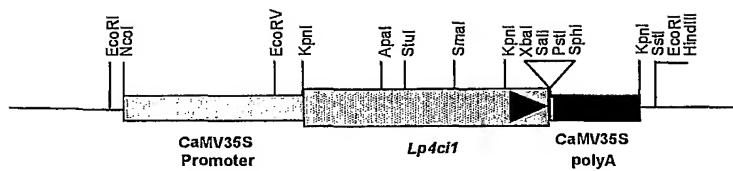
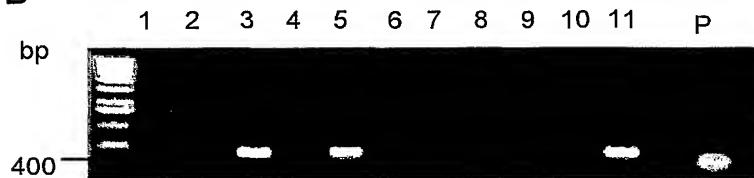
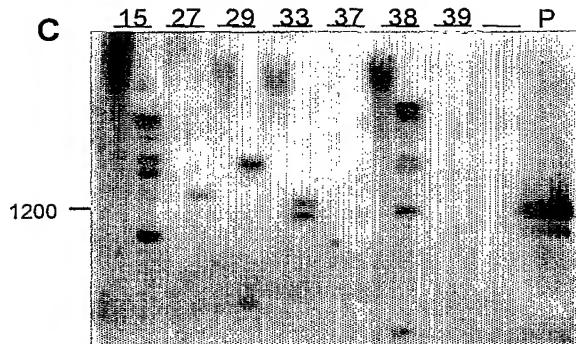
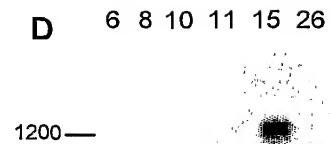
A**B****C****D**

FIGURE 33

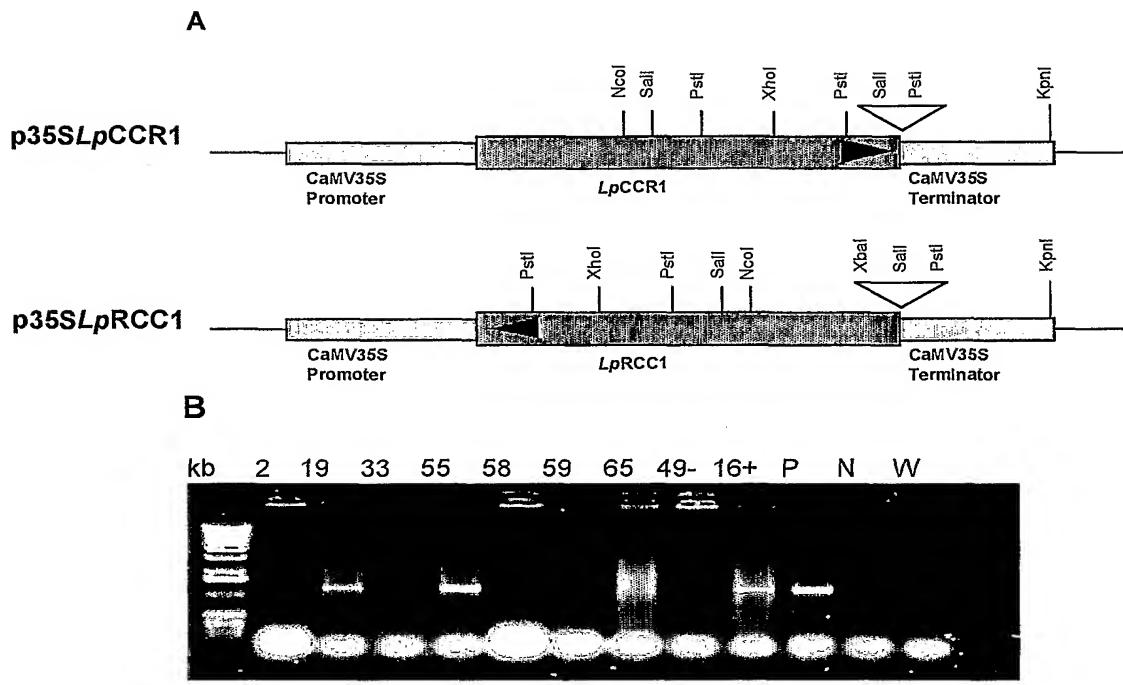
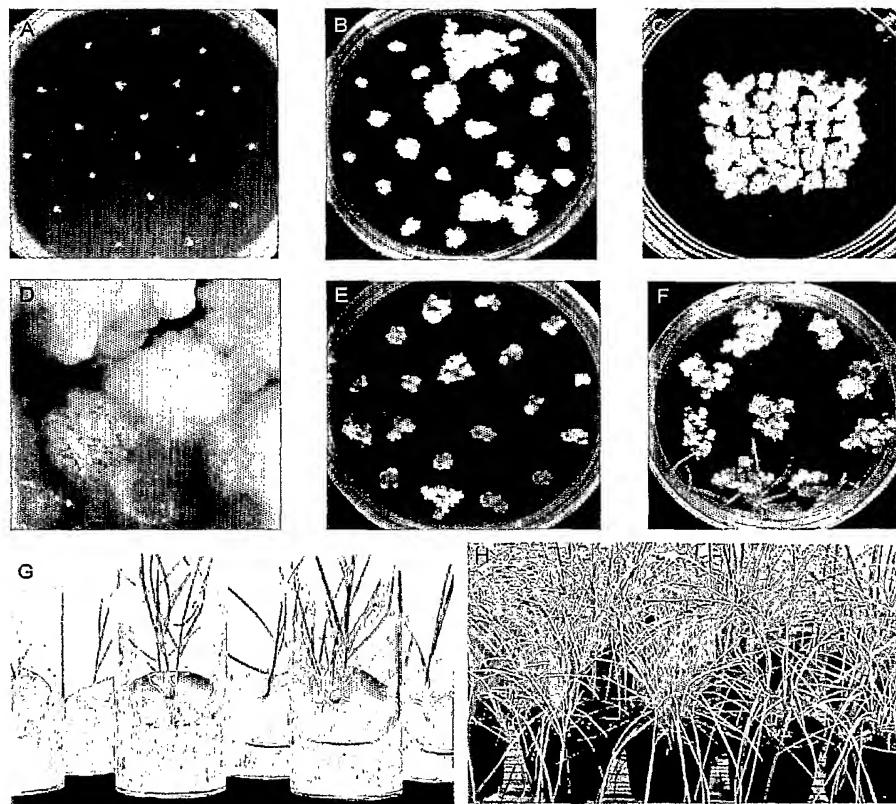


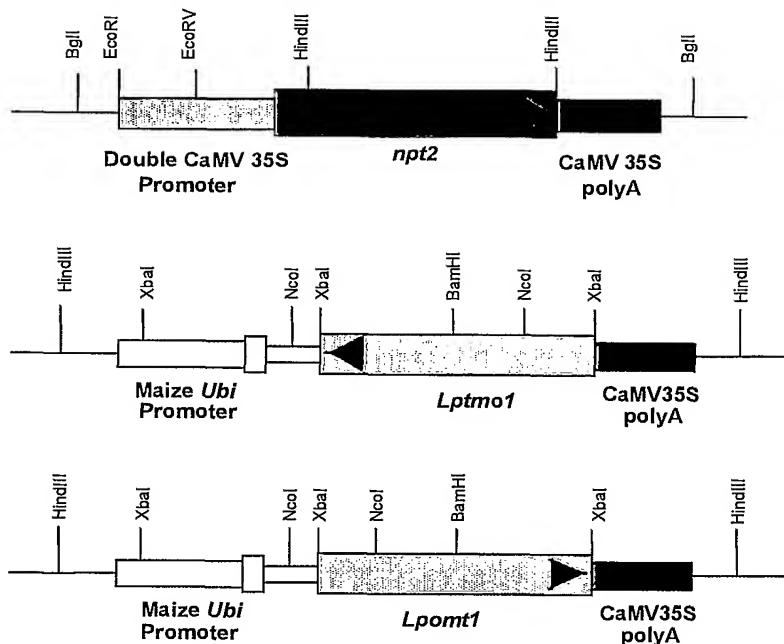
FIGURE 34



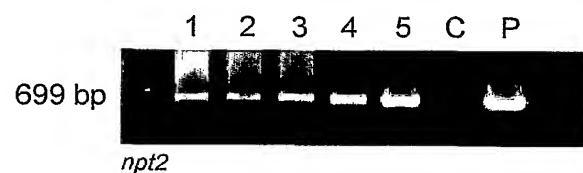
56 / 76

FIGURE 35

A



B



57/76

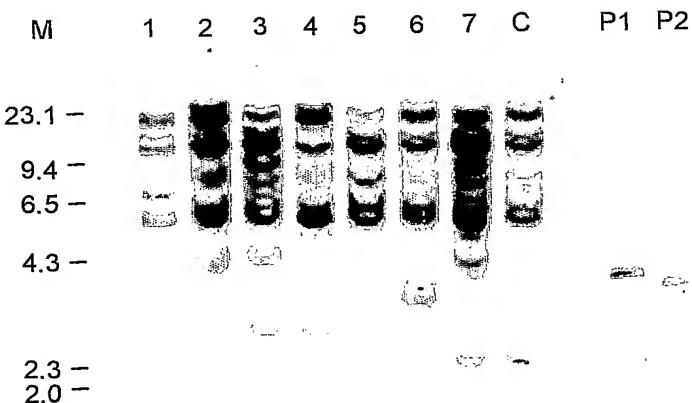
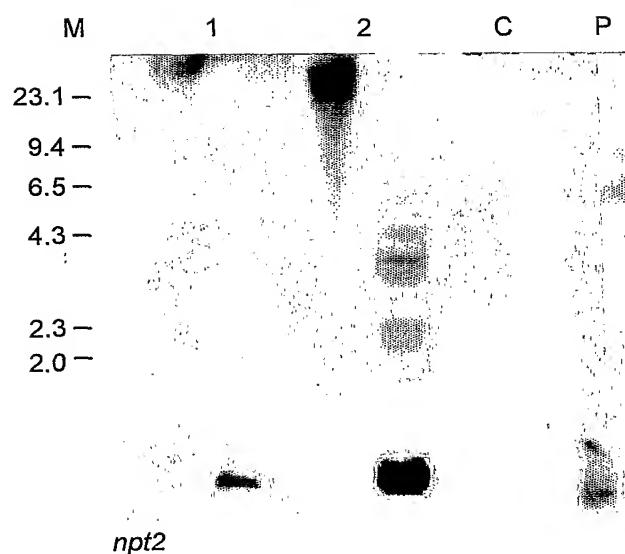
C**D**

FIGURE 35
CONTINUED

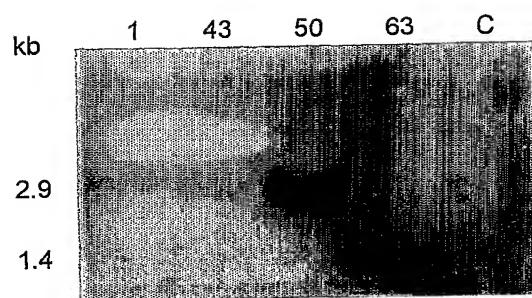
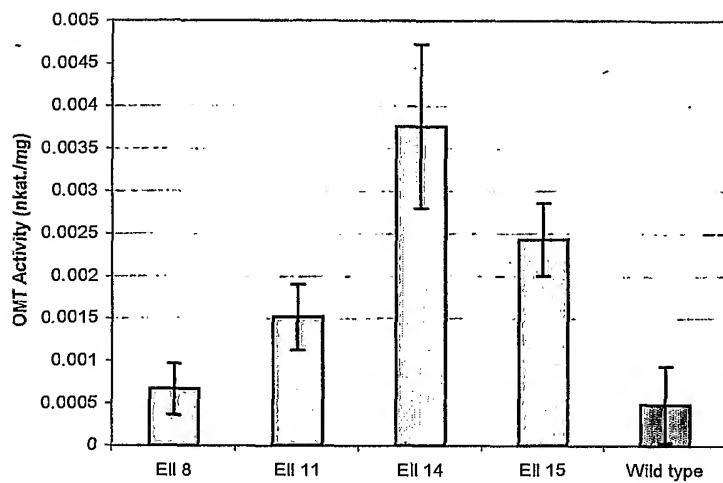
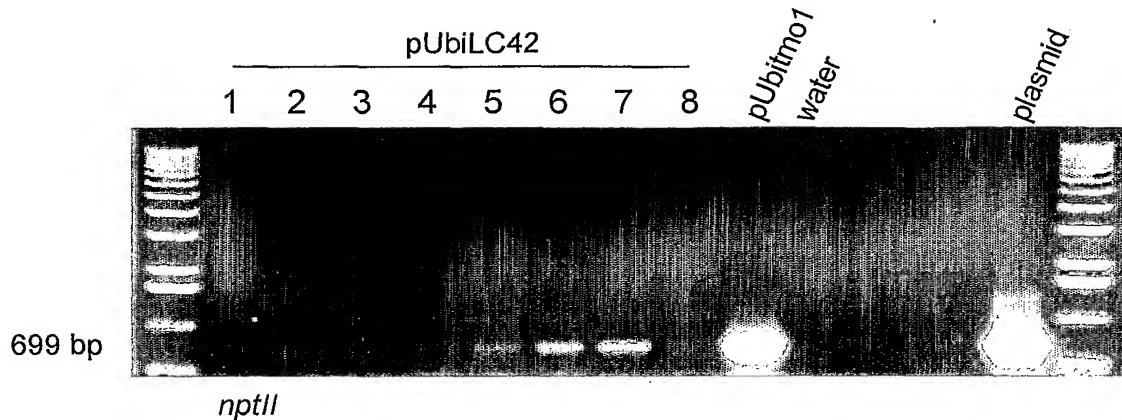
E

FIGURE 36



59/76

FIGURE 37



60/76

-2206	CGGGATCAACTGGATGTCCTTGCGGGCACGGTTCAGGAACAAACGACACATGCAGCAG	-2147
-2146	GGATCTCCTCCAAAGACTCACACAAAGGTGACATGAGCGCCCGCTTTTGAAGCCAAGT	-2087
-2086	TGGCTAAGAAAATCGCAAAGCTTGGTAGTCGGCACCTCAGGATCTGCAACAAAAGGCA	-2027
-2026	CCAAGGGAGCTGCCAACACATCAACCACAACATCATGTTCAAACGCAGTCTCCTCAAGCC	-1967
-1966	TCGAATGCTCAACCGAAAGAGAGGGCAGAAGCTCAACAAAAAAACTCAGCCAACCCAAAGC	-1907
-1906	CCTCGACGTCATCAGAGATTAGGCTCTGAGGACCCGCAGGGAAAGCAACCTGTCAACAAAC	-1847
-1846	CGCATCCGGCAGAAAAGGAGCAAGACCGGAGCAACCCCTCAAGAGGCACACGAAAGACGTC	-1787
-1786	GAAGCCAAGAGGAGACGAGTCGCAGGGACGGCGGACAGGCAGAGAAGGGGCCGTAGAACTC	-1727
-1726	CAAGAGCTGGCGTCCCTCGACCTAGCATCCGAAGCACTGACCGGGGCACTCAATGCATA	-1667
-1666	ACTTTATCTTGATGGCATATGTACTCAAACCCATAACATGTTACCATGCATTATCTATG	-1607
-1606	GAACATTCTCATATAACAATTCTGAGTGGTCAGTGCATAGGAATTTCATTAAACACC	-1547
-1546	AAAAACATAACTGGGGCCTACACACACTTCACAGCATGGAAAATTGTTAGCTTTTAA	-1487
-1486	AGAGTTGCAAAATCTGTCAAGCGAATGTTCTTGATAATTGGAACGAAGCATGTTCCC	-1427
-1426	CATTTCAATGTGTGTCCTTACCCCTAACTAGCACCCGACCAACAAATCTGACCACCT	-1367

FIGURE 38

61/76

-1366	AGTTATATCATCATAGAGACCCACATGTAGGTTGACCCCCATAAACACTTGTGGATATC	-1307
-1306	ATGGAAAATGGCCTTGATCAACACTTCCTACTTGGTACAAATGGTTATGGACTTA	-1247
-1246	CTCAATTAGTGCTTAGAGAGCTTGGCTGCAGACTTGTAGCTTCCAATATTCATAGG	-1187
-1186	TCCCTCCGGAGTGGGCAGCCCCATCTACATAGGCTCAAAACCAGATTTGTAAACATGTT	-1127
-1126	AGACACTTTCAACTTCATCATAGACCATCAAGGAGCTGGCATGTGACAGTGATATATGTA	-1067
-1066	TCAATTACCCATTCAACACGAATAGCTTGCTCATGCATGGTTAGTCTTGC GGCGGGGG	-1007
-1006	CGGGACCATCGAACACACCGCCGGCGGTCA GTAGGCTAGGGTTAGATAAAATCTAGCCG	-947
-946	TTTCATTCAAACTTGTGATATATAATCAAATTAAATAAAACCTTTATTCGTGCAT	-887
-886	TTTTATTTATTTGAGGGCGTGTGGGGACACGGCTGGAAAGTGACATCCCCAACACT	-827
-826	GCACGAAGAAAACCGCTCGCCAAAAATT CGATCCGGCGTCAGTCCTTGGAGACGATT	-767
-766	TGGATGACGCGGCTAGAGATGCTCTAAGTTCTCACGCCATGTTCTTCTATATACA	-707
-706	CACAGCCCCAAGGTCCATGAAAAGTAAAACGGCACGACGACACGCACCGCGACAACCTCA	-647
-646	CATTACGGCACATCGTATTACGGACCACATACAACCTCCACCGCTATTCTCAGCCAAGTC	-587
-586	ATACATGACATGATCCAATGGACGACTTGTGAGCGAAACTAGAACCTTGC GGTTTAG	-527
-526	ATTTTCCAATGTGGATAAGTTGTACCGCGGACTAGCTTACACTTGGTTGAAAAAAGCT	-467

FIGURE 38 CONTINUED

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<211> Length : 12175
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SequenceDescription :

Custom Codon

Sequence Name : CCR1genomicseq (SEQ ID NO: 18)

62/76

-466	TATTGTAGCACGACTTCTCACTGACATAGGAATGTAAACAGTCTCTCCACGCCATGTTTC	-407
-406	TTTCTAGTAGTAGCATACTAGTAGTAACCTCTTGTCTACACACACCCAGGGTCAA	-347
-346	GAAAGGAAAACGGCACGACGGCACCCACCGACGACGACTCCACATCACGGTCGTA	-287
-286	AAAAAAAGTCAAAACTCGCTGACGTGGCACCACCGTGCAGTCAGTCAGTCAGTGACGCGCTCCCT	-227
-226	GCGCAGGTYTCACTtCAAGTTTACCTACCACTGTGGGCCACCGCCAaTGTGGGCCCG	-167
-166	CGAGCTtCTtACTCACTGACCTGTCTCCCACCAGCCTCCTGCCGGTATATTACCCGGC	-107
-106	CCCCAATTCTCTGCCTTCCCACGAGCAGCAGCCGGAGCACGGAATCCGGCCATT	-47
-46	CCTCCACCTTCAGCTCCGCCAAAGATTCCATCCGGCGAGATCCATGGCTCCATCGCG	13
14	GC GG AC CG GC CT CC CG CG AG CT GG GT TT CC GG T C CA AG CT CC GG AC AT CG AG AT CCC G A D A P P A E L V F R S K L P D I E I P	73
74	ACCCACCTGACGCTGCAGGACTACTGCTTCCAGGCCCTGCCGGAGCTCTCGCGCGGCC T H L T L Q D Y C F Q R L P E L S A R A	133
134	TGCCTCATCGACGGGCCACGGCGCCGCTCACCTACGCCGACGTGGACGCCCTCACG C L I D G A T G A A L T Y A D V D A L T	193
194	CGCCGCTGCCGCCGGCTCCGCCCTGGGGTCCGCAAGGGCGACGTGTCATGGCG R R C A A G L R R L G V R K G D V V M A	253
254	CTGCTCCGCAACTGCCCGAGTTGCCCTCGTGTTCCTCGGCCGCCGGCTCGGCC L L R N C P E F A F V F L G A A R L G A	313

FIGURE 38 CONTINUED

63/76

314 GCCACCACCAACCGCCAACCCGTTCTACACGCCAACGAGATCCACCGCCAGGCGACCGCC 373
-----+-----+-----+-----+-----+-----+-----+-----+
A T T T A N P F Y T P H E I H R Q A T A

374 GCCGGGGCCAGGGTCATCGTCACCGAGGCCTGCCCGTCGAGAAGGTGCCGCCCTCGCC 433
-----+-----+-----+-----+-----+-----+-----+
A G A R V I V T E A C A V E K V R A F A

434 GCCGAGAGAG 443
-----+---
A E R

FIGURE 38 CONTINUED

64/76

-6735	TCGACGGCCCGTAATACGACTCACTATAGGGGAAGAATTGGATCATATGGATTG	-6676
-6675	ACACTGGAATTACTCCCATCGGGAGCGTGCAAACAAAAAGGTGTTAGCAAGAACAA	-6616
-6615	CTGGCACATTGCCAGCACAGAATTGTTACAATCATAGAAAGTTTATGACAGGACATT	-6556
-6555	GTTTCAACCGAAAGCAAGATTACAACAATATAATCAAGGGCTGGGTCTGGTGGACATG	-6496
-6495	CTCGGTCCAATGGACGATTATTGCCGAGACCAGCTCAAGGAGTTGACGAGCACACTTA	-6436
-6435	AGCGCCGAGATCTTAAAGGCACCCAAGTCACAAAGTCGCCATCTGCTCTTGGCAGC	-6376
-6375	TCCTGGACATCTCTCGATATTGGCTTGAAAGCCATGACCCATCATAAGCTGAAAGGCT	-6316
-6315	AGGAGGGCACCATAGGTACCGAAGTACGTTGAATACCTCGAGGACCTCCCTCGTGTG	-6256
-6255	ATGGCGAAAGCATCGATCAGCTGCCCAAGGTCTTGTGATCGATCTGGGAAGATC	-6196
-6195	ATCGAGTGCATCCCGTCATGGATCCTTACCTCTGAAGGAGGTCTGAAAAAGCTGG	-6136
-6135	TGAGACCCGAGGGTCATTGACAAAGCATTGCCGGAGAATTATCGGCAATTATCTAGA	-6076
-6075	GCCTCAGCAGGGATGTAGGCAGCTCTGGAGAAAGTGAAGAGGGAGGAGCTCACTAACCA	-6016
-6015	AAATCAAATCGATAAAGCAAAATCGGAAAGGAGGCCAAAGGGGATTACTGAGCAAGGC	-5956
-5955	CAAGGAAGATTGGCGAAGGAGCTCATCTTTCAATGCCCGAGCTCGGCAGCAAGCCT	-5896
-5895	GGATGCCTCTTCATCCTTCAGCCTCTTAGCCCTCGAGCTCATCCTAAAGGAATC	-5836
-5835	AACCTCCTGGGGCCTCGCAGCTATTTATCGCACCCCTCAGCTTCGAGGAAGAAGA	-5776
-5775	CTCGACCTCTTGCAGCCAGTCTGTCAACTCCAGAGAAGTGTATTGGGAGGCGAA	-5716
-5715	GGCCTCCAGAGAAGAGATAACAGCTCACAAATCCTAACAGAGATAAGGAAAATAATTAGA	-5656
-5655	CGAAGAACTGGTTGTCAACAAACTTATAATTGATCAGGGAAATCGTCCCACATGGATAT	-5596
-5595	ATCGTTAAAACAGGAAAAGCTTACAGGTTCCCTGGAGGAGAAGCTGTAACCACGGCAGT	-5536

FIGURE 39

65/76

FIGURE 39 CONTINUED

66/76

FIGURE 39 CONTINUED

67/76

FIGURE 39 CONTINUED

-1995	AGTTGTTCACTTCACTAGTATGAATTCACTAAATCGGGCAATACTCCAAACACTCATTCA	-1936
-1935	CCCCCTAGGCAGGGTAGCTCAGATCAACGTGGGTCTTCATCGAGTTAATGTCGTCA	-1876
-1875	CACGCACACACACGTACGCCAACACACGTGCGCAAACAAAAAGAAAAGACCTT	-1816
-1815	CTCACGTAGCCTAGGTCTTGTCTGTAAGAAAAACCCAGGTCACCCTAGTTCGAACCC	-1756
-1755	AAAATTTTGAAGATACTTAGTAAGATATTTGAAAATAACCGCAAAAGGGAA	-1696
-1695	TTGAAAATATGGACTGGCTGTTGTCCAAAACCACATCTTCGGAGAACCGACGAGGGT	-1636
-1635	ATCTATTGATGGGCTCATACTATACCTGGCATGTGTTGGCAGGCCCTCATGTCGGCC	-1576
-1575	GAGGAAAGCCCGACGCTGAAAATCAGGCCAAGCTTAACCCGGCCGACCAAATACCCA	-1516
-1515	CCAAACCGTTGGGCCATCAGGTTGCAGGGCAGTAGTGAAAACACCGATTCGGG	-1456
-1455	CTACATAGGCCGGCTCGTTGTGGCAACATTCTAGACCTAACGGCGAGTTTCG	-1396
-1395	GGCCGGGCTGCCATGGCAGGTATAGCTCATACGACGTATGACATTGAGCAATTGA	-1336
-1335	TGCAAAGCACGTGAGGTTTATCCCATCCGTGTGGCGTGTAGGGTGTAAATGAATA	-1276
-1275	GGATAATTCTCGCCGAAACTGGTCCAAATTGCTTGAAGTGTCCATATATGATTT	-1216
-1215	AAAGAATGTGACAATAAGATATCCAATTGAAATAGTGTCCGGATACGGTATAGGA	-1156
-1155	TATGGTATAGCAAATAACATGCTGATATGGATTGTCGATATTAAAGATAATCCAA	-1096
-1095	ATGTTTAAACCGCATAATTGATTTGAGTCAGGCAATTGCAATTGAGTTA	-1036
-1035	GCAGTTATTGAGTTCAAAATTATTGGCGAGCATATCTAGTTCTAAATTCTATCACGT	-976
-975	AAATTGTGTTTTAAATAACTACACAAGACTAAAGTTAAATCTCTCAAGATT	-916
-915	GCGAAAATATAGCTATCTACTGATATATATCCGACTATTTGTTTCGGACCGCAT	-856
-855	GCGTCCTATTCCGATTGAATCTGCACTCCGATATCCACATTGAATCTAAACCGAT	-796

FIGURE 39 CONTINUED

69/76

-795	CAATATTGCTCCGATCTAAATCCGGAAAAATATGTGGTGAAGGGATATGGTATAAGCAA -----+-----+-----+-----+-----+-----+-----+-----	-736
-735	ATCCGATTTGATCCATTGTACCTCTAGGCCTGTGCAAGACCTGGAGGAAAGAATGGCGC -----+-----+-----+-----+-----+-----+-----+-----	-676
-675	ATCTGTAGGGTGCAGTCCCACCGTGAAAATGTGAGCTCACCGTATTGTCCCCGATGG -----+-----+-----+-----+-----+-----+-----+-----	-616
-615	AGCATCGAAACGGAGTCGGAACACGATTGCGCCACGTACAGAGCATGCATGATTCCCT -----+-----+-----+-----+-----+-----+-----+-----	-556
-555	TGTATGCGGTCCAGGATCTTAAACTGCCTTCCATTTCAGGAACCTACCGATTGGCTGCA -----+-----+-----+-----+-----+-----+-----+-----	-496
-495	AGCCGTAGCTAGCGGTTGAAGTCACGGCATTGCCGCCCCGATTAACCCACCCGTCGCG -----+-----+-----+-----+-----+-----+-----+-----	-436
-435	CGCGCGGTGGTCGTTCACCGTCCTGCCCTAGGCTACGCACGCGCGCGCAGTTGGGCC -----+-----+-----+-----+-----+-----+-----+-----	-376
-375	AGTTGTAGGTAAGCCGACTCGAGATCACACACCCGGCCTCACCTACTACCTCTGCCGTC -----+-----+-----+-----+-----+-----+-----+-----	-316
-315	GCGGTCAACCGTGTCAACTCACGCCAGGGAGCCACCCGCCACACGGCGCCTAGCTCA -----+-----+-----+-----+-----+-----+-----+-----	-256
-255	TCCCCTCTCACTACTCTTCTCCTCCCTCTCACCTCGCCGTCGACCCAGCTCCGGCT -----+-----+-----+-----+-----+-----+-----+-----	-196
-195	CTATAAATTCCGCACTACTCGAACCAACATGCCAGGCCTTGCCTTTACGACGAATC -----+-----+-----+-----+-----+-----+-----+-----	-136
-135	CTACCAAACCGAGCTACCAGATCCTCTACTAATCGAGCTCCCTACGCTGCTCCGCC -----+-----+-----+-----+-----+-----+-----+-----	-76
-75	GTCTCGTTCCGCCCTCACGCCGGCGTTCTCCGCTCCAAGCTACGTCCGTCCGTC -----+-----+-----+-----+-----+-----+-----+-----	-16
-15	CATATATAGCATCGACATGACCATGCCGAGGTCTGGCTGCCGGAGACACCGCCGCC -----+-----+-----+-----+-----+-----+-----+----- M T I A E V V A A G D T A A A 44	
45	GGTGGTGCAGCCGCCGGAACGGCAGACCGTGTGCGTGACCGCGCCGGGTACAT -----+-----+-----+-----+-----+-----+-----+----- V V Q P A G N G Q T V C V T G A A G Y I 104	
105	CGCGTCGTGGCTCGTCAAGCTGCTGGAGAAGGGTACACCGTCAAGGGCACCGTCAG -----+-----+-----+-----+-----+-----+-----+----- A S W L V K L L E K G Y T V K G T V R 164	
165	GAACCCAGGCATGTCACCCATGCATTCATCATTTCTTAAGTCGTATGCGTTATGCGA -----+-----+-----+-----+-----+-----+-----+----- N P G 224	
225	CTTGTGTATTAACATTGTGGACTGCATGCAGACGACCCGAAGAACGCGCACCTGAGGGC -----+-----+-----+-----+-----+-----+-----+----- D P K N A H L R A 284	

FIGURE 39 CONTINUED

285	GCTCGACGGCGCCGCCGACCGGCTGGTCTCTGCAAGGCCGACCTCCTCGACTACGACGC -----+-----+-----+-----+-----+-----+-----+-----+ L D G A A D R L V L C K A D L L D Y D A	344
345	CATCCGCCGCCATCGACGGCTGCCACGGCGTCTCCACACCGCGTCCCCCGTCACCGA -----+-----+-----+-----+-----+-----+-----+-----+ I R R A I D G C H G V F H T A S P V T D	404
405	CGACCCGTACGTACTCCATAGAACTCGGCACCCCTAGCTCTCTCCGTTCTGTAT -----+-----+-----+-----+-----+-----+-----+-----+ D P	464
465	GTCTGTCACCCTCGATGCCATGGCAGCACGCATGCATGCGCGCGAACGCTAGCTAGAC -----+-----+-----+-----+-----+-----+-----+-----+ -----	524
525	GCTGACCGACTCATTGTGCAGGAGCAAATGGTGGAGCCGGCGGTGAGGGGCACGCAGTAC -----+-----+-----+-----+-----+-----+-----+-----+ E Q M V E P A V R G T Q Y	584
585	GTCATAGACGGCGGGAGGCCGGACGGTGCGGCGGATGGTGTACCTCCATC -----+-----+-----+-----+-----+-----+-----+-----+ V I D A A A E A G T V R R M V L T S S I	644
645	GGCGCCGTCACCATGGACCCCAACCGCGGGCGGACGTGGTCGACGAGTCGTGCTGG -----+-----+-----+-----+-----+-----+-----+-----+ G A V T M D P N R G P D V V V D E S C W	704
705	AGCGACCTCGACTTCTGCAAGAAAACCAGGGTGGGTGCTGCATGCTCAATTATTATTATC -----+-----+-----+-----+-----+-----+-----+-----+ S D L D F C K K T R	764
765	ATAGCTACCCCTTTCTGCACCATGCTGCATTCTTCAAAAACAACCTCTCAAAAGAT -----+-----+-----+-----+-----+-----+-----+-----+ -----	824
825	ATGCTACGTGGTAGTTCTATAGCTGAATTATTACAACCTACCCCTATCGATCACTAC -----+-----+-----+-----+-----+-----+-----+-----+ -----	884
885	CGCCCTAAAAGTGTCAACTTTGAAGGCAACCAAACCATACATGAACGACGATCGTG -----+-----+-----+-----+-----+-----+-----+-----+ -----	944
945	TGCGCTTGTGCGTTATCATTAGCCTCTGTAGCTCTAATTTCACCTATGTACGCATGG -----+-----+-----+-----+-----+-----+-----+-----+ -----	1004
1005	ATAGACGATTGGAAATACAGTTCACTACCTACCATATACTATGCCGAAATCGAACGC -----+-----+-----+-----+-----+-----+-----+-----+ -----	1064
1065	ACACAGGTGTGAGGCAGCAGCCGTCACGAGTTATGCGCCGAAACCGACATCTCGGAATC -----+-----+-----+-----+-----+-----+-----+-----+ -----	1124
1125	TTCAGTCCACAATAAAAAATAGACACACTGGTACCACTACAAAATTATACTCCTACTGTA -----+-----+-----+-----+-----+-----+-----+-----+ -----	1184
1185	TATTGGTAAAACAAAACATTCTTTTATTTGATAGGAGTGCTGCAAATTAAAGTTCT -----+-----+-----+-----+-----+-----+-----+-----+ -----	1244
1245	TTGTGTCATTTCAAAGGAAAAAAACACCTTACCACTTTCTCCCTGCCATCAT -----+-----+-----+-----+-----+-----+-----+-----+ -----	1304

FIGURE 39 CONTINUED

71/76

1305	TTTTTTTACCAAAGTTGTTCTGTCAAATGAACATATATAGTCGGTGCTATGTCA	1364
1365	G TGCCATTACCGGCCACTAGCTAGTAGGACTGCCATGTTCCAGCAAATTGTCTAGTGGAA	1424
1425	CCGGAGTGGCCAAAAGGAGCCAATTATGTAGGGTTGCAAGCAGGATCACACAAAAGCCTC	1484
1485	GCCTCTAGTTCATTTATCAATTAAGTGGTACTTCTCAGGGACCCCCCTTGCAACTCTA	1544
1545	CCATTACATCCGTGAAAATAAAAGCTAGCATCACGCACCAGATTAGTACTCCCTCCGT	1604
1605	TTTTATTTAGTCGCATTCTAGGTTAGCCAAAGTCATACTTGCAAAGTTAACCAAAA	1664
1665	TTATAAGAAAAAAATATCAATAATCATCATACAAAATACATATAATATAAGAGTAAACCT	1724
1725	TATAACGATTCTACAATAGATTTTTTATTGCATATGTCAATATTTTCATAAAATATTT	1784
1785	ACTCAAAATTATAAGGTTGACTTTGACTAAACCCAGAACCTCTTAGAGAGGAAGAAAT	1844
1845	GCATGGGCAAAGCAAATCATGCATATGGCAGGAGTAACATTTTGACTTTCATAGA	1904
1905	AAGTACTGTATGGCACTAACGGCTAAACCGGACACTGGAAGCAAATCGTCACGTGGG	1964
1965	CAATATTATCTACCGTCGCGTCGCCAGTCTCCCCTGCCATGACCATGCTTGGAAATT	2024
2025	AGTCTCGCCGGAGCTGCCGAGTGCATGCATAGTGACGAGTTCAATAGGCCACTATAT	2084
2085	GTGATCATGGCTTTGATTGTCACTTTCTTTTGCCGAAGGATATAGTAGTATTACT	2144
2145	TTCTCTGCTATCACAAAGAAAGAACTGATTGTCTAGTCTAGGTGGTCTCAGAATTCTG	2204
2205	CATGACTCCAGAGTATTCTGATGCCACTTGTGTTATTGCAAGAAACTTAATCGGAG	2264
2265	ACAACCAAAAGCTCATCCATGTCCTGGAACTAGTAGACATAAGAAAATCTCATGGTAT	2324
2325	CAGTTTGCTATTTATCTACAACGTAAACGGCATGTTGGTTTATTAAATTCAAGAACTGG	2384
2385	TACTGCTACGGAAAGGCGGTTGCGGAGCAGGCGGCATCGGAGTTGGCGGGCAGCGCGGC	2444
	N W	
	Y C Y G K A V A E O A A S E L A R O R G	

FIGURE 39 CONTINUED

72/76

2445	G T G G A C C T T G T G G T G G T G A A C C C G G T G C T G G T G A T C G G C C C C T G C T G C A G C C G A C G G T G V D L V V V N P V L V I G P L L Q P T V	2504
2505	A A C G C C A G C A T C G G C C A C A T C C T C A A G T A C C T G G A C G G G T C G G C C A G C A A G T T C G C C A A C N A S I G H I L K Y L D G S A S K F A N	2564
2565	G C C G T G C A G G C G T A C G T G G A C G T C C G C G A C G T G G C C G A C G C C C A C C T C C G C G T C T T C G A G A V Q A Y V D V R D V A D A H L R V F E	2624
2625	T G C G C C G C C G C G T C C G G C C A C C T C T G C G C C G A G C G C G T C C T C C A C C G C G A G G A C G T C C A A A S G R H L C A E R V L H R E D V	2684
2685	G T G C G C A T C C T C G C C A A G C T C T T C C C G A G T A C C C G T C C C C A C C A G G T A C G C G T A C G A C V R I L A K L F P E Y P V P T R	2744
2745	C T G C T T G C T A G C C G T T C C G T T A A T T C C A T T G C T T A A T T G A T T G C A T G A T G C C G C T C C T A A T T T A C T C A C T T G C G T A A C T A A T T G C A T T C A T A T G A T C T A C C A A C C G T G G A G A A A T	2804
2805	T A G C A A G A G T C T G T C G G G C G T C C C G G T C C A G T G C A G T T A A C C T G C A T G T C G A T G G T C T G C A G G T T G C A G C T T A C T T G T G G T T C T T A G T T C A G A G A C A C A G A G C A A T T G G G C A C T A A G C	2864
2865	A A A A C T G A C A T C A C T T G G T A A T T A G G T A G C T C C C A C A C A C T G A A G T G G G T G G A T C C C A T C G G T A G T T A G G G T A A G G G T G G A T A G T A C T G G A C G A G A G C T C G A T C G T G T G T G A A A A A A G C G A G	2924
2925	T G A C C A C C A T T C A C C A T C C A C T G C A A G T A G C T G C T A G T G A A C C A T C C A A C C A G C T C C C T G G A T C A C T C T G C T C C G T C C G T A C C T C A G C T A C C T A C A G A A G C G A C A T G A A C A C A C A G A C	2984
2985	A A A A C T G A C A T C A C T T G G T A A T T A G G T A G C T C C C A C A C A C T G A A G T G G G T G G A T C C C A T C G C C A C A A G G C C G G C T C A C C A T T C G C A T A G G T C A A A C C A A T G T T G G T G A A C G G C A A C A T C G	3044
3045	C C A C A A G G C C G G C T C A C C A T T C G C A T A G G T C A A A C C A A T G T T G G T G A A C G G C A A C A T C G C C A C A A G G C C G G C T C A C C A T T C G C A T A G G T C A A A C C A A T G T T G G T G A A C G G C A A C A T C G	3104
3105	C C A C A A G G C C G G C T C A C C A T T C G C A T A G G T C A A A C C A A T G T T G G T G A A C G G C A A C A T C G T G A C C A C C A T T C A C C A T C C A C T G C A A G T A G C T G C T A G T G A A C C A T C C A A C C A G C T C C C T	3164
3165	T G C C C G T C G C T G A T A T T G C A C G C G T A G C T G T G A C G A A A G T A G G T G G A C T G A C A G A T A C G G A T C A C T C T G C T C C G T C C G T A C C T C A G C T A C C T A C A G A A G C G A C A T G A A C A C A C A G A C	3224
3225	T G C C C G T C G C T G A T A T T G C A C G C G T A G C T G T G A C G A A A G T A G G T G G A C T G A C A G A T A C A C A C A A G G C C G G C T C A C C A T T C G C A T A G G T C A A A C C A A T G T T G G T G A A C G G C A A C A T C G	3284
3285	T G C C C G T C G C T G A T A T T G C A C G C G T A G C T G T G A C G A A A G T A G G T G G A C T G A C A G A T A C C C A C A A G G C C G G C T C A C C A T T C G C A T A G G T C A A A C C A A T G T T G G T G A A C G G C A A C A T C G	3344
3345	T G C C C G T C G C T G A T A T T G C A C G C G T A G C T G T G A C G A A A G T A G G T G G A C T G A C A G A T A C T G C C C G T C G C T G A T A T T G C A C G C G T A G C T G T G A C G A A A G T A G G T G G A C T G A C A G A T A C	3404
3405	T A T C C G C A C G G C A G A A C G C G T A G C A T C A G G G C C A G A A A G C A G C G T G C G T G A T A T C G T A A C A C A T A T C C T C A T T G C C T T C T G C T C G G T T C T G C T A G G A T T G C C A T C T C A G G A G T G C C	3464
3465	T A T C C G C A C G G C A G A A C G C G T A G C A T C A G G G C C A G A A A G C A G C G T G C G T G A T A T C G T A A C T A T C C G C A C G G C A G A A C G C G T A G C A T C A G G G C C A G A A A G C A G C G T G C G T G A T A T C G T A A C	3524

FIGURE 39 CONTINUED

73 / 76

3525	CCAGACGGTCTCACCTGTCATTCTGGGCTACCTGGCATACTACCTCGGTGCCGCTGTG -----+-----+-----+-----+-----+-----+-----+-----+	3584
3585	CCGCTGACCAATTGCGCACGACCCTATAGCAAAACCCATTGCATGTAAGTGCCTCAAG -----+-----+-----+-----+-----+-----+-----+-----+	3644
3645	ATCAGCAGTGACATGTGCAATATAAACCTCAAGTGTGCACTCTAGTGCCTACTGATAAAA -----+-----+-----+-----+-----+-----+-----+-----+	3704
3705	CCGTATAACTGGTGACCCAGTCATTCTCTCTTTTATTTGTTGGACCAAACGAACAC -----+-----+-----+-----+-----+-----+-----+-----+	3764
3765	AGCATGTTATCCATACCAACAAGTGGCGCTGATTTTCAAACACTACACTGGGATCATACT -----+-----+-----+-----+-----+-----+-----+-----+	3824
3825	GGAAACCAAAGCAGGAGAACATCTCGAACCAAGAGATGTTACTAAATTGAAAGAAAA -----+-----+-----+-----+-----+-----+-----+-----+	3884
3885	TGTACTGACAAGTAATCTGCTGAAAGCAAGACACATACTACCTCGGTTCGAACGTGGGAC -----+-----+-----+-----+-----+-----+-----+-----+	3944
3945	ACCATGCCGTGCCATATTGCTAGGCACCACTCTGCCGTCGATTGTATCCAACGGAGG -----+-----+-----+-----+-----+-----+-----+-----+	4004
4005	GAGTATCGATTTGCGAAAGTTCCCTACATACATAGCCGCTCAAGATATAATCTTACGACC -----+-----+-----+-----+-----+-----+-----+-----+	4064
4065	TTCCGTCGAAATCGGTGATACGTCGCAACCTATAGCTAACCTGGCAGAGCATAAAATAAC -----+-----+-----+-----+-----+-----+-----+-----+	4124
4125	TATCTAAGGTTGGGTCTCCCTCTTTCAATCAACCTTCATACCGAATGATGGAGTGT -----+-----+-----+-----+-----+-----+-----+-----+	4184
4185	TTGTGAAAACATCTCTGGTCGACTCAGCATTAGCGCCCTACCAATTCTCTGTGGACAA -----+-----+-----+-----+-----+-----+-----+-----+	4244
4245	TGCCACCTTAAATCGTTTTAGTCATGATTACTCCCCTTATATCTGGCCGTAGT -----+-----+-----+-----+-----+-----+-----+-----+	4304
4305	CCCTCTTTCCATTTCCTGTCTGGTTAAAGTCAAATTAGACTACTAAAACACAGC -----+-----+-----+-----+-----+-----+-----+-----+	4364
4365	AAGATTTATGGAAGGGAGGTAGTGCAAAACAGAAAGTCGATCGAAATGCGTGCCAATT -----+-----+-----+-----+-----+-----+-----+-----+	4424
4425	TGTCGTCGGCGGCCGGACTAAAATGGATCTGCATGTGCATACCGTTCGCGAGTATC -----+-----+-----+-----+-----+-----+-----+-----+	4484
4485	CTGCGAACGGTCGTGTTAGTCACATTAATGTGAGGTTCATGTGATACTCTTGCTTG -----+-----+-----+-----+-----+-----+-----+-----+	4544
4545	AAAGATACTACTACTGCTACCTCGTAGAACTGAATGAAAGTATGTGGACTGTTCAGCTC -----+-----+-----+-----+-----+-----+-----+-----+	4604
4605	TCTGCACATGTCAAATGTCGTTACTCATACCTTCGTCAGAGCATCCTGCGACGCGCGCC -----+-----+-----+-----+-----+-----+-----+-----+	4664
4665	GGTGCCTGAAATTGCGCGTGTGTTAGTCAGAGATCAACGTGAGGTTCATGCGGTACCTA -----+-----+-----+-----+-----+-----+-----+-----+	4724

FIGURE 39 CONTINUED

74/76

FIGURE 39 CONTINUED

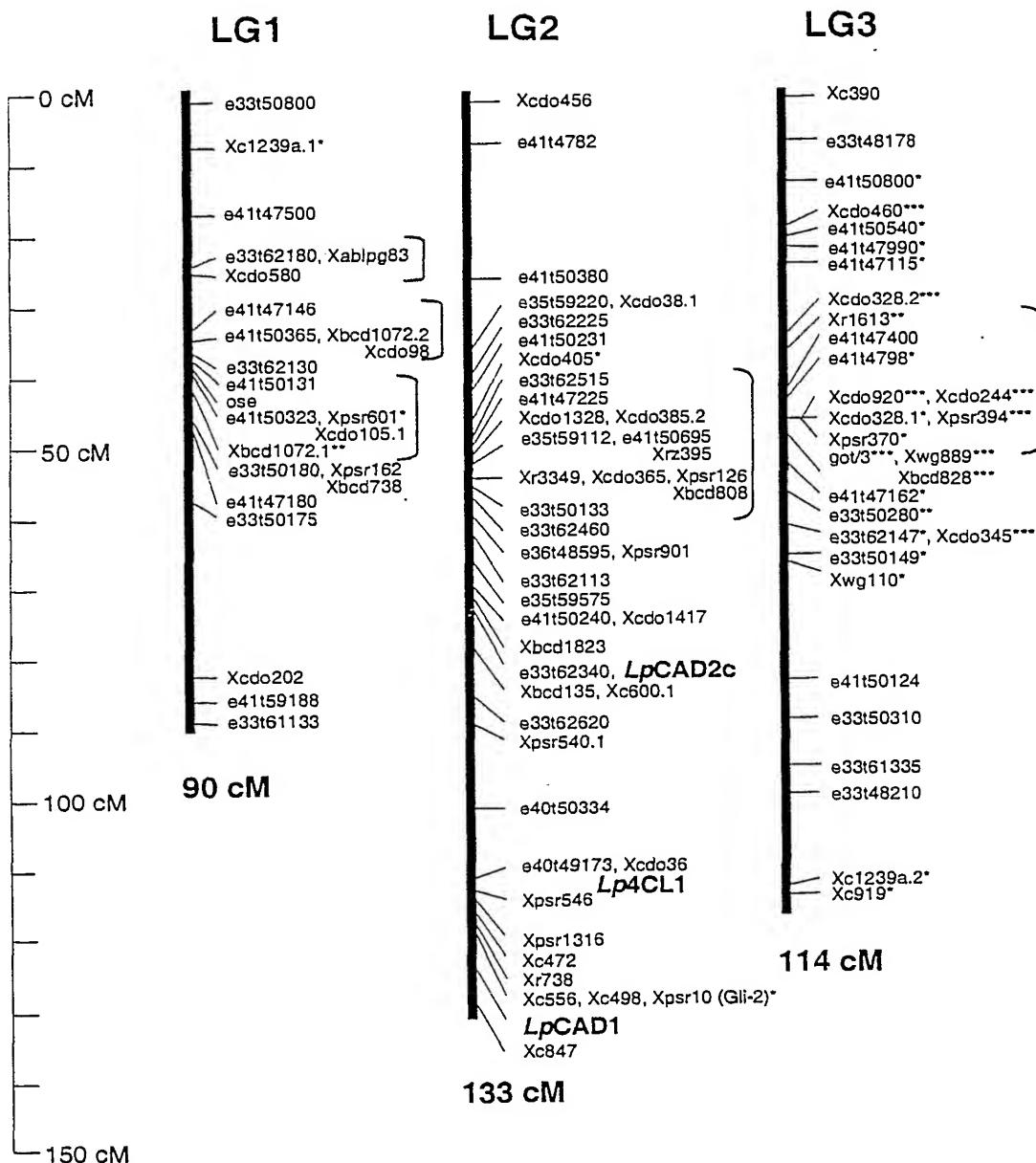


FIGURE 40

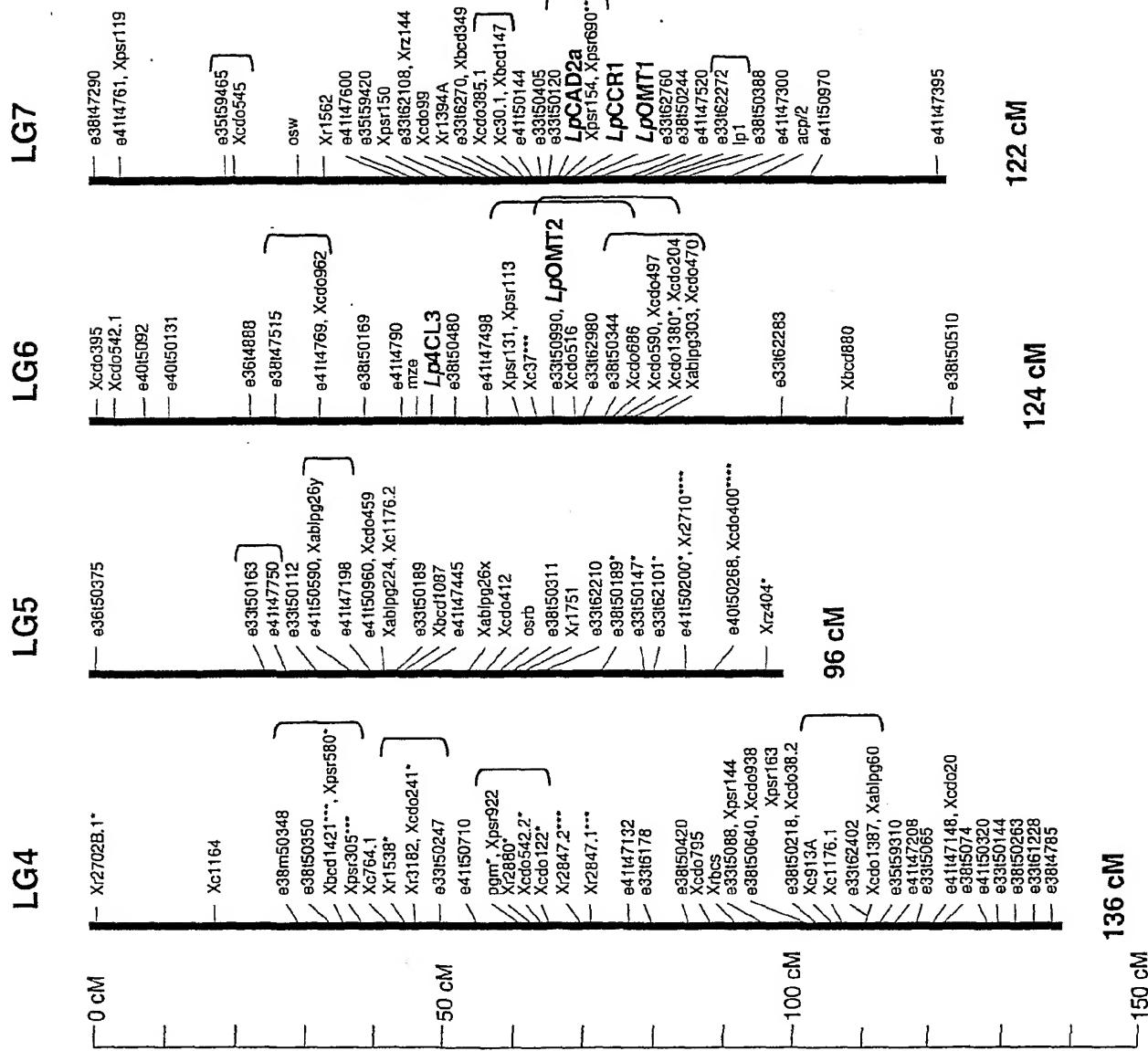


FIGURE 40 CONTINUED

Organization Applicant

Street : 15th Floor, 8 Nicholson Street
City : East Melbourne
State : Victoria
Country : Australia
PostalCode : 3002
PhoneNumber :
FaxNumber :
EmailAddress :

<110> OrganizationName : State of Victoria as represented by Department of Natural Resources and Environment

Organization Applicant

Street : North Terrace
City : Adelaide
State : South Australia
Country : Australia
PostalCode : 5005
PhoneNumber :
FaxNumber :
EmailAddress :

<110> OrganizationName : The University of Adelaide

Organization Applicant

Street : Lisboa 27, Apartado Postal 6-641
City : Mexcio
State : DF
Country : Mexico
PostalCode : 06600
PhoneNumber :
FaxNumber :
EmailAddress :

<110> OrganizationName : International Maize and Wheat Improvement Center

Organization Applicant

Street : Waite Road
City : Glen Osmond
State : South Australia
Country : Australia
PostalCode : 5064
PhoneNumber :
FaxNumber :
EmailAddress :

<110> OrganizationName : State of South Australia as represented by South Australian Research and Development Institute

Organization Applicant

Street : Military Road
City : Lismore
State : New South Wales
Country : Australia
PostalCode : 2580
PhoneNumber :
FaxNumber :
EmailAddress :

<110> OrganizationName : Southern Cross University

Organization Applicant

Street : Level 3, 84 William Street
City : Melbourne
State : Victoria
Country : Australia

PostalCode : 3000
 PhoneNumber :
 FaxNumber :
 EmailAddress :
<110> OrganizationName : Dairy Research and Development Corporation

Application Project

<120> Title : Modification of Lignin Biosynthesis
<130> AppFileReference : 40494788
<140> CurrentAppNumber : AU PQ8154
<141> CurrentFilingDate : 2001-06-14

Earlier Applications

<150> PriorAppNumber : AU PQ8154
<151> PriorFilingDate : 2000-06-14

Sequence

<213> OrganismName : Lolium perenne
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cactg cctgaagatg 2040gggttcatggc ttcatgtctaa tcatttcgt cagaaaggca cttctagcat ata
tgttcca 2100ccctttgttt catttggaaatg attgtattcc agctagtggc cagtgcactga gtaaggatg
2160gggataaaaatg ttttgtctac gttttctttt acgctactt ctcattggg ggttacaatg 2220tat
caggggaa ttcgtgattt aagttaatca agattggttc aattataaaaa aaaaaaaaaa 2280aaaa
2284

<212> Type : DNA
<211> Length : 2284
 SequenceName : 4CL1cDNA (SEQ ID NO: 1)
 SequenceDescription :

Custom Codon

 Sequence Name : 4CL1cDNA (SEQ ID NO: 1)

Sequence

```

<213> OrganismName : Lolium perenne
<400> PreSequenceString :
MITVAAPEVQ QPQIAAAAAA VEAAAPEATT IFRSRLPDID IPTHMLHDY CFATAASAPD      60APCLITAAT
G KTYTFAETHL LCRKAAAALH GLGVRHGDRI MLLLQNSVEF ALAFFGASML      120GAVSTAANPF CTPQEIH
KQL VASGAKLVVT QSAYVDKLRH EAFPRIGEAL TVITIDEDDG      180TPDGCQPFWA LVSAADENSV PESPI
SPDDA VALPYSSGTT GLPKGVVLTH GGLVSSVAQQ      240VDGENPNLHM RAGEDVVLCV LPLFHIFSLN SVL
LCALRAG AAVMLMPRF E MGAMLEGIER      300WRVTVAAVVP PLVLALAKNP GVEKHDLSI RIVLSGAAAPL G
KELEDALRG RLPQAIFQGQ      360YGMTEAGPVL SMCPAFAREP TPAKSGSCGT VVRNAQLKVV DPDTGVSLSGR
NLPGECIRG      420PQIMKGYLND PVATAATIDV EGWLHTGDIG YVDDDDEVFI VDRVKELIK KGFQVPPA
EL      480EALLIAHPSI ADAAVVPQKD DAAGEVPVAF VVRAADSDIA EEAIKEFVSK QVVFYKRLHK      54
OYVFTHAIPKS ASGKILRKEL RAKLAAPATA RVVHGFMLII SIRKALLAYM FHLLFHLEDC      600IPASGQ
                                         606

```

```
<212> Type : PRT
<211> Length : 606
      SequenceName : 4CL1pep (SEQ ID NO: 2)
      SequenceDescription :
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Sequence

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<213> OrganismName : Lolium perenne
<400> PreSequenceString :
cggcacgagc gccattcctc caccctcagc tccggccaaa gatttccatc cggcgagatc      60catgggctc
c atcgcggcgg acgcgcctcc cgccgagctg gtgttccgggt ccaagctccc      120ggacatcgag atcccgaa
ccc acctgacgct gcaggactac tgcttcagc gcctggcga      180gtctccgcg cgccgctgccc tcatc
gacgg cgccacgggc gccgcgtca cctacggga      240ggtgacgccc ctgtcccgc gctgcgcgc ggg
gctgcgc cgcctcggcg tcggcaaggg      300cgacgtcgtc atggcgctcc tccgcaactg ccccgagttc g
ccttcgtgt tcctcggcgc      360ggcccggtc ggcgcgcaca ccaccacccgca accccgttc tacacgcccc
acagagatcca      420ccgccaggcc accgcgcggc gggccagggt catcgtaacc gaggccgtcgcc cgctcgagg
aa      480ggtgcgcgcc ttccgcggcc agagaggat tcccgctcgcc tcgtcgacg agggcgctcg      54
0cgccggctgc ctcccggttc ccgagactct gtcggggaa gaaagcggggg acgcgggttcg      600cgacgagg
cg gtcgaccccg acgacgttgtt ggcgtcgccg tactcggtcg gcaccacccg      660cctgcccgg ggcgtc
atgc tcacccacccg cagcctcgatc accagcgatc cccagcagg      720ggacggttag aaccggaaacc tgca
cttcag ctcgatcgac gtgtgtgtt ggcgtcgatc      780gctgttccac atctactcgatc tcaactcggt gc
tgctcgcc ggtctccggc cgggtgtcg      840ga
tcgtgatc atgcgcgaatc tcgaccacgg cgccgtgggt gacctgggtc gcacgcacgg      900cgtcaccgtg
gcgcattatcg tgccgcctat cgtggggag atcgccaaaga gcgcgcgggt      960gaccgcgcgc gacctggcg
t ccattccggct ggtcatgtcg ggggcggcgc ccattgggaa      1020ggagctcgacg gacgcgttca tggccaa
gat ccccaacgcgat gtcgtcgcc aggatatgg      1080gatgaccgag gccggccctgt tgctggcgat gtcg
tggcc ttccgcggaaaggccgttcg      1140ggtcgaatcc gggttccgtcg gcaccgtcgat cagaacgcgac ggg
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ccggcaagca      1260gatcatggaa ggttacctaa atgtccggt ggcacaaag aaccatttg acaaggacgg
      1320ttggctgat actgggtgaca ttggttatgt cgatgtatc gacgagatct ttattgtcg      1380c
agactgaag gagataatta aatataaggg attccaaatc cttccggcg aacttgaagc      1440ccttctcatt
    acacaccctg aaatcaaggg tgctgtgtc gtatcgatgc aagacgaaact      1500tgctgggtgaa gttccgggt
tg cgtttgttgcgatcgatc ggttccggaa tcagcgaaaa      1560cgagatcaag cagttcgatc caaaagg
aggat tttttctac aagaggatct gcaaaatgtt      1620cttcgcggat tccattccaa agatccatc tggc
aagatc ctcaggaagg acctgagatc      1680aaag
ctcgcc gcaggccatc ccacgcgttcc taccacacag tccaaaatgc aagtccatc      1740tattgtttcc ca
acccatc caccctgtc ccacaccatc taatgttctt aatataaaacg      1800gaaatttata catatagaag
ggctgttgcgtt ttttactaga tttgttccaaat atatgtatc      1860cttggtaggc cgtatgtatc taatctgtc
a tttgtatagata ccgcctttt tttgttccaaat aatgttatac cgtgtactatc atatgttgc
ttc agggagatca aaaaaaaaaaa      1980aaaaaaaaaa aa
1992

```

```
<212> Type : DNA
<211> Length : 1992
      SequenceName : 4CL2cDNA (SEQ ID NO: 3)
      SequenceDescription :
```

Custom Codon

Sequence Name : 4CL-2cDNA (SEQ ID NO: 3)

Sequence

<213> OrganismName : *Lolium perenne*
<400> PreSequenceString : MGSTAADAPP AEIVFERSKLP DIELIPTHIITL ODYCEFORLPE LSARACIJDG ATGAALTYGE 60VDALSRRCAG

A GLRRLGVGKG DVVMALLRNC PEFAFVFLGA ARLGAATTAA NPFYTPHEIH 120RQATAAGARV IVTEACA
 VEK VRAFAAERGI PVSVSDEGVDF GGCLPFAETL LGEESEGERFV 180DEAVDPDDVV ALPYSSGTTG LPKGV
 MLTHR SLVTSAQQV DGENPNLHFS SSDVLLCVP 240LFHIYSLNSV LLAGLRAGCA IVIMRKFDHG ALV
 DLVRTHG VTVAPFVPPV VVEIAKSARV 300TAADLASIRL VMSGAAPMGK ELQDAFMAKI PNAVLGQGYG M
 TEAGPVPLAM CLAFAKEPFA 360VKSGSCGTVV RNAELKIVDP DTGASLGRNL PGEICIRGKQ IMKGYLNDPV
 ATKNTIDKD 420WLHTGDIGYV DDDDEIFIVD RLKEIKYKG FQVPPAELEA LLITHPEIKD AAVVSMQD
 EL 480AGEVPVAFVV RTEGSEISEN EIKQFVAKEV VFYKRICKVF FADSIPKSPS GKILRKDLRA 54
 OKLAAGIPSSN TTQSKS 556
 <212> Type : PRT
 <211> Length : 556
 SequenceName : 4CLpep (SEQ ID NO: 4)
 SequenceDescription :

Sequence

<213> OrganismName : Lolium perenne

<400> PreSequenceString :

cggcacgaga tctcccacga ctaattttaga agaagattta cttagtctct gcttctcgct 60cgatcgccg
 g ccggtaggat agctagctag ctactcgtac tagaccatta ccatgggttc 120cgtgccggag gagtcag
 tgg tggcggtggc accggcggag acgggtttcc ggtcgaagct 180ccccgacatc gagatcaaca acgag
 cagac gctcagagc tactgcttcg agaagatggc 240cgaggctcg tcccgccccct gcatcatcga cgg
 ccagacg ggccgcctcc acacccatc 300ggaggtcgac tccctgaccc gtccgcgcgc ggcggggctg c
 gccgcacatgg gcgtgggaa 360gggcgacgtg gtatgaaacc tgctgcgcaa ctgcccggag ttcccttct
 ccttcctggg 420cgccgcgcg ctgggcgcgc ccaccaccac cgccaaccc ttctacaccc cgcacgag
 at 480ccaccgcacg gcccggccgg cgggcgcacaa gctgatcgatg accgaggcct gcggcgtgg 54
 0gaaggtgtcg gagttcgccg cgggggggggg cgtgcccgtg gtcaccgtcg acggggaggcg 600cgacgggt
 gc gtggacttcg cggactgtat cggccggag gagctggccg aggccggacga 660ggccggggtc ctcccc
 gacg acgtcgtcgc ctcgcctac tcctccggca ccaccgggt 720cccccaaggcgc gtcatgctca cccca
 ccgcacg ctcgtcacc acgtcgtcccc agtgtgtcg 780cggtcgaac cctaactgtgt gttcaacaa gg
 acgacgcg ctgtgtgcc tgctggcgt 840ct
 tccacatc tactcgtc acacgggtct gctggcgcccc ctccgcgtcg ggcgcgcacat 900cgatcatcatg
 cgcacatcg acgtcgtcgc gctgggtggac ctgcgtccgcg cgcacccgc 960caccatcgcc ccattcg
 c cgcacatcg cgtggagatc gccaagagcg accgcgtcgg 1020cgccgacgac ctcgcacatcca tccgcac
 ggt gctctccggc gccgcgcacca tgggcaagga 1080cctccaggac gccttcatgg ccaagatccc caacg
 ccgtg ctccggcagg ggtacggat 1140gaccggaggc gggccgggtgc tggccatgtg cctggcggtc ggc
 aaggagc cgttcaaggt 1200caagtccggg tgctgcggaa ccgtggcgccg caacgcggag ctcaaggtcg t
 cgaccccg 1260caccggcgc tccctccggc ggaaccagcc tggcagatt tgctccggg ggaaggcagat
 1320catgataggt tacctgaacg acccagagtc gaccaagaac accatcgaca aggacggctg 1380g
 ctgcacacc ggagacatcg gcttggtgga tgacgacgac gagatctca tcgtcgacag 1440gctcaaggag
 atcatcaagt acaagggtctt ccaagtggcg cggccggagc tcgaggccct 1500cctccctcacg aacccgg
 gg tcaaggacgc cgccgtcgtt ggggtgaagg atgatctctg 1560cggcgaagtc ccgtcgccct tcatta
 agag gatcaagga tctgagatca acgagaacga 1620gatcaagcaa ttctgtctcaa aggagggttgt tt
 tacaag aggtcaaca aggtctactt 1680cacc
 gactcc attcccaaga acccttccgg caagatcata aggaaggact tgagagccag 1740gctcgccgct gg
 catccca cgaatgtcg cgcgcggaga agctaaggcc cgcttctcg 1800gaacgcagtc acccatggtg
 ctgttttaggt gctgttatag accacaccaa atggggaaag 1860aaaactacggg aggggatcat attatgt
 g caggagat cagttgttg attccgttctg 1920cttgcgtat gttgataaaaa tgaatgata taataga
 tgt gttgtttat ttttgacca 1980tgtaagaaca aggctgtttt atacactact tatttttga aaaaa
 aaaaa aaaaaaaaaa 2038

<212> Type : DNA

<211> Length : 2038

SequenceName : 4CLcDNA (SEQ ID NO: 5)

SequenceDescription :

Custom Codon

Sequence Name : 4CLcDNA (SEQ ID NO: 5)

Sequence

<213> OrganismName : Lolium perenne

<400> PreSequenceString :

MGSVPEESVV AVAPAETVFR SKLPDIEINN EQTLQSYCFE KMAEVASRPC IIDGQTGASY 60TYTEVDSL
 R RAAAGLRRMG VGKGDDVVMNL LRNCPEFAFS FLGAARLGAA TTTANPFYTP 120HEIHRQAEAA GAKLIVT
 EAC AVEKVLEFAA GRGVPVVTVD GRRDGCVDFA ELIAGEELPE 180ADEAGVLPDD VVALPYSSGT TGLPK
 GVMLT HRSILVTSVAQ LVDGSNPVC FNKDDALLCL 240LPLFHIYSLH TVLLAGLRVG AAIVIMRKFD VGA
 LVLDLVRM HRITIAPFVP PIVVEIAKSD 300RVGADDLASI RMVLSGAAPM GKDLQDAFMA KIPNAVLGQG Y
 GMTEAGPVLM AMCLAFAKEP 360FKVKSGSCGT VVRNAELKVV DPDTGASLGR NQPGEICVRG KQIMIGYLND

PESTKNTIDK 420DGWLHTGDIG LVDDDDEIFI VDRLKEIIKY KGFQVAPAEI EALLLTNPEV KDAAVVGV
 KD 480DLCGEVPVAF IKRIEGSEIN ENEIKQFVSK EVVFYKRINK VYFTDSIPKN PSGKILRKDL 54
 ORARLAAGIPT EVAAPRS 557
 <212> Type : PRT
 <211> Length : 557
 SequenceName : 4CLpep (SEQ ID NO: 6)
 SequenceDescription :

Sequence

 <213> OrganismName : Lolium perenne
 <400> PreSequenceString :
 ggcacgagga atcctaccaa accgagctac cagatccttc tctactaatac gagctcccta 60cgctgctcc
 g cctgtcttcg ttccgcctc accgcccggcc gtttctccgc tccaagctac 120gtccgtccgt ccacata
 tat agcatcgaca tgaccatcgc cgaggtcgtg gtcgccccggag 180acaccgcccgc cgcgggtgggt cagcc
 ccccg ggaacgggca gaccgtgtgc gtgaccggcg 240ccggccggta catcgctcg tggctcgta agc
 tgctgct ggagaagggg tacaccgtca 300agggccaccgt caggaaccca gacgacccga agaacgcgc a
 ctgaggggcg ctcgacggcg 360ccggccgaccg gctggctc tcgaaggccg acctccctcgat ctcgacgc
 atccggcgcg 420ccatcgacgg ctgccacggc gtctccaca cgcgtcccc cgtcaccgc gaccccg
 gc 480aaatggtgg gccggcggtg aggggcacgc agtacgtcat agacgcggcg gggaggccg 54
 0gcacggcgcg gggatggtg ctcacccctc ccacccgcgc cgtcaccatg gaccccaacc 600gcggggccg
 ga cgtggctc gacgagtcgt gctggagcga ctcgacttc tgcaagaaaa 660ccaggaactg gtactc
 ctac gggaggccg ttggcgagca ggcggcatcg gagttggcgc 720ggcagcgcgg cgtggacattt gtgg
 tggta acccggtgct ggtatcgcc cccctgctc 780agccgacggt gaacgcgc caca tc
 ctcagaatc ctcggacggg tcggccagca 840ag
 ttcgcacaa cgcgtcgacg gcgtacgtgg acgtccgcga cgtggccgac gcccaccc 900gcgtcttcga
 gtgcgcgcg cgtccggcc gccacccctcg cgcgcgcgc gtcctccacc 960gcgaggacgt cgtgcgc
 catc ctcgcacaaacgc tctcccgatc gtaccccgatc cccaccaggat 1020gctctgatga gacgaacccg
 agc catacaagat gtcgaacccatc aagctccagg 1080acctcgact ctagttccagg ccgggtgagcc agtcc
 ctgta cgagacgggtg aagagccctc 1140aggagaaggg ccacccctcg gtcgcgcg agcaggcaga ggc
 ggacaag gaaaccctag 1200ctggcgact gcaaggcagggttaccatcc gacatgagg aacaagaat
 aaccatgtc 1260cataactgcta ctgtcatgt aaccagctgt tgaatgccta aaatctaagt tcttgtaata
 1320ctgtgttgg tcatgtggac tagattgatc gaataaacat ctctacacaaa gttgctaaa 1380a
 aaaaaaaaaaaaaa 1395
 <212> Type : DNA
 <211> Length : 1395
 SequenceName : CCR1cDNA (SEQ ID NO: 7)
 SequenceDescription :

Custom Codon

 Sequence Name : CCR1cDNA (SEQ ID NO: 7)

Sequence

 <213> OrganismName : Lolium perenne
 <400> PreSequenceString :
 MTIAEVVAAG DTAAAVVQPA GNGQTVCVTG AAGYIASLWV KLLLEKGYTV KGTVRNPDDP 60KNAHLRALD
 G AADRLVLCKA DLLDYDAIRR AIDGCHGVFH TASFVTDDEP QMVEPAVRGT 120QYVIDAAAEE GTVRRMV
 LTS SIGAVTMDPN RGPDVVVDES CWSLDLFCKK TRNWYCYGKA 180VAEQAASELA RQRGVDLVVV NPVLV
 IGPLL QPTVNASHG ILKYLDGSAS KFANAVQAYV 240DVRDVADAHL RVFECAAASG RHLCAERVLH RED
 VVRILAK LFPEYPVPTR CSDETNPRKQ 300PYKMSNQKLQ DLGLEFRPVS QSLYETVKSL QEKGHLPVLS E
 QAEADKETL AAELQAGVTI 360RA
 362
 <212> Type : PRT
 <211> Length : 362
 SequenceName : CCR1pep (SEQ ID NO: 8)
 SequenceDescription :

Sequence

 <213> OrganismName : Lolium perenne
 <400> PreSequenceString :
 ggcacgagca acaagtcatc aatggcgaa ggcttgcggg cgctcggtt ggctgcgagg 60gacgcctcc
 g gtcacccctcg cccttacatcg ttctcgagaa gcgtccgaa ggacgacat 120gtgacgatca aggtgtat
 ctt ctgcggatc tgccacactg acctccacat catcaagaac 180gactggggca acgcctcta cccca
 tcgtc ccaggccatcg agatcggtgg cgtcgtcgcc 240agcgtcgca ggcggcgtcag cagcttcaag ggc
 ggcaca cggctggcg 300ctcgactcctt gccgcacccatc ctacagctgc agcaagggtt a

cgagaactt ctgccccacc 360ctgacgctca cctccaaacgg cgtcgacggc ggccggcgcca ccaccagggg
 cggcttctcc 420gacgtctcg tcgtcaacaa ggactacgtc atccgcgtcc cggacaacct gcccctgg
 cc 480ggcgccac ctctcctctg cgccggcgac acagtctaca gccctatggt ggagtgacggc 54
 Octcaacgcac ccgggaagca cytcggcgac gtcggcctgg gcgggctgg ccacgtcgcc 600gtcaagtt
 cg gcaaggcctt cgggatgacc gtcacccgtca tcagctcctc ggacagagaag 660cgcgacgagg cgctcg
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 catggggc agatggtcgt ggtgggccc 840cc
 cagcaagc cgctcgagct cccggccttc gccatcatcg gccgctcgcc 900ggggacggca
 cccggcagcg cgcacatgc caggccatgc tcgacttcgc gggcaagcac 960ggcattacccg cgcacgtcg
 a ggtcgtaag atggactacg gtcaacaccc ccatcgagcg 1020gctagagaag aacgacgtca ggtaccc
 ctt cgtcatcgac gtcggcgcc gccacccgtca 1080gggcacccgccc gcttaacttg tgctacacaa tgtgg
 acgcg cgctcggtt gtcggaaaaaa 1140aggttcgccc gtcacagcc acatgaacaa gtcaatgagt cgt
 tggtgtg ttgttatct 1200tcattccaca tatgggacgc agttccagat tttcatgtca aataattgcg t
 cgtgtcgcc 1260ttgtcaagac tcaaataaaaaaaa gaaaaaaa 1320aaaaaa
 1320aaaaaa
 <212> Type : DNA
 <211> Length : 1325
 SequenceName : CAD1cDNA (SEQ ID NO: 9)
 SequenceDescription :

Custom Codon

Sequence Name : CAD1cDNA (SEQ ID NO: 9)

Sequence

<213> OrganismName : Lolium perenne
 <400> PreSequenceString :
 MAEGLPALGW AARDASGHLS PYSFSRSVPK DDDVTIKVLF CGICHTDLHI IKNDWGNALY 60PIVPGHEIV
 G VVASVGSGVS SFKAGDTVGV GYFLDSCRTY YSCSKGYENF CPTLTLTSNG 120VDGGGATTQG GFSDVLV
 VNK DYVIRVPDNL PLAGAAPLLC AGVTVYSPMV EYGLNAPGKH 180XGVVGLGGLG HVAVKFGKAF GMTVT
 VISSS DRKRDEALGR LGADAFLVSS DPEQMKAAG 240TMDGIIDTVS AGHPIVPLLD LLKPMGQMVV VGA
 PSKPLEL PAFAIIGGGK RLAGSGTGGSV 300AHCQAMLDFA GKHGITADVE VVKMDYQHR HRAAREERRQ V
 PLRHRRLRRQ PPAGHRRRLTC 360ATQCGRALVW SRKRFAGSQP HEQVNESLVC CLSSFHIWDA VPDFHV
 407
 <212> Type : PRT
 <211> Length : 407
 SequenceName : CAD1pep (SEQ ID NO: 10)
 SequenceDescription :

Sequence

<213> OrganismName : Lolium prenne
 <400> PreSequenceString :
 ggcacagtc gcctcaacg tcttccctta accggccgtc cctacgcttg caccaccacc 60acgcacaga
 c agagcagttt cccagcccccc gccggaaaccg gatggcaccc acggccggccg 120agcagacggc gcaccac
 cag cacaccaggaa aggccgtggg gctggcgccg cgcgacgacg 180ccggccacct ctccccgctc gccat
 cacac ggaggagcac aggagacgac gatgtggta 240taaaagattt gtactcgacg atctgccact ctg
 acctgca cgcctgaag aacgactgaa 300agaactcaag gtacccgatg atccccggc acgagatcgc c
 ggcgaggc acggagggtgg 360gcaagaacgt gagcaagttc aaggccggc accgcgtggg cgtcgggtgc
 atggtaact 420cgtggcggtc gtgcgagagc tgccgacaagg gttcgagaa ccactgccc ggcgtat
 cc 480tcacccataa ctcggctgac gtgcacggca cggcaccta cggccgtctc tccagcatgg 54
 0tgggtgggtca cgagccgttc gtggccgggt tcccgacgc catggcgctg gacaaggccg 600cggcgtc
 ct gtgcggccgc ataccgtgt acagcccat gaagtaccac gggctcaacg 660ttcccggtc gcaccc
 cggc gtgcggggc tggccgggtt gggccacgtt gggctcaagt 720tcggcaaggc cttcgaaatg aaag
 tgcgg tgcgtact gtcggccgggg aagaaggagg 780aggccctggg gccgctggc gccgacgcgt tc
 atccgtcg caaggacgcc gacgagatga 840ag
 gctgtgat agcaccatgg atggcatcat aaacacggta tctgcaaaca tccccctgac 900ccctcttcc
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 t tcgctctagt tgccacgaat aagaccctgg ccggagcat 1020catcgccggc atgagcgaca cgcaggaa
 gat gctggaccc tcggcgacggc acggcgatgac 1080ggccgacatc gaggtggctcg cgcggagatg tggta
 acacg gccttggagc gccttggccaa 1140gaacgacgtc aggtatcgat tcgtcatcgacatcgcaac acc
 ctcgaca atggccggc 1200caccaccggag tgaacgtact cagcactgt tacatctac gttgttccac t
 gtagtgct 1260ccgtatgtaaaa caataaaacga tcaaaaactct tgcattggg tgcattggg tagacatggt
 1320tggggcgag gaaactgagt tgaaggatgg atggataaaa aaaaaaaaaa aaaaaaaaaa 1377
 <212> Type : DNA
 <211> Length : 1377

SequenceName : CAD2cvEllettcDNA (SEQ ID NO: 11)
 SequenceDescription :

Custom Codon

Sequence Name : CAD2cvEllettcDNA (SEQ ID NO: 11)

Sequence

<213> OrganismName : Lolium perenne
<400> PreSequenceString :
MAPTAAEQTE HHQHTRKAVG LAARDAGHL SPLAITRRST GDDDVVIKIL YCGICHSDLH 60ALKNDWKNS
R YPMIPGHEIA GEVTEVGKNV SKFKAGDRVG VGCVMNSCRS CESCDKGFE 120HCPGMILTYN SVDVDGT
VTY GGYSSMVVHV ERFVVRFPDA MPLDKGAPLL CAGITVYSPM 180KYHGLNVPGHL HGVLGLGGGL GHVAV
KFGKA FGMKVTVISS SPGKKEEALG RLGADAFIVS 240KDADEMKA VI APWMASXNTV SANIPLTPLF GLL
KPNKMI MVGLPEKPIE IPPFALVATN 300KTLAGSIIGG MSDTQEMLDL AAKHGVTA DI EVVGAEVYNT A
LERLAKNDV RYRFVIDIGN 360TLDNVAATTE
370
<212> Type : PRT
<211> Length : 370
SequenceName : CADcvEllettepep (SEQ ID NO: 12)
SequenceDescription :

Sequence

<213> OrganismName : Lolium perenne
<400> PreSequenceString :
gcagcggttn caaatcgccg gtcctggggt ggaagtgnag cagtggaaag atgtgtgcga 60gggggttg
t tttggatgna agacaggcgg gccagtggag aacaagagag aacgcgagag 120gccaaggat ccgcagc
ccc gcaaacaagg cctagattt ggttaagttt gggtcgctc 180agacaccgcg gcatccctt taggt
ggtcc gcgcgctgga ccgtatttt atcttagttt 240accatttcg acgcgcagac acgagatgga tgg
tgca gtcgtw agagatgacc taagtacaar 300aacctctccc cgagctccg ccatccgtca cttaccgagc g
acaaaagctt cccacttcat 360cacactcagc ccagcaagca tactgtatggt gagcgcactc gggctgtgc
ccaccgacc 420cacgcacatcc aaaaccaact ctactttca ccmcaccaac aaaagacaaa atatggtg
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5119

<212> Type : DNA
<211> Length : 5119
SequenceName : LpOmt1promoter (SEQ ID NO: 13)
SequenceDescription :

Custom Codon

Sequence Name : Lp0mt1promoter (SEQ ID NO: 13)

Sequence

<213> OrganismName : *Lolium perenne*

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 <211> Length : 4555
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 SequenceDescription :

Custom Codon

Common Name

Sequence Name : CAD2cvBarlanogenomic (SEQ ID NO: 14)

SequenceDescription :

Sequence

<213> Organi

<400> PreSequenceString :

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TVY SPMKYHGLNV PGLHLGVLGL GGLGHVAVKF GKAFGMKVTV 180ISSSPGKKEE ALGRIGADAF IVSKD
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312

<212> Type : PRT
<211> Length : 312
SequenceName : CAD2cvBarlanopep (SEQ ID NO: 15)
SequenceDescription :

Sequence

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<213> OrganismName : Lolium perenne
<400> PreSequenceString :
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<212> Type : DNA
<211> Length : 1378
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SequenceDescription :

Custom Codon

Sequence Name : CAD2cubBarlanoCDNA (SEQ ID NO: 16)

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<213> OrganismName : Lolium perenne
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SequenceDescription :

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Sequence Name : 4CL2promoterseq (SEQ ID NO: 17)

Sequence

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INTERNATIONAL SEARCH REPORT

International application No.

PCT/AU01/00699

A. CLASSIFICATION OF SUBJECT MATTERInt. Cl.⁷: A01H 5/00, C12N 9/00, C12N 9/02, C12N 9/04, C12N 9/10, C12N 15/29

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

GenBank, EMBL, PDB Nucleic Acids, GenPept, TREMBL, SWISS-PROT, PIR, Medline, ChemAbs, AGRICOLA, WPIDS

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	US 5 952 486 A (BLOKSBERG <i>et al</i>) 14 September 1999 whole of document Civardi L <i>et al</i> , "Molecular Cloning and Characterization of two cDNAs Encoding Enzymes Required for Secondary Cell Wall Biosynthesis in Maize", <i>NATO ASI Series, Volume H 104 (Cellular Integration of Signalling Pathways in Plant Development)</i> , 1998, pages 135-146 whole of document	1-26
X	WO 99/31243 A (INTERNATIONAL PAPER COMPANY) 24 June 1999 whole of document	1, 2, 4-12, 14-17, 20a, 19b, 23-26
X		1-3, 6-13, 16, 17, 19a, 19b, 21, 23-26

Further documents are listed in the continuation of Box C See patent family annex

* Special categories of cited documents:	"T"	later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
"A" document defining the general state of the art which is not considered to be of particular relevance	"X"	document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
"E" earlier application or patent but published on or after the international filing date	"Y"	document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	"&"	document member of the same patent family
"O" document referring to an oral disclosure, use, exhibition or other means		
"P" document published prior to the international filing date but later than the priority date claimed		

Date of the actual completion of the international search

23 July 2001

Date of mailing of the international search report

29 August 2001

Name and mailing address of the ISA/AU

AUSTRALIAN PATENT OFFICE
PO BOX 200, WODEN ACT 2606, AUSTRALIA
E-mail address: pct@ipaaustralia.gov.au
Facsimile No. (02) 6285 3929

Authorised officer

GARETH COOK

Telephone No : (02) 6283 2541

INTERNATIONAL SEARCH REPORT

International application No.
PCT/AU01/00699

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	GenBank accession AF052223, Heath RL <i>et al</i> , "Lolium perenne 4-coumarate-CoA ligase 4CL3 mRNA, complete cds", 7 March 2000.	1-3, 6-13, 16, 17, 19a, 19b, 21, 23-26
X	GenPept accession AAF37734, Heath RL <i>et al</i> , "4-coumarate-CoA ligase 4CL3 [Lolium perenne]", 7 March 2000.	1-3, 6-13, 16, 17, 19a, 19b, 21, 23-26
X	GenBank accession AF052222, Heath RL <i>et al</i> , "Lolium perenne 4-coumarate-CoA ligase 4CL2 mRNA, complete cds", 7 March 2000.	1-3, 6-13, 16, 17, 19a, 19b, 21, 23-26
X	GenPept accession AAF37733, Heath RL <i>et al</i> , "4-coumarate-CoA ligase 4CL2 [Lolium perenne]", 7 March 2000.	1-3, 6-13, 16, 17, 19a, 19b, 21, 23-26
X	GenBank accession AF052221, Heath RL <i>et al</i> , "Lolium perenne 4-coumarate-CoA ligase 4CL1 mRNA, complete cds", 7 March 2000.	1-3, 6-13, 16, 17, 19a, 19b, 21, 23-26
X	GenPept accession AAF37732, Heath RL <i>et al</i> , "4-coumarate-CoA ligase 4CL1 [Lolium perenne]", 7 March 2000.	1-3, 6-13, 16, 17, 19a, 19b, 21, 23-26
X	WO 98/39454 A (ZENECA LIMITED) 11 September 1998 whole of document	1, 2, 4, 6-12, 14, 17, 20a, 19b, 22-26
X	Pichon M <i>et al</i> , "Cloning and characterization of two maize cDNAs encoding Cinnamoyl-CoA Reductase (CCR) and differential expression of the corresponding genes", <i>Plant Molecular Biology</i> , 1998, 38:671-676 whole of document	1, 2, 4, 6-12, 14, 17, 20a, 19b, 22-26
X	GenBank accession AJ231134, Selman-Housein G <i>et al</i> , "Saccharum officinarum mRNA for cinnamoyl-CoA reductase" 25 January 2000	1, 2, 4, 6-12, 14, 17, 20a, 19b, 22-26
X	WO 93/05159 A (IMPERIAL CHEMICAL INDUSTRIES PLC) 18 March 1993 whole of document	1, 2, 5-12, 15-17, 19b, 23-26
X	WO 93/24638 A (ZENECA LIMITED) 9 December 1993 whole of document	1, 2, 5-12, 15-17, 19b, 23-26
X	Baucher M <i>et al</i> , "Down-regulation of cinnamyl alcohol dehydrogenase in transgenic alfalfa (<i>Medicago sativa</i> L.) and the effect on lignin composition and digestibility", <i>Plant Molecular Biology</i> , 1999, 39:437-447 whole of document	1, 2, 5-12, 15-17, 19b, 23-26
X	GenBank accession AF010290, McAlister FM <i>et al</i> , "Lolium perenne cinnamyl alcohol dehydrogenase mRNA, complete cds", 23 September 1997	1, 2, 5-12, 15-17, 19b, 23-26
X	GenPept accession AAB70908, McAlister FM, "cinnamyl alcohol dehydrogenase [Lolium perenne]", 22 September 1997	1, 2, 5-12, 15-17, 19b, 23-26

INTERNATIONAL SEARCH REPORT

International application No. PCT/AU01/00699

C (Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	Heath R <i>et al</i> , "cDNA Cloning and Differential Expression of Three Caffeic Acid O-Methyltransferase Homologues from Perennial Ryegrass (<i>Lolium perenne</i>)," <i>Journal of Plant Physiology</i> , 1998, 153:649-657 whole of document	17, 18, 19b, 20b, 23-26
X	GenBank accession AF033540, Heath RL <i>et al</i> , "Lolium perenne caffeic acid O-methyltransferase (OMT3) mRNA, complete cds", 29 January 1999	17, 18, 19b, 20b, 23-26
X	GenBank accession AF033539, Heath RL <i>et al</i> , "Lolium perenne caffeic acid O-methyltransferase (OMT2) mRNA, complete cds", 29 January 1999	17, 18, 19b, 20b, 23-26
X	GenBank accession AF033548, Heath RL <i>et al</i> , "Lolium perenne caffeic acid O-methyltransferase (OMT1) mRNA, complete cds", 29 January 1999	17, 18, 19b, 20b, 23-26
X	GenBank accession AF010291, McAlister FM <i>et al</i> , "Lolium perenne bispecific caffeic acid/hydroxyferulic acid O-methyltransferase mRNA, complete cds", 3 June 1998	17, 18, 19b, 20b, 23-26
X	Capellades M <i>et al</i> , "The maize caffeic acid O-methyltransferase gene promoter is active in transgenic tobacco and maize plants," <i>Plant Molecular Biology</i> , 1996, 31:307-322 whole of document	17, 18, 19b, 20b, 23-26
	Note that the application as filed contains two claims numbered 19 and two claims numbered 20. To distinguish them, 19a and 20a refer to the first claims 19 and 20, and 19b and 20b refer to the second claims 19 and 20.	

Supplemental Box

(To be used when the space in any of Boxes I to VIII is not sufficient)

Continuation of Box No: II, Observations where unity of invention is lacking

The international search report has been drawn up in respect of the entire international application but the International Searching Authority is of the opinion that the application does not appear to comply with the requirements of unity of invention as set forth in the PCT regulations (Article 34(3), Rule 68(1) PCT).

The separate groups of invention are:

1. Claims 1, 2, 6 to 12, 16, 17, 19a, 19b and 23 to 26 (partial) and claims 3, 13 and 21 (complete) are to 4-coumarate-CoA ligase (4CL) from ryegrass (*Lolium*), the nucleotide sequence encoding it, the promoter from its gene and various uses of them. 4CL from ryegrass is considered to be the first "special technical feature".
2. Claims 1, 6 to 11, 16, 17, 19a, 19b and 23 to 26 (partial) are to 4CL from fescue (*Festuca*), the nucleotide sequence encoding it, the promoter from its gene and various uses of them. 4CL from fescue is considered to be the second "special technical feature".
3. Claims 1, 2, 6 to 12, 16, 17, 20a, 19b and 23 to 26 (partial) and claims 4, 14 and 22 (complete) are to cinnamoyl-CoA reductase (CCR) from ryegrass, the nucleotide sequence encoding it, the promoter from its gene and various uses of them. CCR from ryegrass is considered to be the third "special technical feature".
4. Claims 1, 6 to 11, 16, 17, 19b, 20a and 23 to 26 (partial) are to CCR from fescue, the nucleotide sequence encoding it, the promoter from its gene and various uses of them. CCR from fescue is considered to be the fourth "special technical feature".
5. Claims 1, 2, 6 to 12, 16, 17, 19b and 23 to 26 (partial) and claims 5 and 15 (complete) are to cinnamyl alcohol dehydrogenase (CAD) from ryegrass, the nucleotide sequence encoding it, the promoter from its gene and various uses of them. CAD from ryegrass is considered to be the fifth "special technical feature".
6. Claims 1, 6 to 11, 16, 17, 19b, and 23 to 26 (partial) are to CAD from fescue, the nucleotide sequence encoding it, the promoter from its gene and various uses of them. CAD from fescue is considered to be the sixth "special technical feature".
7. Claims 17, 18, 19b and 23 to 26 (partial) and claim 20b (complete) are to caffeic acid O-methyltransferase (OMT) gene promoter from ryegrass and various uses of it. Caffeic acid OMT from ryegrass is considered to be the seventh "special technical feature".
8. Claims 17, 18, 19b and 23 to 26 (partial) are to caffeic acid O-methyltransferase (OMT) gene promoter from fescue and various uses of it. Caffeic acid OMT from fescue is considered to be the eighth "special technical feature".

In order for there to be unity between the four types of enzymes claimed, they have to share a significant structural element, that is a structural element that defines the specific biological activity of the enzymes, and the significant structural element must be disclosed in the specification. No significant structural element has been identified as being shared by the four types of enzymes, hence there is lack of unity between the enzymes. In addition, all four types of enzymes are known in the prior art, for example, in US 5 952 486. Hence unity of invention is also lacking between the enzymes from ryegrass and the enzymes from fescue.

Note that the application as filed contains two claims numbered 19 and two claims numbered 20. To distinguish them, 19a and 20a refer to the first claims 19 and 20, and 19b and 20b refer to the second claims 19 and 20.

INTERNATIONAL SEARCH REPORT
Information on patent family-members

International application No.
PCT/AU01/00699

This Annex lists the known "A" publication level patent family members relating to the patent documents cited in the above-mentioned international search report. The Australian Patent Office is in no way liable for these particulars which are merely given for the purpose of information.

Patent Document Cited in Search Report				Patent Family Member			
US	5 952 486	AU	44036/97	BR	9711745	EP	929 682
		US	5 850 020	WO	98/11205	ZA	9710451
		ZA	9810574	US	6 204 434		
WO	99/31243	US	6 252 135	ZA	9811568		
WO	98/39454	AU	63041/98	EP	970 222		
WO	93/05159	AU	16581/92	BR	9205934	CA	2 109 222
		EP	584 117	US	5 451 514	US	6 066 780
WO	93/24638	AU	43345/93	BR	9306445	EP	643 774
		US	5 633 439				
END OF ANNEX							